

Author Accepted Manuscript (AAM)

Adenosine–Dopamine interactions in the pathophysiology and treatment of CNS disorders

K. Fuxe; D. Marcellino; D.O. Borroto-Escuela; M. Guescini; V. Fernández-Dueñas; S. Tanganelli; A. Rivera; F. Ciruela; L.F. Agnati

Published article: Fuxe K, Marcellino D, Borroto-Escuela DO, et al. Adenosine–Dopamine Interactions in the Pathophysiology and Treatment of CNS Disorders. *CNS Neuroscience & Therapeutics*. 2010;16(3):e18–e42. doi:10.1111/j.1755-5949.2009.00126.x

This document is the peer-reviewed author accepted manuscript. The final Version of Record is available at the publisher via the DOI link above.

Repository deposit note: please cite the published version when referencing this work.

Adenosine-Dopamine interactions in the pathophysiology and treatment of CNS disorders

K. Fuxe¹, D. Marcellino¹, D.O. Borroto-Escuela¹, M. Guescini², V. Fernández-Dueñas³, S.Tanganelli⁴, A. Rivera⁵, F. Ciruela³ and L.F. Agnati⁶

¹*Department of Neuroscience, Karolinska Institutet, Stockholm, Sweden,* ²*Department of Biomolecular Sciences, University of Urbino 'Carlo Bo', Italy,* ³*Unitat de Farmacologia, Departament Patologia i Terapèutica Experimental, Facultat de Medicina, Universitat de Barcelona, L'Hospitalet de Llobregat, Barcelona, Spain,* ⁴*Department of Clinical and Experimental Medicine, Section of Pharmacology, University of Ferrara, Ferrara, Italy* ⁵*Department of Cell Biology, University of Malaga, Malaga, Spain,* ⁶*Department of Biomedical Sciences, University of Modena, Modena, and IRCCS Lido, Venice, Italy*

Adenosine-dopamine interactions in the CNS have been studied for many years in view of their relevance for disorders of the central nervous system and their treatments. The discovery of adenosine and dopamine receptor containing receptor mosaics (higher-order receptor heteromers) in the striatum opened up a new understanding of these interactions. Initial findings indicated the existence of A_{2A}R-D₂R heterodimers and A₁R-D₁R heterodimers in the striatum that were followed by indications for the existence of striatal A_{2A}R-D₃R and A_{2A}R-D₄R heterodimers. Of particular interest was the demonstration that antagonistic allosteric A_{2A}-D₂ and A₁-D₁ receptor-receptor interactions take place in striatal A_{2A}R-D₂R and A₁R-D₁R heteromers. As a consequence, additional characterization of these heterodimers led to new aspects on the pathophysiology of Parkinson's disease, schizophrenia, drug addiction and L-DOPA-induced dyskinesias relevant for their treatments. In fact, A_{2A}R antagonists were introduced in the symptomatic treatment of Parkinson's disease in view of the discovery of the antagonistic A_{2A}R-D₂R interaction in the dorsal striatum that leads to reduced D₂R recognition and

$G_{i/o}$ coupling in striato-pallidal GABAergic neurons. In recent years, indications have been obtained that $A_{2A}R$ - D_2R and A_1R - D_1R heteromers do not exist as heterodimers, rather as receptor mosaics. In fact, A_{2A} - CB_1 - D_2 receptor mosaics and A_{2A} - D_2 - $mGlu_5$ receptor mosaics have been discovered using a sequential BRET-FRET technique and by using the BRET technique in combination with bimolecular fluorescence complementation. Thus, other pathogenic mechanisms beside the well-known alterations in the release and/or decoding of dopamine in the basal ganglia and limbic system are involved in Parkinson's disease, schizophrenia and drug addiction. In fact, alterations in the stoichiometry and/or topology of A_{2A} - CB_1 - D_2 and A_{2A} - D_2 - $mGlu_5$ receptor mosaics may play a role. Thus, the integrative receptor-receptor interactions in these receptor mosaics give novel aspects on the pathophysiology and treatment strategies, based on combined treatments, for Parkinson's disease, schizophrenia and drug addiction.

1. Introduction

1.1 Role of dopamine and adenosine as volume transmission signals in the CNS

The first observation that led to the discovery of a mode of communication different from synaptic transmission was the appearance of extra-neuronal dopamine (DA) fluorescence around midbrain DA nerve cells following amphetamine, a catecholamine (CA) releasing drug, treatment in reserpine-nialamide-L-DOPA treated rats [1]. Locally applied CA into the striatum was also found to migrate to the neuropil [2, 3]. Such observations taken together with the existence of global monoamine terminal networks in the central nervous system (CNS, see above) and with several other observations in the literature, in particular the demonstration of large numbers of nonjunctional monoamine varicosities by Descarries [4], led Agnati and Fuxe to propose the existence of volume transmission (VT) as complementary transmission to the well-known wiring transmission (WT), or synaptic transmission, in the CNS [5]. VT in the CNS was introduced as an

extracellular fluid (ECF) and cerebrospinal fluid (CSF) form of transmission [5-11]. In this way, VT signals are chemical signals like neurotransmitters, trophic factors, ions, peptides, etc. that migrate by diffusion and convection from the source cells to the target cells in the ECF and CSF in VT channels of the extracellular space and the ventricles as a consequence of energy gradients that create migration.

Evidence exists and suggests that the main mode of communication of all the central DA neurons is short distance VT in the μm range in which DA primarily reaches extrasynaptic receptors; see [10-14]. Thus, DA via VT in the local circuits of the medium-sized striatal neurons reaches and activates extrasynaptic D₁R- and D₂R-containing Receptor Mosaics (RM) (see below) on the dendritic spines of such neurons [15]. In many regions, DA exists as a diffusing VT signal in the ECF in concentrations that vary with the pattern of DA release and has a major impact on the modulation of the polymorphic wiring networks in the CNS [16, 17]. In this way it becomes possible to understand how the DA terminal networks have such a powerful role in CNS functions involving mood, reward, fear, cognition, attention, arousal, motor function, neuroendocrine and autonomic function and indeed play a central role in neuropsychopharmacology.

Adenosine is an endogenous nucleoside and functions as a neuromodulator in many areas of the central nervous system (CNS), see [18-20]. It is a normal cellular constituent and its intracellular concentration is dependent on the breakdown and synthesis of ATP, which is metabolized to AMP. Adenosine is then formed from AMP, through the action of a 5'-nucleotidase, and the intracellular and extracellular concentrations are kept in equilibrium by means of equilibrative transporters. The two main metabolic pathways of adenosine removal depend on the enzymes adenosine deaminase (ADA) (mostly intracellular) and adenosine kinase. Extracellular adenosine concentration depends on intracellular adenosine and also on extracellular ATP (released as a neurotransmitter or as an intracellular signal, from neurons or glial cells) that is rapidly hydrolyzed to adenosine and other metabolites. However, the main source of extracellular adenosine is likely intracellular adenosine released from active cells in response to an increased metabolic demand [19, 20]. Based on its presence in and its release into the

ECF, together with the demonstration of extrasynaptic adenosine receptors, adenosine likely represents an important VT signal [18].

The two major adenosine receptors in the CNS are the adenosine A₁ and A_{2A} receptors. A₁R are widely distributed in the brain and are mainly expressed in the hippocampus, cerebellum and neocortical areas. On the other hand, A_{2A}R have a much more restricted brain distribution, in which the striatum contains the highest density in the brain and where they are specially concentrated in the GABAergic striato-pallidal neurons [11, 17, 18] together with the D₂R. The D₁R are instead predominantly found in the direct pathway, the striato-entopeduncular and striato-nigral GABAergic neurons, see [14, 15]).

It has been many years since the hypothesis was first introduced that adenosine-dopamine interactions in the brain primarily take place via receptor-receptor interactions in A_{2A}R-D₂R and A₁R-D₁R heteromers located perisynaptically at glutamate synapses on the striato-pallidal and striato-entopeduncular/nigral GABAergic neurons, respectively, see [21-25]. However, although it may be a rather infrequent event, it is possible that adenosine, via VT into DA synapses may directly modulate synaptic DA transmission via synaptic A_{2A}R-D₂ and A₁R-D₁R heteromers.

1.2. The concept of receptor mosaic and its implication: stoichiometry versus topology

Already in 1980 it was proposed by Fuxe and Agnati that assemblages of receptors could operate as integrative input units of membrane associated molecular circuits [26, 27]. This postulation was supported by indirect evidence on the existence of receptor-receptor interactions obtained through an analysis of the effects of neuropeptides on the binding characteristics of monoamine receptors in membrane preparations from discrete brain regions [28-30]. As a logical consequence for the indications of direct physical interactions between neuropeptide and monoamine receptors, the well-known terms heteromerization versus homomerization were introduced by the Agnati

and Fuxe teams as well as by other groups to describe this kind of interaction between different types of GPCRs, see [21, 23, 31-37].

Allosteric events were postulated to be the molecular mechanism for intra-membrane interactions in multimeric assemblages of receptors. Thus, the term receptor mosaic (RM) [8, 35, 38, 39] was introduced for assemblies of multiple receptors of the same or different kinds ($n \geq 3$) in the plasma membrane as a more meaningful term than higher-order heteromers, which nevertheless is highly relevant from a stoichiometric point of view. The term receptor mosaic indicates the “integrated output” of such an input unit, since it also stresses the concept that topology (spatial localization in the plane of the membrane) and integrative function of the receptor assemblage are deeply interconnected. In other words, the emergent properties of the receptor assemblage, or its integrated output, depend on the location and the order of activation of the participating receptors as well as on the type of allosteric interactions (entropic and/or enthalpic) within such an integrative receptor mosaic [16, 17]. Already in the 1982 [39] it was proposed that formation of a RM and/or its allosteric change could have a role in the molecular basis for the engram by leading to a transient and/or permanent change of the synaptic efficacy (i.e., the synaptic weight). The term RM maintains that allostery is any ligand-induced change in protein conformation and/or dynamics but also includes the functional characteristic of allostery, namely that one ligand alters the functional response of another ligand through a conformational change in the binding site of the second ligand, see [16, 17, 40-42]. It is then possible that a GPCR can have very different biochemical properties leading to a different pharmacology through interactions with another GPCR [15-17, 21, 35, 43-45].

The field of receptor-receptor interactions has opened up new targets for drug development [15, 17, 21, 44] and several strategies can be exploited to develop new drugs based on receptor-receptor interactions in receptor heteromers, see [46]. Higher-order receptor heteromers (receptor mosaics) also offer several additional targets for drug development. Novel drugs may be developed to modify the composition of RMs, their topography, the order of activation as well as allosteric regulators modulating the functional state of the individual receptors in the RM. Drugs may affect, e.g. (I) the synthesis and

release of receptor oligomeric building blocks from the endoplasmic reticulum, (II) the insertion of such building blocks into the plasma membrane, (III) the internalization of RMs, (IV) the adapter and scaffolding proteins organizing the RMs and (V) ligand induced receptor assembly.

The potential importance of developing allosteric modulators has also been suggested since they may, *inter alia*, substantially affect the allosteric mechanisms within the RM leading to changes in its integrative function [47-49] in addition to affects on RM assemblage, G protein, β -arrestin coupling and receptor recognition [41, 50]. An example is the discovery of an allosteric D₂R antagonist homocysteine (Hcy), which reduces D₂R agonist binding and D₂R function by its apparent binding to the third intracellular loop (IC3) of the D₂R [47, 48]. This discovery opens up the development of new antipsychotic drugs based on the development of allosteric Hcy agonist analogues and antiparkinsonian drugs based on the development of allosteric Hcy antagonist analogues.

In the present review we will focus on the A_{2A}R-D₂R heteromers and A_{2A}R-D₂R-containing RM as well as the A₁R-D₁R heteromers and the A₁R-D₁R-containing RM as targets for the development of novel treatments of CNS diseases.

2. A_{2A}-D₂-like receptor heteromers and A_{2A}R-D₂R containing receptor mosaics
 - 2.1. A_{2A}R-D₂R heteromers
 - 2.1.1. Biochemical and functional findings

Initially, to tackle the study of receptor heteromers traditional biochemical protocols were used and some of those methods include microscopy-based procedures, such as co-immunolocalization, and immobilized protein-protein interaction assays, such as co-immunoprecipitation. The invasive nature of these technical approaches to study protein-protein interactions still have the disadvantage of altering the natural state of the cell and therefore, may not represent its real structure. This is even more critical with membrane proteins, like GPCRs, due to their highly hydrophobic framework and the need for detergents to extract proteins

from membranes. Either working with aqueous solutions or with detergents, the composition and organization of the membrane is altered, which can be a source of false results. Nevertheless, besides the inherent technical problems associated with these methods they have been shown to give accurate results, and they are still very useful to confirm close interactions between GPCRs. During the last decade, a new set of technologies based on the use of fluorescent-fused proteins have been developed to overcome the invasive nature of the immobilized protein-protein interaction assays. These new approaches, centered on the use of various adaptations of resonance energy transfer (RET) techniques (e.g. fluorescence-RET and bioluminescence-RET), have favored the possibility of carrying out “*in vivo*” real time experiments. Thus, the use of BRET and FRET techniques has emerged as powerful tools to study GPCR oligomerization. A_{2A}R-D₂R heteromers [51-54] may exist in the dorsal and ventral striato-pallidal GABA pathway in which activation of A_{2A}R reduce D₂R recognition, coupling, and signaling together with A_{2A}R and D₂R homodimers (Fig. 1) [21, 23, 25, 51-55]. A large number of studies using the above mentioned approaches (e.g. coimmunoprecipitation, FRET and BRET), as well as biochemical binding and signaling, behavioral pharmacology, and microdialysis techniques, have corroborated the existence of A_{2A}R-D₂R heteromers [24, 51-53, 55-61]. Interestingly, it has been also suggested that A_{2A}R-D₂R heteromers may be predominantly located on the dendritic spines in the perisynaptic zones of DA terminals and glutamate synapses but also on glutamate terminals in the local circuits of the striato-pallidal GABAergic neurons [24, 55, 57, 60, 62, 63].

In the striato-pallidal GABAergic neuron, this heteromer may exist in equilibrium on the neuronal surface membrane together with A_{2A}R and D₂R homomers. It seems possible that higher-order A_{2A}-D₂ RM of unknown stoichiometry and topology may also exist and contain e.g., D₂R homodimers and A_{2A}R homodimers. In such a case, antagonistic A_{2A}R-D₂R interactions can still take place by assuming that the A_{2A}R can enhance the negative cooperativity in such participating D₂R homodimers. Such events may also take place in the A_{2A}R-D₃R and A_{2A}R-D₄R heteromers (see below). A major component of the interface in the A_{2A}R-D₂R heteromer is the electrostatic interaction between the positively charged arginine-rich epitope in the N-

terminal domain of the IC3 of the D₂R and negatively charged epitopes in the C-terminal tail of the A_{2A}R, especially the epitope (aa 370-378) containing a phosphorylatable serine (Fig. 2) [52, 64]. Thus, phosphorylation events may modulate the strength of the receptor-receptor interactions within the A_{2A}R-D₂R heteromer and RM. These results were also supported by studies using D₁R-D₂R chimeras [65]. In addition, microdialysis experiments indicate that in awake, freely moving rats the A_{2A}R agonist CGS 21680 when intrastrially co-perfused with the D₂R-D₃R agonist quinpirole (10 μM), was able to fully counteract the quinpirole-induced reduction of extracellular GABA levels in the globus pallidus, see [61], where CGS 21680 itself did not produce any significant effects on its own.

The antagonistic A_{2A}R-D₂R interaction in the brain has been demonstrated in many publications including at the level of D₂R agonist recognition and animal behavior (see [23, 24, 55, 60, 63, 66-69]. These results make it likely that the A_{2A}R-D₂R heteromer strongly modulates the excitability in the striato-pallidal GABAergic neurons via its ability to counteract D₂R signaling to multiple effectors. The A_{2A}R-induced counteraction of the D₂R-induced inhibition of the Ca²⁺ influx over the L-type voltage dependent Ca²⁺ channels (Cav 3.1 channels) via the activation of phospholipase C and protein phosphatase-2B (calcineurin) [70] may be of special importance [15, 60]. The G protein involved may be G_{i/o} and/or G_{q/11} with the release of the βγ subunits. The counteraction of this cascade by A_{2A}R leads to increased phosphorylation of this calcium channel and its increased opening favoring an upstate of the striato-pallidal GABAergic neuron, see [71]. A_{2A}R activation also has been shown to counteract D₂R-induced intracellular calcium responses in cotransfected mouse fibroblast and human neuroblastoma cell lines [72, 73]. Moreover, the D₂R agonist-induced reduction of firing rates in the DA denervated striatum was enhanced by A_{2A}R antagonists and attenuated by A_{2A}R agonists [69]. The D₂R in the A_{2A}R-D₂R heteromers may also be coupled to G_{i/o} since in cultured striatal neurons A_{2A}R agonists can counteract the D₂R induced inhibition of forskolin stimulated cyclic adenosine monophosphate (cAMP) production without being active when given alone [15, 24, 53, 55, 60]. It is also likely that A_{2A}R activation through inhibition of the G_{i/o} coupling of the D₂R with the G_{i/o} trimer remaining

at the D₂R will also interfere with the protein kinase B (Akt)-Glycogen synthase kinase 3 (GSK-3) signaling cascade induced by D₂R stimulation through its β-arrestin 2 signaling [74]. Thus, β-arrestin 2 can no longer become effectively linked to the D₂R, since the G_{i/o} trimer is not sufficiently split and removed from the D₂R in the presence of A_{2A}R activation (Fig.3). It should also be considered that the A_{2A}R-D₂R receptor-receptor interaction also leads to a conformational state of the D₂R less able to bind and activate the β-arrestin 2 as is the case for G_{i/o}.

There also exists a reciprocal interaction between A_{2A}R-D₂R in as much as D₂R can inhibit the A_{2A}R-induced increase in cAMP accumulation via G_{i/o} at the level of the adenylate cyclase (AC), an interaction which also can take place between A_{2A} and D₂ homomers, see [24, 55, 75]. Removal of the D₂R brake on the A_{2A}R signaling would therefore, also lead to increased striatal excitability since it will result in increased protein kinase A (PKA) activity causing increased phosphorylation of α-amino-3-hydroxy-5-methyl-4-isoxazole-propionate (AMPA) and N-methyl-D-aspartic acid receptors (NMDA) and of dopamine and cAMP regulated neuronal phosphoprotein (DARPP-32) at Thr34 position with inhibition of protein phosphatase-1 further enhancing the phosphorylation and the activity of these ion channel receptors, see [60, 76-80]. Such events would also favor the up-state of the striato-pallidal GABAergic neurons. Based on the above, it seems likely that the A_{2A}R-D₂R heteromer, via its antagonistic receptor-receptor interaction, plays a crucial role in moving the striato-pallidal GABAergic neurons towards the up-state by the antagonism of D₂R signaling, see [15].

2.1.2. Relevance to Parkinson's disease and its treatment

Based on the antagonistic A_{2A}R-D₂R interaction described above, the development of A_{2A}R antagonists to target these A_{2A}R-D₂R heteromers in the dorsal striato-pallidal GABA pathway was initiated [24, 59, 63, 81, 82]. It was also demonstrated that the antagonistic A_{2A}R-D₂R interaction remained and was even increased in the striatal membranes from rat models of Parkinson's disease (PD) [68, 81]. In a number of PD models, A_{2A}R antagonists, like SCH 58261, have been found to dose-dependently increase locomotor activity in

combination with sub-threshold doses of L-DOPA and D₂R agonists in reserpinized mice. In fact, A_{2A}R antagonists consistently reverse Parkinsonian deficits in non-human primates and rodents; see [60, 61, 68, 81]. It should be noted that in contrast to caffeine [83], subchronic A_{2A}R antagonist treatment in models of Parkinson's disease does not result in tolerance development [84, 85], which represents a prerequisite for clinical development. Therefore, similar antiparkinsonian effects may be seen with acute and chronic treatment of A_{2A}R antagonists.

Such results could elegantly be explained by the hypothesis that A_{2A}R antagonists target the A_{2A}R-D₂R heteromer and increase D₂R signaling in these heteromers. These A_{2A}R-D₂R heteromers have been found to be constitutive [51] and thus exist in the absence of agonist activation of the receptors. Therefore, it is not likely that A_{2A}R antagonist treatment will disrupt the A_{2A}R-D₂R heteromers but conformational changes may develop in the heteromer thereby altering its integrative activity. So far it has not been possible to see differences in the affinity and blocking activity of A_{2A}R antagonists at A_{2A}R belonging to A_{2A}R-D₂R heteromers or to A_{2A}R homomers (unpublished data). The A_{2A}R antagonist istradefylline (KW-6002) has been used in clinical trials and found to have interesting anti-Parkinsonian and anti-dyskinetic properties [60, 68, 78, 86]. Initial clinical studies using KW-6002 showed symptomatic but rather modest improvement in relatively advanced PD patients with dyskinetic complications [87, 88]. However, bradykinesia, muscle rigidity and resting tremor in PD patients may be improved after A_{2A}R blockade.

Based on our hypothesis, treatment with an A_{2A}R antagonist alone should in fact produce only modest effects in PD except during early stages of PD when DA is still being released from remaining DA terminals. Instead the A_{2A}R antagonist should ideally be given in Parkinsonian patients in combination with close to threshold doses of L-DOPA and/or D₂R agonist. Clinical results observed so far would be in agreement with this view. Thus, the A_{2A}R antagonist would be targeting the A_{2A}R-D₂R heteromer to enhance D₂R signaling.

A_{2A}R antagonists, by acting on the A_{2A}R-D₂R heteromer will enhance D₂R signaling at the soma-dendritic level and lead to a reduction in the activity

of the striato-pallidal GABA/enkephalin pathway. In this way, motor inhibition will be reduced and the motor drive will be partly restored. A combined treatment with L-DOPA will still be optimal since it will restore D₁R activity in the direct pathway which helps motor initiation. The direct pathway becomes integrated with the indirect pathway in the globus pallidus interna and zona reticulata of the substantia nigra to optimally inhibit their GABA projections to the motor thalamus. In this way, the GABA inhibition of the excitatory glutamate thalamo-cortical pathway to the motor cortices will be removed and movements restored

To understand the role of A_{2A}R antagonists in PD it becomes important to note that there also exists a reciprocal interaction by which D₂R inhibits the A_{2A}R signaling at the level of AC as discussed above, either in the A_{2A}R-D₂R heteromer or between A_{2A}R homomers and D₂R homomers (see above). In this way, we can understand why A_{2A}R antagonists alone can counteract haloperidol-induced catalepsy: namely, by blocking the excessive A_{2A}R signaling that results from the removal of D₂R-induced inhibition of AC by haloperidol. This mechanism can also help to explain some therapeutic effects of A_{2A}R antagonists in advanced PD, see [60].

When discussing the role of A_{2A}R and D₂R we should also consider the balance between A_{2A}R homomers versus D₂R homomers and A_{2A}R-D₂R heteromers (Fig.1). L-DOPA treatment may lead to a disruption of the balance between them that ultimately leads to increases in A_{2A}R signaling versus D₂R signaling. This would help to explain the reduction of the therapeutic effects of L-DOPA and the appearance of L-DOPA-induced dyskinesias after prolonged treatment. Our hypothesis is that a L-DOPA-induced co-internalization of A_{2A}R-D₂R heteromers and D₂R homomers leads to a compensatory up-regulation of A_{2A}R and an increase in A_{2A}R homomers. To support this hypothesis, increases of A_{2A}R mRNA and A_{2A}R immunoreactivity (IR) have also been demonstrated in both animal models of L-DOPA-induced dyskinesias and in dyskinetic PD patients, see [89]. The resulting up-regulation of A_{2A}R leads to increases in PKA and phosphorylated DARPP-32 at Thr34 and increased inhibition of PP-1. This will result in an increase in protein phosphorylation including ion channels, which may help stabilize pathological RM formed under the influence of the transcriptional panorama

caused by the L-DOPA-induced excessive D₂R activation [52, 64]. This may lead to a repeated appearance of the abnormal pattern of firing in the striato-pallidal GABA pathway and contribute to dyskinesias. This hypothesis can help explain the reported anti-dyskinetic effect of A_{2A}R antagonists, which cannot be explained by the enhancement of D₂R signaling in the heteromer since this would worsen dyskinesias. We have also postulated, based on this hypothesis, that A_{2A}R antagonists will help to counteract the disappearance of the therapeutic effects of L-DOPA after long-term treatment, see [60, 68].

Taken together, the demonstrated anti-Parkinsonian effect of A_{2A}R antagonists in clinical studies has given 'proof of concept' that intramembrane receptor-receptor interactions in receptor heteromers can lead to the development of novel therapies. The A_{2A}R in the A_{2A}R-D₂R heteromer in the dorsal striato-pallidal GABAergic neurons is the major target for A_{2A}R antagonists when used in treatment of PD.

A_{2A}R antagonists may have not only anti-Parkinsonian properties but also neuroprotective and anti-dyskinetic properties. Recently it has been found that inactivation of forebrain A_{2A}R fully counteracts nigral DA nerve cell degeneration in mouse model of Parkinson's disease associated with a block also of nigral gliosis [90]. In line with these results, an attenuation of the DA nerve cell degeneration and of the gliosis was observed after treatment with the A_{2A}R antagonist SCH58261 [90]. However, the molecular mechanism for its neuroprotective effect on DA cells, as found by Schwartzschild *et al.*, see [60, 68], is still unclear, although an increase in retrograde trophic signaling from the striatum has been proposed by Fuxe *et al.* [60].

When discussing the potential neuroprotective effects of A_{2A}R antagonists in Parkinson's disease the epidemiological evidence should also be considered. There exists an inverse association between intake of coffee and caffeine in Japanese-American men and the risk of development of Parkinson's disease [91]. These results were strengthened by a study in a larger and more ethnically diverse cohort of prospectively followed men showing again an inverse relationship between consumption of caffeinated but not decaffeinated coffee and the risk of development of Parkinson's disease [92]. This link between consumption of caffeine in coffee and lower incidence of Parkinson's disease strongly favor the view that A_{2A}R antagonists

may have neuroprotective actions in Parkinson's disease. Thus, it is generally believed that caffeine exerts its psychoactive actions through blockade of A₁R and A_{2A}R in the CNS [20, 83]. Overall, A_{2A}R antagonists therefore clearly offer a realistic opportunity to improve PD treatment.

2.1.3. Relevance to schizophrenia and its treatment

The indicated existence of A_{2A}R-D₂R heteromers with antagonistic A_{2A}R-D₂R interactions in the ventral striato-pallidal GABA pathway has also introduced the strategy of using A_{2A}R agonists for the treatment of schizophrenia. In this way, based on these antagonistic interactions, A_{2A}R agonist treatment would lead to a reduction in the high affinity state of the D₂R and a reduction of its G_{i/o} coupling [24, 25, 55, 93]. It is also known that the common feature of antischizophrenic drugs of the D₂R antagonist type is the antagonism of the D₂R-β-arrestin 2 interaction and thus of Akt-GSK-3 signaling [94, 95]. As discussed, the observations suggest that A_{2A}R agonists may also interfere with the D₂R-β-arrestin 2 interaction (Fig. 3). Also it has been shown that the A_{2A}R agonist CGS-21680 reduces protein phosphatase 2A activity in the murine heart [96] which increases Akt activation and thereby blocks the GSK-3 signaling (Fig. 3).

The classical treatment in schizophrenia is the use of DA receptor antagonists such as haloperidol (a typical antipsychotic) to block the D₂R on both nigro-striatal and mesolimbic DA neurons [5, 97-101]. This leads to motor and mental effects through the blockade of excessive D₂R-mediated DA transmission in the nigro-striatal and meso-limbic DA systems, respectively; see [25, 55, 93]. Atypical antipsychotic drugs, like benzamides, that display reduced motor side effects may at least in the case of remoxipride block only subpopulations of D₂R [102]. The reason for this may be that remoxipride-sensitive D₂R receptors are part of a particular receptor heteromer [15, 17, 24, 45] that provide a unique pharmacology to these D₂R and make them remoxipride-sensitive. According to the Seeman hypothesis of schizophrenia [101], the major error in psychosis is an increased proportion of D₂R in the high affinity state that results in the development of D₂R supersensitivity. This makes our proposal on the antipsychotic potential of A_{2A}R agonists of special interest since they can, via the A_{2A}R-D₂R heteromer,

preferentially reduce the high affinity agonist state of the D₂R in both the dorsal and ventral striatum.

However, the current combined glutamate/DA hypothesis of schizophrenia states that the meso-limbic DA neurons are hyperactive due to reduced NMDA receptor function of cortical glutamate systems. This results in the reduced activity of the descending cortical glutamate projections to the ventral tegmental area (VTA) giving rise to the meso-cortical and meso-limbic DA systems. In this way, the VTA GABA interneurons that inhibit the firing of the meso-limbic DA neurons have reduced activity and as a consequence the meso-limbic DA neurons become hyperactive and inhibition of the ventral striato-pallidal GABA pathway becomes increased [15, 45, 103]. The ventral striato-pallidal GABAergic neurons integrate and transfer the emotional information from the limbic system via the medial dorsal (MD) nucleus to the prefrontal cortex (PFC) [15]. The increased activity of these D₂R reduces the glutamate drive to the prefrontal cortex and further worsens the hypoglutamatergic state in schizophrenia. As it was postulated early on [104], DA receptors in meso-limbic DA transmission are a major target for antipsychotic drugs that improve the emotional state of the schizophrenic patients.

We have proposed that the A_{2A}R agonists may be used as anti-schizophrenic drugs through their antagonism of D₂R signaling in the A_{2A}R-D₂R heteromer in the soma-dendritic region of the ventral striato-pallidal GABA pathway. This would help to reestablish the glutamate drive from the mediodorsal thalamic nucleus to the prefrontal cortex [15]. In fact, A_{2A}R agonists strongly reduce the D₂R agonist binding affinity in the nucleus accumbens shell and core [105] diminishing both D₂R recognition and G protein coupling. The A_{2A}R-D₂R heteromer also exists on the glutamate terminals of the local circuits in the striato-pallidal GABAergic neurons and counteracts D₂R-induced inhibition of glutamate release upon A_{2A}R activation. This increase in glutamate release after A_{2A}R agonist treatment will also contribute to enhance the excitability of the ventral striato-pallidal GABA pathway and thereby add to the antipsychotic activity of A_{2A}R agonists. It is important to note that D₂ autoreceptors are not directly modulated by A_{2A}R agonists since A_{2A}R does not exist in the DA terminal networks. In this way,

A_{2A}R agonist treatment will not affect the function of the D₂ autoreceptor to further contribute in lowering DA release.

The anti-schizophrenic potential of A_{2A}R agonists is further underlined through behavioral analysis in the amphetamine and phencyclidine rat models of schizophrenia [106] and in the *Cebus apella* monkey model of schizophrenia [107] in which it demonstrated an atypical antipsychotic profile. Therefore, A_{2A}R agonist treatment represents a new strategy for the treatment of schizophrenia, especially in combination with very low doses of atypical and/or typical D₂R antagonists, to further reduce the development of extrapyramidal side effects, see [15, 45].

Disturbances in the A_{2A}R molecular mechanisms especially in the A_{2A}R-D₂R receptor heteromer in schizophrenia should be considered since deficits in the operation of the antagonistic A_{2A}R-D₂R interaction may increase the vulnerability to develop the disease [15]. However, it should be noted that treatment with A_{2A}R antagonists in PD has so far not led to an increase in psychotic episodes in PD patients, see [68], possibly due to a low adenosine and in particular DA tone in the nucleus accumbens and other parts of the ventral striatum in the PD patients on A_{2A}R antagonist monotherapy.

Dual-probe microdialysis evidence from the Tanganelli group (Ferraro *et al.*, unpublished data (see [15]), in awake, freely moving rats supports the A_{2A}R agonist treatment strategy in schizophrenia. It was demonstrated that the D₂R-D₃R agonist quinpirole (10 µM) when superfused in nucleus accumbens was able to reduce accumbal extracellular GABA levels while increased extracellular GABA levels in the MD nucleus, medial division. The actions of the D₂R agonist were counteracted by the A_{2A}R agonist CGS 21680 (1 µM) when co-superfused with quinpirole into the nucleus accumbens. These results provide for the first time the functional evidence for a connection between the ventral striato-pallidal GABA pathway with the MD medial division which is known to innervate the prefrontal cortex via widespread glutamate projections via the ventral pallidal-MD GABA pathway, see [108].

Furthermore, in agreement, Ferraro *et al.* (unpublished data) found using dual probe microdialysis that extracellular levels of glutamate in the prefrontal cortex were reduced after accumbal superfusion with quinpirole (10 µM), see

[15]. The A_{2A}R agonist CGS 21680 (1 μM) when co-superfused with the D₂R agonist in the nucleus accumbens not only counteracted the D₂R agonist induced reduction of prefrontal glutamate levels, but even resulted in a small rise of extracellular levels of glutamate in the prefrontal region. These results give functional neurochemical evidence that A_{2A}-D₂ receptor-receptor interactions in the nucleus accumbens have substantial relevance for the activity of the MD glutamate projections to the prefrontal cortex and schizophrenia in view of hypoglutamatergia and hypofunction in the dorsolateral prefrontal cortex of schizophrenic patients, see [109].

Within the nucleus accumbens, quinpirole, upon local superfusion, reduced the extracellular levels of accumbal glutamate probably via inhibition of glutamate release from cortico-striatal glutamate terminals known to possess D₂R. These glutamate terminals may also possess A_{2A}R that interact with D₂R and are known to release glutamate [110]. In agreement, the A_{2A}R agonist CGS 21680 superfused into the nucleus accumbens in the present experiments produced a substantial and prolonged release of glutamate (Ferraro *et al.*, unpublished data); see [15]. Interestingly when CGS 21680, at a concentration by itself ineffective on cortical extracellular glutamate levels, was co-perfused in the nucleus accumbens with quinpirole, it significantly antagonized the reduction of extracellular glutamate levels induced by quinpirole in the prefrontal cortex (Ferraro *et al.*, unpublished data), see [15].

Taken together, these results give evidence of the importance of antagonistic A_{2A}R-D₂R interactions in both the ventral striato-pallidal GABA pathway and in the cortico-striatal glutamate terminals in the control of the prefrontal glutamate projections via the ventral pallidum and MD, a loop with major disturbances in schizophrenia. Jones, Popken and colleagues [109] have demonstrated that there exists a subnucleus specific loss of nerve cells in the medial thalamus of schizophrenics. Through stereological counts, a 30% loss of nerve cells was demonstrated in the MD nucleus primarily confined to the parvocellular and densocellular subnuclei. It is of substantial interest that the parvocellular part projects to the dorsolateral parts of the prefrontal cortex and other regions known to be compromised in schizophrenia, see [109]. These results underline the relevance of the present strategy of targeting the A_{2A}R-D₂R heteromer in the nucleus accumbens by

A_{2A}R agonist treatment alone or in combination with low doses of D₂R antagonists to help restore the glutamate drive from MD to the dorsolateral prefrontal cortex.

2.1.4. Relevance to cocaine addiction and its treatment

An increase in A_{2A}R in the nucleus accumbens has been observed after extended cocaine self-administration, see [111]. Following a 10-day cocaine self-administration procedure, one that increased accumbal D₁R, D₂R and D₃R signaling through a cocaine-induced increase in extracellular DA levels via its well-known blockade of the DA transporter, gave rise to a compensatory up-regulation of A_{2A}R that diminished during a cocaine withdrawal period. Behavioral pharmacological results have demonstrated that A_{2A}R antagonists reinstate cocaine self-administration [112] and that A_{2A}R agonists diminish the reinforcing effects of cocaine [113]. Furthermore, A_{2A}R agonists counteract the development and expression of sensitization to the locomotor activation effects of cocaine [114]. In fact, antagonistic A_{2A}R-D₂R interactions have been demonstrated in the nucleus accumbens at both the binding and behavioral level and at the level of neuronal function [24, 55, 60, 93, 105].

A direct action of cocaine on the A_{2A}R is not involved in producing this probable rise of accumbal A_{2A}R signaling in the animals with cocaine still present in the brain [111]. The disappearance of the A_{2A}R rise during the withdrawal period may help to explain the increased reinforcing efficacy of cocaine in animals after 7-days of cocaine withdrawal [115]. This could involve a compensatory mechanism to increase signaling via accumbal D₂R and D₃R by reducing the A_{2A}R brake on D₂R and D₃R signaling [111]. The results indicate a putative role of antagonistic A_{2A}R-D₂R interactions at the membrane (in A_{2A}R-D₂R heteromers) and cytoplasmatic level in the nucleus accumbens in the prevention of development of cocaine addiction. These results open up the possibility that A_{2A}R agonists can represent cocaine antagonists to be used in the prevention of cocaine addiction. This is not supported by the fact that the lack of A_{2A}R signaling reduces the reinforcing

efficacy of cocaine [116] in A_{2A}R knockout mice but may be explained by a reorganization of the D₂R containing RM in these transgenic mice.

The observed rise of A_{2A}R in the nucleus accumbens after extended cocaine self-administration may depend on the existence of an atypical cAMP response element (CRE) in the core promoter of the A_{2A}R gene [117]. CREB (CRE binding protein) diminishes cocaine reward in this region [118] and is enabled by increased activation of the extracellular signal-related kinase [119]. A_{2A}R and D₂R are collocated in the ventral striato-pallidal GABAergic pathway [120-122] and cocaine-induced activation of the D₂R can produce an increase in CREB phosphorylation via several intracellular mechanisms, see [15, 60, 111].

It should also be considered that the A_{2A}R up-regulation reflects not only increases in e.g. A_{2A}R-D₂R heteromers but also increased formation of A_{2A}R homomers that further increase the excitability of the striato-pallidal GABAergic neurons [24, 55, 60, 89] by counteracting D₂R-mediated inhibition of these neurons. Another mechanism could be that persistent D₂-like receptor activation sensitizes A_{2A}R signaling at the level of the AC via the release of $\beta\gamma$ dimers from the activated G_i proteins [24, 55, 60, 89, 123-125]. It is of interest that the increased density of A_{2A}R in the nucleus accumbens after cocaine self-administration demonstrated a reduced affinity of the A_{2A}R antagonist binding sites. This may, *inter alia*, reflect the formation of novel A_{2A}/D₂-like RM with a different stoichiometry and/or topology that produces conformational changes in the receptors of this RM, which lead to altered allosteric interactions and changes in the affinity of the A_{2A}R.

2.2. A_{2A}R-D₃R and A_{2A}R-D₄R heteromers

2.2.1. Biochemical and functional findings

A study by Torvinen *et al.* [126] demonstrated a specific and high FRET efficiency in cells transiently co-transfected with A_{2A}R-YFP and D₃R-GFP² receptors providing evidence that A_{2A}R and D₃R receptors form an A_{2A}R-D₃R heteromer (Fig. 2). Also similar to the D₂R, the D₃R contains the arginine-rich epitope in its IC3 that makes possible an electrostatic interaction with the carboxyl terminus of the A_{2A}R [55]. Evidence was also obtained in

membranes prepared from stably transfected CHO cell lines, in which A_{2A}R activation reduces D₃R agonist binding and D₃R signaling. This provided the evidence for an antagonistic A_{2A}R-D₃R interaction in A_{2A}R-D₃R heteromers similar to the antagonistic interaction observed in the A_{2A}R-D₂R heteromer. A_{2A}R-D₃R heteromers may therefore exist in the nucleus accumbens where the A_{2A}R and D₃R are co-distributed provided that they are co-expressed in the same neuron. In view of the existence of D₃R dimers and tetramers in brain [127] the existence of higher order A_{2A}R-D₃R heteromers (A_{2A}-D₃ RM) should be considered.

The existence of A_{2A}R-D₄R heteromers has also been postulated based on the existence of the arginine-rich epitope in the IC3 loop of the D₄R, which can interact with the negatively charged epitopes in the A_{2A}R carboxyl terminus (Fig. 2) [55]. Recently, it has also been possible to demonstrate the existence of A_{2A}R-D₄R heteromers through BRET experiments in transiently co-transfected A_{2A}R-D₄R in cell lines (Borroto-Escuela *et al.* unpublished data). The A_{2A}R and D₄R may be codistributed especially in the island (striosome, patch) striato-nigral GABAergic system at the soma-dendritic level [128], where it is postulated that A_{2A}R-D₄R heteromers may exist. The striatal island system is involved in cognitive, reward and motivational functions, see [129, 130], which may be modulated by the postulated A_{2A}R-D₄R heteromers.

2.2.2. Relevance to CNS diseases and their treatments

The D₃R is being considered a target for novel anti-schizophrenic drugs that display a D₃R antagonist profile [131, 132]. Therefore A_{2A}-D₃ RM offer possibilities for novel treatment strategies of this disease that might include the combined use of novel D₃R antagonists and A_{2A}R agonists in order to reduce D₃R signaling.

Indirect indications have been obtained for a rise in D₃R density in the nucleus accumbens in cocaine self-administering animals after experiencing cocaine withdrawal [111] which are in line with previous results that demonstrated increases in D₃R binding after cocaine withdrawal that was also associated with an increase in cocaine-seeking behavior [133, 134]. D₃R antagonists counteract cocaine seeking and cocaine enhanced reward and

may be used in treatment of cocaine addiction [131, 135, 136]. Furthermore, an up-regulation of D₃R mRNA levels was found in reward networks of human cocaine fatalities [137]. Therefore, the reduction not only of the antagonistic A_{2A}R-D₂R interaction but also of the antagonistic A_{2A}R-D₃R interaction in animals after 7-days of cocaine withdrawal [111] may contribute to the increased motivation to self-administer cocaine [115]. The A_{2A}R may also have a role in cocaine addiction through its potential modulation of D₄R in the island striato-nigral DA system in view of the demonstration of A_{2A}R-D₄R heteromers in cotransfected cell lines (Borroto-Escuela *et al.*, unpublished data).

2.3. A_{2A}R-D₂R containing receptor mosaics

2.3.1. A_{2A}-D₂R-mGlu₅ receptor mosaic

2.3.1.1. Biochemical and functional findings

The existence of functional A_{2A}R-D₂R-mGlu₅R oligomers in the GABAergic striato-pallidal neuron has often been discussed based on the high and selective co-expression of mGlu₅R, D₂R and A_{2A}R in these particular cells, on the demonstration of A_{2A}R-D₂R heteromers (see above) and A_{2A}R-mGlu₅R [138] heteromers and on the existence of strong multiple interactions between the three receptors [15]. The existence of neurotransmitter receptor heteromers in general and A_{2A}R-D₂R heteromers in particular is now broadly accepted and reinforced by the fact that the functional meaning of heteromerization is being revealed. Thus, the heteromerization of neurotransmitter receptors and their existence as RM have been demonstrated in neuronal cells as functional entities that possess different biochemical characteristics with respect to the individual components of the RM. Therefore, the heteromer might be considered as a molecular switch that fine-tunes the information flow between neurons, thus the signaling mediated by a single stimulated receptor within the heteromer might be, from a qualitative and/or quantitative point of view, different to that expected when all the receptors are simultaneously stimulated. Interestingly, the existence of RM or higher-order receptor heteromers has been recently demonstrated [139, 140].

Taking advantage of the recent fluorescence-based approaches to study protein-protein interactions, we have recently demonstrated the existence of higher-order A_{2A}R-D₂R-mGlu₅R oligomers or RM (Fig. 4). Initially, by using bimolecular fluorescence complementation (BiFC), we visualized for the first time the occurrence of mGlu₅R-D₂R heterodimers in living cells [139]. Furthermore, the combination of BiFC and BRET techniques allowed us to detect the existence of receptor oligomers containing more than two protomers, namely A_{2A}R-D₂R-mGlu₅R higher-order oligomers or RM (Fig. 4) [139]. Thus, this new experimental approach has allowed the study of the quaternary structure of A_{2A}-D₂-mGlu₅ RMs.

Interestingly, by using triple-labeling post-embedding immunogold and detection at the electron microscopic level, the precise simultaneous distribution of A_{2A}R, D₂R and mGlu₅R in striatal neurons has been performed. It is noticeable that these three receptors co-distributed in post-synaptic structures along the extra-synaptic and peri-synaptic plasma membrane of spines that establish asymmetrical, putative glutamatergic, synapses with axon terminals [139]. Overall, this is the first direct anatomic evidence for mGlu₅R, D₂R and A_{2A}R co-distribution in the same neuronal compartment and supports the notion of that these receptors form a RM in GABAergic striatopallidal neurons.

2.3.1.2. Relevance to CNS diseases and treatments

The A_{2A}-D₂-mGlu₅ RM may mainly be in operation to produce activation of the cortico-striatal glutamate synapse and the striato-pallidal GABAergic neurons when motor inhibition of certain movements is required [15]. The increased firing in the glutamate terminals will release glutamate and co-stored ATP that will result in an increased formation of adenosine and lead to the increased activation of both pre- and postjunctional A_{2A}R and mGlu₅R receptors that synergize to counteract the D₂R signaling in the glutamate terminals and in the striato-pallidal GABAergic neurons. In this way, the firing of the striato-pallidal GABAergic neurons can develop without being restrained by the D₂R signaling that is aimed to silence the striato-pallidal GABAergic neurons. Once the firing in the cortico-striatal glutamate pathways

slows down the balance between glutamate and DA signaling, the RM will reach another set-point dependent on the movements to be initiated.

Parkinson's disease. The development of mGlu₅R antagonists are yet another strategy for treatment of PD based on their ability to enhance D₂R recognition and signaling in these RM in the dorsal striato-pallidal GABAergic pathway by the removal of the antagonistic mGlu₅R-D₂R interaction. In addition, mGlu₅R antagonists block the ability of mGlu₅R to enhance NMDA receptor signaling, which will also favor anti-Parkinsonian actions by reducing the excitability of the striato-pallidal GABAergic neurons and thus their ability to cause motor inhibition, see [141].

It should be noted that the ability of mGlu₅R antagonists to produce motor activation requires both A_{2A}R and D₂R, which underlines their interdependence and supports the concept of A_{2A}R-D₂R-mGlu₅R RM [142]. The synergism of A_{2A}R and mGlu₅R antagonists to increase locomotion in reserpinized mice [142, 143] can be elegantly explained by the existence of A_{2A}-D₂-mGlu₅ RM where A_{2A}R-mGlu₅R synergize to counteract D₂R signaling, see [68].

The postulated A_{2A}-D₂-mGlu₅ RM in the striato-pallidal GABAergic neurons with multiple receptor-receptor interactions is therefore, a novel target for anti-Parkinsonian drugs. The above results have led to the proposal that mGlu₅R antagonists especially in combination with A_{2A}R antagonists or drugs with both A_{2A}R and mGlu₅R antagonist properties are symptomatic anti-Parkinsonian drugs of special value particularly in view of their neuroprotective properties, see also [15, 60].

Schizophrenia. The postulated A_{2A}-D₂-mGlu₅ RM may also exist in the ventral striato-pallidal GABAergic neurons and in the glutamate terminal networks of the ventral striatum. In fact, A_{2A}R and mGlu₅R agonists synergize when co-superfused into the nucleus accumbens to increase GABA release in the ventral pallidum [144]. Evidence for a role of mGlu₅R in schizophrenia-related behavior in rodents such as prepulse inhibition has also been obtained [145]. Therefore, combined treatment with A_{2A}R and mGlu₅R agonist drugs or drugs with combined A_{2A}R and mGlu₅R agonist properties may be an effective novel strategy for treatment of schizophrenia based on the synergistic A_{2A}R-mGlu₅R interaction which should be able to override the pathologically increased D₂R

signaling in this RM that may potentially be present in schizophrenia. The addition of a low dose of a D₂R antagonist to the A_{2A}R-mGlu₅R agonist treatment should also be considered.

2.3.2. A_{2A}-CB₁-D₂ receptor mosaic

2.3.2.1. Biochemical and functional findings

In previous work [146, 147] indications were obtained for the existence of cannabinoid-dopamine CB₁R-D₂R heteromers in co-transfected HEK-293 cells based on FRET analysis and for an antagonistic CB₁R-D₂R interaction based on D₂R binding analysis. In 2005, studies using co-immunoprecipitation in HEK-293 cells gave further indications for CB₁R-D₂R heteromers with an enhanced formation after concurrent activation of the two receptors. It was noticed that in this heteromer CB₁R signaling, in part, switches from the inhibition of the AC to a pertussis toxin-insensitive activation of AC [148], see also [149].

Colocalization of D₂R and CB₁R in striatum was first observed in 2002-2003 [150, 151] and have been later found particularly in cortico-striatal glutamate terminals, in the soma and dendrites of ventral striato-pallidal GABAergic neurons and in local collaterals of the striato-pallidal GABAergic neurons, see [152, 153]. Therefore, a chemical anatomical basis for CB₁R-D₂R interactions exists and results indicate that endocannabinoids, via inhibitory feedback, can counteract D₂R-mediated responses [154-156]. Recently, novel evidence has been obtained for the existence of CB₁R-D₂R heteromers based on FRET analysis in HEK-293 cells [157]. Thus, FRET data show a strong and specific FRET signaling in cells co-transfected with cDNA of vectors encoding for D₂R-GFP² and CB₁R-YFP but not in various controls including mixtures of cells expressing D₂R-GFP² or CB₁R-YFP or co-transfections with GFP² or YFP without being tagged to the corresponding receptor. Of substantial interest is the finding of antagonistic D₂R binding modulation by CB₁R agonists that reduce D₂R agonist affinity in striatal membranes. Similar results were also observed in the nucleus accumbens shell demonstrated by quantitative receptor autoradiography [157]. In agreement, behavioral analysis revealed that CB₁R agonists can counteract

D₂R agonist-induced hyperlocomotion, an effect that was blocked by rimonabant, a CB₁R antagonist, which also can enhance the action of the D₂-like receptor agonist quinpirole [157]. These results clearly suggest that antagonistic intramembrane CB₁R-D₂R interactions exist in CB₁R-D₂R heteromers in the ventral and dorsal striatum and lead to reduced D₂R signaling, increased excitation and firing of the striato-pallidal GABAergic neurons and counteraction of D₂R-induced hyperlocomotion.

Of special interest in this behavioral analysis was the observation that the A_{2A}R antagonist, MSX-3, could prevent the ability of the CB₁R agonist CP 55,940 to counteract the D₂R agonist-induced hyperlocomotion [157]. These results indicated the involvement of A_{2A}R that is also known to exist in the cortico-striatal glutamate terminals and the soma-dendritic regions of the striato-pallidal GABAergic neurons in the antagonistic intramembrane CB₁R-D₂R interaction (see above). In line with these results, A_{2A}R-CB₁R heteromers were also demonstrated in transiently transfected HEK-293 cells and in neuroblastoma cells in which CB₁R signaling was entirely dependent upon A_{2A}R activation [158]. Finally, it was also shown that motor depression caused by CB₁R agonists was blocked by A_{2A}R antagonists. From the accumulating evidence, the existence of A_{2A}-CB₁-D₂ RM was therefore postulated [157].

The sequential BRET-FRET (SRET) technique was developed specifically for the identification of trimeric receptor mosaics [140]. Through a combination of BRET and FRET, trimeric receptor mosaics could finally be identified. This method was the essential technique to finally identify receptor mosaics, [15-17, 35, 43, 157]. Using the SRET technique, the A_{2A}-CB₁-D₂ RM was the first RM to be identified in living cells, and this discovery is in line with the indications for its existence obtained in previous work on the brain, see [157]. This RM is an integrator of DA, adenosine and endocannabinoid signals.

The present hypothesis for the operation of this putative A_{2A}-CB₁-D₂ RM in the brain states that the antagonistic CB₁R-D₂R interaction activated by the D₂R-induced release of endocannabinoids into the extracellular fluid removes the D₂R brake on A_{2A}R signaling to AC, see [15], by an inhibitory feedback mechanism by the activation of CB₁ receptors in putative A_{2A}-CB₁-D₂ RM in the ventral striato-pallidal GABAergic neurons and in cortico-striatal

glutamate terminals. The increase in A_{2A}R signaling in the ventral striato-pallidal GABAergic neurons will, via DARPP-32 phosphorylation at Thr34, strongly contribute to the markedly increased activity in ventral striato-pallidal GABAergic neurons by the inhibition of protein phosphatase-1, which will enhance the phosphorylation of ion channels and ion channel-linked receptors. The simultaneous release of A_{2A}R signaling in striatal glutamate terminals will increase glutamate release and therefore lead to an increase in the glutamate drive of the ventral and dorsal striato-pallidal GABAergic neurons. Such an operation of this RM may be the molecular basis for the observed blockade of the D₂R agonist-induced locomotor hyperactivity and contribute to CB₁R agonist-induced motor inhibition [157]. It seems likely that the former is mainly in operation as an inhibitory feedback mechanism to reduce an exaggerated and prolonged activation of D₂R that will produce a prolonged silencing of the striato-pallidal GABAergic neurons. The release of the D₂R brake on A_{2A}R-induced activation of AC probably plays a major role in making this possible.

2.3.2.2. Relevance to CNS diseases and their treatments

Parkinson's disease. The results from Marcellino *et al.* [157] clearly suggest that CB₁R antagonists may represent novel symptomatic anti-Parkinsonian drugs by enhancing D₂R signaling to uphold its brake on A_{2A}R signaling at the level of AC in the putative A_{2A}-CB₁-D₂ RMs. These molecular events may explain the enhancement of D₂R agonist-induced hyperlocomotion by the CB₁R antagonist and its ability to counteract the CB₁R agonist-induced inhibition of D₂R-induced hyperlocomotion.

It also seems likely that low doses of CB₁R and A_{2A}R antagonists may synergize to enhance D₂R-mediated anti-Parkinsonian actions in early Parkinson's disease where DA release remains to a substantial degree from the remaining DA nerve terminal networks. The A_{2A}R antagonist will also act by interfering with the ability of A_{2A}R to inhibit D₂R signaling in the A_{2A}-D₂-mGlu₅ RM as discussed above. In contrast, in late PD with a very low DA tone, it becomes necessary to add low threshold doses of L-DOPA or D₂R agonists to achieve substantial therapeutic activity from CB₁R and A_{2A}R

antagonists. Through the possibility to use low doses of the two and/or three drugs, side effects such as dyskinesias may be effectively reduced.

In line with these results, CB₁R antagonists can strongly enhance the stereotypies caused by combined treatment with D₁R and D₂R agonists [159]. Furthermore, increased CB₁R binding and G protein-coupling has been observed in the basal ganglia of patients with Parkinson's disease and in the MPTP marmoset model of Parkinson's disease [160]. These neurochemical effects are counteracted by L-DOPA therapy suggesting that the CB₁R changes observed, represent a CB₁R receptor up-regulation in response to reduced D₂R signaling in PD that fails to elicit the release of endocannabinoids [161] and thus fails to activate the inhibitory feedback via the CB₁R. Similar results have been observed by Strömberg, Andersson and colleagues [162] after chronic haloperidol blockade of the D₂R. This mechanism can also help explain the reduced expression of cannabinoid CB₁R mRNA in the basal ganglia of postmortem brain of Parkinsonian patients [163] as a result of the dopaminergic treatment. The CB₁R antagonists should therefore act, as postulated above, to counteract the D₂R-activated inhibitory feedback activation of the CB₁R in the A_{2A}-CB₁-D₂ RM with the aim to bring down D₂R signaling. The enhancement of D₂R signaling in this RM should be optimized by a combined treatment with CB₁R and A_{2A}R antagonists in order to block the two allosteric mechanisms of antagonizing the D₂R in A_{2A}-CB₁-D₂ RM.

Schizophrenia. Antagonistic CB₁R-D₂R interactions may also exist in postulated A_{2A}-CB₁-D₂ RM in the ventral striato-pallidal GABAergic neurons and in the cortico-accumbal glutamate terminals. The possibility is open that CB₁R agonists may possess antipsychotic properties by their ability to reduce D₂R signaling in this pathway and in the afferent glutamate terminals that will lead to a reduction of positive symptoms of schizophrenia. This is however, in apparent disagreement with the fact that Δ^9 -tetrahydrocannabinol (THC) is reported to exacerbate psychotic symptoms in schizophrenia, see [164]. However, these actions may be exerted at CB₁R in other brain regions *inter alia* the cerebral cortex.

These novel observations may help explain the findings that the increased CSF levels of anandamide found in schizophrenic patients are

inversely correlated with psychotic symptoms [165]. Thus, an over activity of D₂R-mediated DA transmission in the ventral striatum may lead to an increased formation of the endocannabinoid anandamide with an increased inhibitory feedback on the D₂R signaling via the CB₁R-D₂R antagonistic interaction in the A_{2A}-D₂-CB₁ RM on the glutamate terminals and in the ventral striato-pallidal GABAergic neurons. In this way, the excessive D₂R-mediated inhibition of the ventral striato-pallidal GABAergic neurons may be reduced by anandamide, which can act as an agonist in the postulated A_{2A}-CB₁-D₂ RMs. The precipitation of psychotic periods by cannabis use may also be related to the reduction of anandamide signaling in the brain [165] that leads to a reduction in the antagonistic CB₁R-D₂R interaction.

Cocaine addiction. The present indications of antagonistic CB₁R-D₂R interactions in RM in the ventral striato-pallidal GABAergic pathway may help explain the ability of the CB₁R agonist WIN 55,512-2 to counteract the rewarding actions of cocaine in intracranial self-stimulation experiments [166]. Thus, D₂R participate in mediating cocaine reward by inhibiting the activity in this reward-regulating pathway, see [111]. In line with this hypothesis, the CB₁R agonist WIN 55,212-2 can also reduce cocaine self-administration [167]. Our interpretation is that the CB₁R agonist, via the antagonistic CB₁R-D₂R interaction, leads to an increased activity in the ventral striato-pallidal GABAergic neurons thereby reducing the reward value of cocaine. As stated above, A_{2A}R agonists can also reduce cocaine self-administration. Therefore, we postulate that combined treatment with A_{2A}R agonists and CB₁R agonists that preferentially activates the A_{2A}R and CB₁R in A_{2A}-CB₁-D₂ RM should represent an interesting and novel strategy for preventing the development of cocaine abuse.

3. A₁R-D₁R heteromers and A₁R-D₁R containing receptor mosaics

3.1. A₁R-D₁R heteromers

3.1.1. Biochemical and functional findings

A₁R and D₁R were shown to co-immunoprecipitate in co-transfected Ltk- fibroblast cells [168], a phenomenon that appeared specific, since co-immunoprecipitation was not detected in A₁R-D₂R co-transfected Ltk-fibroblast cells. The A₁R-D₁R co-immunoprecipitation was observed in the

absence of A₁R or D₁R receptor agonist exposure, thereby indicating their constitutive formation. However, the A₁R-D₁R co-immunoprecipitation was substantially reduced after a 1 h treatment with the D₁R agonist SKF 38393, giving evidence that D₁R activation leads to disruption of the A₁R-D₁R heteromeric receptor complex. This disruption did not occur if combined treatment with SKF 38393 and the A₁R agonist *R*-PIA was applied. Thus, these initial results indicated that A₁R and D₁R form receptor heteromers at least in cell lines. Later, co-immunoprecipitation experiments indicated that they may also exist in striatum [169]. Further evidence that an A₁R-D₁R heteromer exists was recently obtained, since specific BRET and FRET signals can be detected between fluorophore-tagged A₁R and D₁R upon transient co-transfection in cell lines [170]. The A₁R-D₁R heteromer may exist on the cell surface membrane together with A₁R homodimers [171] and D₁R homodimers [172], and it is not known if the A₁R-D₁R heteromer is preferred.

Competition experiments with the D₁R antagonist [³H]-SCH 23390 versus dopamine were performed in striatal membrane preparations as well as in membrane preparations from an A₁R-D₁R co-transfected Ltk- cell line [173, 174]. The A₁R agonist CPA, in the nanomolar range, caused a marked reduction in the proportion of D₁R in its high affinity state. These effects were mimicked by the GTP analogue Gpp (NH)p (100 μM). These results make it likely that A₁R activation in the A₁R-D₁R heteromer leads to an uncoupling of the D₁R to its G_{s/olf} protein.

The possible role of the G_i protein in the A₁R-D₁R interaction at the binding pocket level was studied using pertussis toxin, since it inactivates the G_i protein coupled to the A₁R. It was found that pertussis toxin treatment blocked the effects of low but not high concentrations of the A₁R agonist on D₁R binding characteristics [174]. However, it could not be determined if the D₁R modulation by the high 10 μM concentration of CPA was due to activation of the low affinity A₁R receptors or to the activation of the remaining high affinity A₁R, since pertussis toxin-induced ribosylation of the G_i protein was not complete.

This problem could however be solved by involving adenosine deaminase (ADA) in the analysis, which metabolizes adenosine to inosine. This enzyme binds to A₁R as an ectoenzyme [175] and is necessary to obtain

the high affinity binding state of A₁R [176]. An irreversible inhibitor of ADA, deoxycoformycin (DCF), was found to fully counteract the effects of high and low concentrations of CPA on the binding characteristics of the D₁R. The blockade of the enzymatic activity of ADA was shown not to be involved in this action of DCF. These results give evidence that it is the high affinity state of A₁R that is responsible for the interaction with D₁R, at least at the level of the binding pocket [177]. Thus, ADA may directly bind to the A₁R, which is necessary for the A₁R high affinity state to develop. This state has a protein conformation such that the A₁R antagonistically interacts with the D₁R. It follows that functional A₁R-D₁R heteromer requires ADA to be bound to A₁R, underlining the important role of receptor interacting proteins in heteromers [24, 170].

In the A₁R-D₁R co-transfected fibroblast cell line, the expected antagonistic interaction at the level of AC level was observed after A₁R and D₁R co-activation [174]. The antagonistic A₁R-D₁R interaction found at both the recognition and AC levels was found to be correlated, indicating that the inhibitory interaction in the A₁R-D₁R heteromer was involved in causing the A₁R-mediated inhibition of D₁R signaling.

The available evidence suggests that there exist antagonistic intramembrane A₁R-D₁R interactions in the dorsal and ventral striatum and in the prefrontal cortex [24, 25, 170, 178]. This involves also an ability of A₁R agonists to antagonistically modulate D₁R antagonist binding sites in the nucleus accumbens and the prefrontal cortex that cause a reduction of their affinity.

Agonist-induced co-aggregation and co-desensitization of A₁R-D₁R heteromers

Permeabilized cells were used and the A₁R agonist R-PIA (100 nM, 1 h) caused aggregates of A₁R-D₁R, while the D₁R agonist SKF 38393 (10 μM, 1 h) caused clusters of D₁R alone, in line with the D₁R agonist induced disruption of the A₁R-D₁R heteromers [168]. It is of substantial interest that combined treatment with the two agonists, which maintains the heteromerization of A₁R and D₁R, reduced the co-aggregation of A₁R-D₁R. In this case, a clear-cut decrease in D₁R signaling to the AC was observed. Thus, upon co-activation of A₁R and D₁R in the heteromer, with no formation

of co-aggregates and maintenance of A₁R-D₁R heteromers, a desensitization of D₁R signaling occurs. This desensitization may involve an uncoupling of the D₁R in the A₁R-D₁R heteromer to the G_s protein. In contrast, A₁R-D₁R co-aggregates (activation of A₁R alone) or D₁R aggregates (activation of D₁R alone) did not result in desensitization of D₁R signaling [168]. In contrast to the A₁R-D₁R co-transfected fibroblast cell line, combined treatment with A₁R and D₁R agonists produced co-aggregates in cortical nerve cells in culture. Such differential actions may be caused by differences in the stoichiometry and in adapter and scaffolding proteins interacting with the A₁R-D₁R heteromers but nevertheless indicate an important role of A₁R and D₁R agonists in modulating the cotrafficking of A₁R-D₁R heteromers in the forebrain and thus in D₁R function.

Function. An A₁R antagonistic modulation of D₁R signaling was observed in the regulation of transcription factors in the striatum after DA terminal denervation based on analysis of the immediate early gene NGFI-A and *c-fos* mRNA levels and of GABA release in the striato-entopeduncular GABAergic pathway as studied with microdialysis [178]. The same year behavioral indications for antagonistic A₁R-D₁R receptor interactions were also obtained with adenosine A₁R antagonists potentiating the motor effects of D₁R agonists [179]. Collectively, this work in the 1990s indicated the existence of antagonistic intramembrane A₁R-D₁R interactions reducing D₁R signaling in the direct pathways (see [24]).

3.1.2. Relevance to CNS diseases and their treatments

The antagonistic interaction in A₁R-D₁R heteromers, where D₁R is the crucial receptor in view of its important behavioral role, offers a new way to modulate D₁R signaling, namely to reduce striatal D₁R signaling with A₁R agonists and enhance it with A₁R antagonists (see [24, 25]). This offers a therapeutic potential for A₁R antagonists in Parkinson's disease as seen also by A₁R antagonist enhancement of D₁R-induced locomotion. Often, A₁R drugs in low doses by themselves have only weak effects in the behavioral models used but can strongly modulate the D₁R signaling [179, 180]. This is also beautifully illustrated through microdialysis studies and through the induction

of immediate early genes (IEG) (see above and [181]). In addition, A₁R agonists can strongly counteract the D₁R agonist-induced oral dyskinesias in rabbits [173] indicating a therapeutic potential of A₁R agonists in L-DOPA-induced dyskinesias in parkinsonian patients by targeting the striatal A₁R-D₁R heteromer.

A₁R agonists also counteract D₁R agonist-induced electroencephalography (EEG) arousal in rats [182] probably by targeting the postulated A₁R-D₁R heteromers in the frontal cortex. A₁R antagonists targeting these heteromers may therefore, have a potential therapeutic role in attention deficit hyperactivity disorders (ADHD) by increasing EEG arousal provided it will lead to increased attention. This proposal is supported by the demonstration of sedative-hypnogenic properties of adenosine analogues that could in part be mediated via A₁R-D₁R heteromers in the frontal cortex. In fact, the A₁R agonist CPA but not the A_{2A}R agonist CGS 21680 prevents EEG arousal due to D₁R activation [182].

3.2. Putative A₁-D₁-D₃ receptor mosaics

As mentioned earlier, a large array of experimental approaches utilizing heterologous expression systems have been used to demonstrate receptor-receptor interactions. Interestingly, by using some of these approaches, namely BRET, FRET and acceptor photobleaching FRET, a D₁R-D₃R interaction has been demonstrated [183]. Interestingly, in membrane preparations from bovine striatum the D₃R agonist R(+) 7-OH-DPAT was found to shift the D₁R agonist competition curve to the left ([³H]-SCH23390 versus SKF-38393) demonstrating that D₃R activation increases the affinity of the D₁R agonist binding sites [183]. These results suggest the existence of synergistic intramembrane D₃R-D₁R interactions at the level of D₁R recognition in striatal D₃R-D₁R heteromers. In line with these findings, the D₃R agonist PD 128907 is also shown to enhance the actions of the D₁R agonist SKF 38393 on locomotion in reserpinized mice [183].

Based on previous work by the Schwartz and Sokoloff group [184-186], see also [23, 131, 173] and in line with the recent findings of Marcellino *et al.* [183] it seems likely that D₃R-D₁R synergism in their receptor heteromer in the

direct striatal pathway may contribute to L-DOPA-induced dyskinesias. D₃R antagonists acting on the D₃R-D₁R heteromer may therefore have anti-dyskinetic properties. In fact, induction of D₃R expression may be one mechanism for behavioral sensitization to L-DOPA [187]. D₃R antagonists blocking D₃R-D₁R synergism may also have anti-cocaine reward properties in view of the D₃R up-regulation found e.g. in reward circuitries of human cocaine fatalities [188] and where both D₁R and D₃R are involved in cocaine actions [187, 189].

Interestingly, Surmeier *et al.* [190] found that around 50% of the striato-nigral and striato-entopeduncular GABAergic neurons show D₃R expression. There is the possibility that at least some of these neurons also express A₁R and that not only A₁R-D₁R heteromers but also A₁-D₁-D₃ RM may be formed in these neurons of the direct pathway. Such a RM composed of three different GPCR types may be an especially important integrative center for transmitter and modulator signals with D₁R as the hub receptor and the other two as receptors important for proper D₁R function. Therefore, it may be the case that combined treatment with D₃R antagonists and A₁R agonists could be rational to reduce D₁R signaling especially in combination with low doses of D₁R antagonists. Overall, this approach may bring about an improved treatment of L-DOPA-induced dyskinesias and of cocaine addiction with reduced side effects in view of the lower doses of D₁R antagonists that can be used, see [15].

4. Final comments

Dopamine receptors in general, and D₂R and D₁R in particular, may be considered as the functionally most important receptors in several types of striatal GPCR heterodimers. Interestingly, in view of their strong actions on multiple effectors in the striato-pallidal and striato-entopeduncular/nigral GABAergic neurons, it is postulated that these dopamine receptors-containing oligomers are mainly located outside the glutamate and DA synapses (see above). The standard treatment, e.g. with L-DOPA and/or DA agonists [191] in PD builds on the activation of these D₂R and D₁R by moderate to high doses, while the accessory A_{2A}R and A₁R and others in the different heterodimers and RM are not targeted.

A novel principle may now be used to develop D₂R agonists for treatment of PD based on the existence of various D₂R containing heterodimers (two receptors) and RM (three or more receptors; higher-order oligomers) since the conformational state of the D₂R and likely its pharmacology probably differs from one receptor assembly to another. It may also be due to different receptor-protein interactions, and also influenced by the local molecular histology of the surface membrane of discrete striato-pallidal nerve cell populations and of striatal glutamate and DA nerve terminal networks. Thus, the full or partial agonist pharmacology of D₂R in terms of potency and efficacy may show substantial differences among various types of RM such as A_{2A}-D₂-mGlu₅ RM versus A_{2A}-CB₁-D₂ RM versus D₂ monomers and homodimers, and A_{2A}R-D₂R and CB₁R-D₂R heterodimers etc. The development of specific D₂R agonists, specifically for the D_{2S} autoreceptor may be especially hopeful since the D_{2S} autoreceptor participates in unique RM versus the postjunctional D_{2L} receptors. In this way, RMs of D_{2S} autoreceptors and non- α 7 nicotinic receptors have been found in the striatal DA nerve terminals [192], see also [193] as well as direct protein-protein interactions of the D_{2S} autoreceptor with the DA transporter which are disrupted in schizophrenia [194]. This should give exciting new possibilities to develop novel and more selective D₂R agonist drugs for treatment of Parkinson's disease by preferentially acting on certain postjunctional RM in the striato-pallidal GABAergic neurons and their glutamate inputs.

The same principle may also be used to develop novel D₂R antagonists for the treatment of schizophrenia. Thus, the potency and efficacy of full and partial D₂R antagonists and their inverse D₂R agonist activity may vary among the different D₂R assemblies, due to differences in the conformational state of the participating D_{2L} postjunctional and D_{2S} autoreceptors which give them differences in D₂R antagonist pharmacology. The major target may be the postjunctional D₂R in the ventral striato-pallidal GABAergic neurons inhibiting the glutamate drive to the prefrontal cortex in view also of likely increases in meso-limbic DA activity in schizophrenia, see [15, 17, 24, 25, 103]. It should be mentioned that the atypical antipsychotic drug remoxipride, a selective D₂R antagonist, unlike haloperidol *in vivo* blocks only a subpopulation of D₂R in nigro-striatal and meso-limbic/cortical regions

as evaluated by the protection against the N-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ) induced decreases in D₂R binding due to its irreversible inactivation of D₂R [102]. It was suggested that the blockade of this D₂R subpopulation *in vivo* was the basis for its antipsychotic activity and its atypical antipsychotic profile with reduced extrapyramidal side effects. The underlying mechanism may be that this selective D₂R antagonist can only bind and block D₂ receptors in distinct receptor heterodimers and receptor mosaics present in these regions due to their unique D₂R antagonist pharmacology. This may now be tested by studying how the potency and efficacy of remoxipride to block D₂ receptors varies in cell lines upon co-transfection with other receptors known to form heterodimers and receptor mosaics with the D₂R. This approach may also be valid for the treatment of cocaine addiction (see above) where novel and more selective D₂R antagonists for certain D₂R-containing heterodimers and RM in the ventral striatum may offer improvement of treatment.

The other novel principal strategy based on the different D₂R- and D₁R-containing receptor mosaics is the targeting of their accessory receptors that antagonistically interact via receptor-receptor interactions with the D₂R (key receptor) involving e.g. the A_{2A}R, and the D₁R and the A₁R, which can involve combined treatment with very low doses of D₂R agonist/antagonists and D₁R agonist/antagonists, respectively.

In PD, a combined treatment will make possible the use of very low to low doses of L-DOPA and D₂R agonists since in combination with A_{2A}R antagonists the inhibitory impact of these receptors on D₂R is removed [15, 17, 24, 60]. Thus, a reduction of the collateral effects of L-DOPA and D₂R agonists may be observed along with improvement of hypokinesia, resting tremor and rigidity [68]. In early PD, it may be possible to substantially delay the onset of L-DOPA and D₂R agonist treatment by introducing monotherapy or combined therapy with A_{2A}R or CB₁R antagonists which also may delay the neurodegeneration of the nigral DA cells, see [15, 17].

In schizophrenia, low doses of D₂R antagonists with reduced collateral effects may be used since a combined treatment with low doses of A_{2A}R agonists will synergistically reduce D₂R signaling at the membrane and the cytoplasmic level in the ventral striato-pallidal GABAergic neurons [15, 17,

45, 62]. Thus, with this combined treatment anti-schizophrenic actions may be obtained with reduced extrapyramidal side effects and monotherapy with A_{2A}R agonists should also be considered in view of their atypical antipsychotic profile [15, 106]. A similar combined treatment strategy may also be used against development of cocaine addiction.

5. Future research

It becomes important to discover the A_{2A}R-D₂R and A₁R-D₁R heterodimers and RM *in vivo* within the brain and their functional role. This may be accomplished through the generation of BAC transgenic mice [195] with fluorescently tagged wild-type adenosine and dopamine receptors together with mice that express mutant adenosine and dopamine receptor that cannot heteromerize but remain functional. Therefore the heteromers demonstrated can then be analyzed, e.g. in mice models of Parkinson's disease, schizophrenia, etc., in order to understand their role in the pathogenesis and treatments of neurological and mental diseases.

The biochemical *in vitro* work should unravel the pharmacology of the various adenosine and dopamine heterodimers and RM assisted by computerized modeling of the heteromers and RM and bioinformatic analysis of the participating receptors. In this way, adenosine and dopamine receptor interfering drugs may be discovered that preferentially interact with distinct heterodimers and RM of adenosine and dopamine receptors [15, 17].

It is crucial to characterize the receptor interfaces of the various heteromers and RM through a combined biochemical and bioinformatic analysis that includes mathematical approaches. In this way, novel drugs can be developed that directly target the receptor interface in this way mimicking the adenosine-dopamine receptor-receptor interactions that take place via the receptor interfaces through allosteric mechanisms.

Finally, it becomes important to also establish and characterize the role of allosteric modulators in the adenosine and dopamine receptor heterodimers and RM [47] and how they become integrated with the receptor-receptor interactions to modulate the orthosteric sites and the coupling of the receptors to G proteins and β -arrestins. The understanding of such integrative processes is also vital for drug development in neuropsychopharmacology.

Acknowledgements

This work was supported by grants from the Swedish Research Council (04X-715), Torsten and Ragnar Söderberg Foundation, Hjärnfonden and Marianne and Marcus Wallenberg Foundation to KF and grants SAF2008-01462 and Consolider-Ingenio CSD2008-00005 from Ministerio de Ciencia e Innovación to FC. FC and VF-D belong to the “Neuropharmacology and Pain” accredited research group (Generalitat de Catalunya, 2009 SGR 232).

Figure Legends

Figure 1. Schematic illustration of A_{2A}R and D₂R homodimers and A_{2A}R-D₂R heterodimer. The striato-pallidal GABAergic neurons might co-express A_{2A}R and D₂R homo- and heterodimers (dashed box) at the plasma membrane. Adenosine and dopamine can potentially interact with both homo- and heterodimers converging in the control of adenylate cyclase function an integrated cellular response is generated. The functional balance between these three oligomers determines the final adenylate cyclase output and thus the eventual cellular response. The antagonistic allosteric A_{2A}R-D₂R interaction in the heterodimer (dashed box) is shown (filled black arrow) as well as the negative and positive coupling of D₂R and A_{2A}R to the adenylate cyclase, respectively.

Figure 2. Illustration of positively charged arginine-rich epitopes ²¹⁵VLRRRRKRVN²²⁴ (D₂R), ²¹⁶KQRRRKRI²²³ (D₃R), and ³⁷⁷TRRRRRRAK³⁸⁵ (D₄R) in the N-terminal part of the third intracellular loop of D₂R, D₃R, D₄R, electrostatically interacting with negatively charged C-terminal epitopes of the A_{2A}R (³⁷⁰SAQEpSQGNT³⁷⁸, ³⁸⁸HELKGVCPPEPGLDDPLAQDGAGVS⁴¹²). The most important residues in the A_{2A}R appear to be the phosphorylated serine in the ³⁷⁰SAQEpSQGNT³⁷⁸ in the C-terminal epitope of the A_{2A}R (see [52, 55, 64]). These electrostatic interactions represent important hot spots in the receptor interface of the A_{2A}R-D₂R, A_{2A}R-D₃R and A_{2A}R-D₄R heteromers. The prototype was the A_{2A}R-D₂R heteromer (dashed box).

Figure 3. Schematic representation of protein kinase B/glycogen synthase kinase 3 (Akt/GSK3) signalling networks regulated by dopamine D₂R. (left) The stimulation of D₂R lead to an initial change in receptor conformation that mediate the activation of G_{i/o} protein, leading to inhibition of adenylyl cyclase, subsequently to receptor phosphorylation by G-protein receptor kinase and the recruitment of β -arrestin. The recruitment of β -arrestin results in the formation of a signalling complex that comprises at least β -arrestin, PP2A and Akt. The formation of this complex result in the deactivation of Akt by protein phosphatase 2A (PP2A) and the subsequent stimulation of GSK-3 that mediates dopamine-dependent behaviors. (right) The antagonistic allosteric A_{2A}R-D₂R interaction in the heterodimer could reduce β -arrestin recruitment, resulting in an enhancement of Akt phosphorylation, thus inhibiting GSK3 (see text).

Figure 4. Schematic representation of putative A_{2A}-D₂-mGlu₅ RM and A_{2A}R-D₂R heterodimer in the striato-pallidal GABAergic neurons (A) and A₁-D₁-D₃ RM and A₁R-D₁R heterodimer in the striato-entopeduncular/nigral GABAergic neurons (B). In panel A, the antagonistic A_{2A}R-D₂R and mGlu₅R-D₂R interactions in the A_{2A}-D₂-mGlu₅ RM are shown as well as the antagonistic A_{2A}R-D₂R interactions in the heterodimer. The interactions at the level of adenylyl cyclase are also indicated in which D₂R inhibits and A_{2A}R activates this enzyme. D₂R signalling from the putative RM and/or the heterodimer control the excitability of the striato-pallidal GABAergic neurons by gating ion channels. In panel B, the antagonistic A₁R-D₁R and facilitatory D₁R-D₃R interactions in the putative A₁-D₁-D₃ RM are shown as well as antagonistic A₁R-D₁R interactions in the heterodimer. The interactions at the level of adenylyl cyclase are also indicated in which D₁R activates and A₁R and possibly D₃R inhibits this enzyme. The activation of PKA contributes not only to activate intracellular pathways that lead to the phosphorylation of DARPP-32, MEK, and CREB, but also to phosphorylation events that lead to an increase in the activity of cation channels.

References

1. Fuxe K, Ungerstedt U. Histochemical, Biochemical and Functional Studies on Central Monoamine Neurons after Acute and Chronic Amphetamine Administration. In: Costa E, Garattini S, editors. Amphetamines and Related Compounds. New York: Raven Press; 1970. p. 257-88.
2. Agnati LF, Bjelke B, Fuxe K. Volume Transmission in the Brain. *Am Scientist*. 1992;80:362-73.
3. Ungerstedt U, Butcher LL, Butcher SG, Anden NE, Fuxe K. Direct chemical stimulation of dopaminergic mechanisms in the neostriatum of the rat. *Brain Res*. 1969 Jul;14(2):461-71.
4. Descarries L, Beaudet A, Watkins KC. Serotonin nerve terminals in adult rat neocortex. *Brain Res*. 1975 Dec 26;100(3):563-88.
5. Agnati LF, Fuxe K, Zoli M, Ozini I, Toffano G, Ferraguti F. A correlation analysis of the regional distribution of central enkephalin and beta-endorphin immunoreactive terminals and of opiate receptors in adult and old male rats. Evidence for the existence of two main types of communication in the central nervous system: the volume transmission and the wiring transmission. *Acta Physiol Scand*. 1986 Oct;128(2):201-7.
6. Agnati LF, Leo G, Zanardi A, Genedani S, Rivera A, Fuxe K, et al. Volume transmission and wiring transmission from cellular to molecular networks: history and perspectives. *Acta Physiol (Oxf)*. 2006 May-Jun;187(1-2):329-44.
7. Fuxe K, Agnati L. Volume transmission in the brain. New York: Raven Press; 1991.
8. Fuxe K, Agnati L. Cell-Cell Communication through the Extracellular Space. In: Squire LR, editor. *Encyclopedia of Neuroscience* Oxford: Academic Press; 2009. p. 655-64.
9. Fuxe K, Agnati LF. Two Principle Modes of Electrochemical Communication in the Brain: Volume versus Wiring Transmission. In: Fuxe K, Agnati LF, editors. *Volume Transmission in the Brain: Novel Mechanisms of Neuronal Transmission*. New York: Raven Press; 1991a. p. 1-9.

10. Fuxe K, Dahlstrom A, Hoistad M, Marcellino D, Jansson A, Rivera A, et al. From the Golgi-Cajal mapping to the transmitter-based characterization of the neuronal networks leading to two modes of brain communication: Wiring and volume transmission. *Brain Res Rev.* 2007 Aug;55(1):17-54.
11. Fuxe K, Dahlstrom A, Jonsson G, Marcellino D, Guescini M, Dam M, et al. The discovery of Central Monoamine Neurons Gave Volume Transmission to the Wired Brain. *Prog Neurobiol.* 2009 Oct 20.
12. Agnati LF, Fuxe K. Volume transmission as a key feature of information handling in the central nervous system possible new interpretative value of the Turing's B-type machine. *Progress in Brain Research.* 2000;125:3-19.
13. Jansson A, Descarries L, Cornea-Hebert V, Riad M, Verge D, Bancila M, et al. Transmitter-Receptor mismatches in central dopamine serotonin and neuropeptide systems. In: Walz W, editor. *The Neuronal Environment: Brain Homeostasis in Health and Disease.* Totowa, NJ: Humana Press; 2002. p. 83-107.
14. Rice ME, Cragg SJ. Dopamine spillover after quantal release: rethinking dopamine transmission in the nigrostriatal pathway. *Brain Res Rev.* 2008 Aug;58(2):303-13.
15. Fuxe K, Marcellino D, Rivera A, Diaz-Cabiale Z, Filip M, Gago B, et al. Receptor-receptor interactions within receptor mosaics. Impact on neuropsychopharmacology. *Brain Res Rev.* 2008a Aug;58(2):415-52.
16. Agnati LF, Guidolin D, Leo G, Carone C, Genedani S, Fuxe K. Receptor-receptor interactions: a novel concept in brain integration. *Prog Neurobiol.* 2009 Oct 19.
17. Fuxe K, Marcellino D, Guidolin D, Woods AS, Agnati LF. Brain receptor mosaics and their intramembrane receptor-receptor interactions: molecular integration in transmission and novel target for drug development *J Acupunct Meridian Stud.* 2009;2:1-25.
18. Ferre S, Fuxe K. Adenosine as a volume transmission signal. A feedback detector of neuronal activation. *Prog Brain Res.* 2000;125:353-61.

19. Fredholm BB. Astra Award Lecture. Adenosine, adenosine receptors and the actions of caffeine. *Pharmacol Toxicol.* 1995 Feb;76(2):93-101.
20. Fredholm BB, Battig K, Holmen J, Nehlig A, Zvartau EE. Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev.* 1999 Mar;51(1):83-133.
21. Agnati LF, Ferre S, Lluís C, Franco R, Fuxe K. Molecular mechanisms and therapeutical implications of intramembrane receptor/receptor interactions among heptahelical receptors with examples from the striatopallidal GABA neurons. *Pharmacol Rev.* 2003 Sep;55(3):509-50.
22. Ferre S, Fredholm BB, Morelli M, Popoli P, Fuxe K. Adenosine-dopamine receptor-receptor interactions as an integrative mechanism in the basal ganglia. *Trends Neurosci.* 1997 Oct;20(10):482-7.
23. Fuxe K, Agnati LF, Jacobsen K, Hillion J, Canals M, Torvinen M, et al. Receptor heteromerization in adenosine A2A receptor signaling: relevance for striatal function and Parkinson's disease. *Neurology.* 2003 Dec 9;61(11 Suppl 6):S19-23.
24. Fuxe K, Ferre S, Genedani S, Franco R, Agnati LF. Adenosine receptor-dopamine receptor interactions in the basal ganglia and their relevance for brain function. *Physiol Behav.* 2007 Sep;92(1-2):210-7.
25. Fuxe K, Ferre S, Zoli M, Agnati LF. Integrated events in central dopamine transmission as analyzed at multiple levels. Evidence for intramembrane adenosine A2A/dopamine D2 and adenosine A1/dopamine D1 receptor interactions in the basal ganglia. *Brain Res Brain Res Rev.* 1998 May;26(2-3):258-73.
26. Agnati LF, Fuxe K, Zini I, Lenzi P, Hokfelt T. Aspects on receptor regulation and isoreceptor identification. *Med Biol.* 1980 Aug;58(4):182-7.
27. Fuxe K, Agnati LF, Benfenati F, Cimmino M, Algeri S, Hokfelt T, et al. Modulation by cholecystokinins of 3H-spiroperidol binding in rat striatum: evidence for increased affinity and reduction in the number of binding sites. *Acta Physiol Scand.* 1981 Dec;113(4):567-9.
28. Agnati LF, Fuxe K, Zoli M, Pich EM, Benfenati F, Zini I, et al. Aspects on the information handling by the central nervous system: focus on cotransmission in the aged rat brain. *Prog Brain Res.* 1986;68:291-301.

29. Fuxe K, Agnati LF, Benfenati F, Celani M, Zini I, Zoli M, et al. Evidence for the existence of receptor-receptor interactions in the central nervous system. Studies on the regulation of monoamine receptors by neuropeptides. *J Neural Transm Suppl.* 1983;18:165-79.
30. Fuxe K, Agnati LF, Harfstrand A, Andersson K, Mascagni F, Zoli M, et al. Studies on peptide comodulator transmission. New perspective on the treatment of disorders of the central nervous system. *Prog Brain Res.* 1986;66:341-68.
31. Agnati LF, Tarakanov AO, Ferre S, Fuxe K, Guidolin D. Receptor-receptor interactions, receptor mosaics, and basic principles of molecular network organization: possible implications for drug development. *J Mol Neurosci.* 2005;26(2-3):193-208.
32. Angers S, Salahpour A, Bouvier M. Dimerization: an emerging concept for G protein-coupled receptor ontogeny and function. *Annu Rev Pharmacol Toxicol.* 2002;42:409-35.
33. Bulenger S, Marullo S, Bouvier M. Emerging role of homo- and heterodimerization in G-protein-coupled receptor biosynthesis and maturation. *Trends Pharmacol Sci.* 2005 Mar;26(3):131-7.
34. Franco R, Ferre S, Agnati L, Torvinen M, Gines S, Hillion J, et al. Evidence for adenosine/dopamine receptor interactions: indications for heteromerization. *Neuropsychopharmacology.* 2000 Oct;23(4 Suppl):S50-9.
35. Fuxe K, Canals M, Torvinen M, Marcellino D, Terasmaa A, Genedani S, et al. Intramembrane receptor-receptor interactions: a novel principle in molecular medicine. *J Neural Transm.* 2007 Jan;114(1):49-75.
36. Milligan G. G protein-coupled receptor dimerization: function and ligand pharmacology. *Mol Pharmacol.* 2004 Jul;66(1):1-7.
37. Prinster SC, Hague C, Hall RA. Heterodimerization of g protein-coupled receptors: specificity and functional significance. *Pharmacol Rev.* 2005 Sep;57(3):289-98.
38. Agnati LF, Franzen O, Ferre S, Leo G, Franco R, Fuxe K. Possible role of intramembrane receptor-receptor interactions in memory and learning via formation of long-lived heteromeric complexes: focus on motor learning in the basal ganglia. *J Neural Transm Suppl.* 2003(65):1-28.

39. Agnati LF, Fuxe K, Zoli M, Rondanini C, Ogren SO. New vistas on synaptic plasticity: the receptor mosaic hypothesis of the engram. *Med Biol.* 1982 Aug;60(4):183-90.
40. Kenakin TP. Allosteric agonist modulators. *J ReceptSignal Transd.* 2007;27:247-59.
41. Kenakin TP. Seven transmembrane receptors as nature's prototype allosteric protein: de-emphasizing the geography of binding. *Mol Pharmacol.* 2008 Sep;74(3):541-3.
42. Kenakin TP. 7TM receptor allostery: putting numbers to shapeshifting proteins. *Trends Pharmacol Sci.* 2009 Sep;30(9):460-9.
43. Agnati LF, Guidolin D, Leo G, Fuxe K. A boolean network modelling of receptor mosaics relevance of topology and cooperativity. *J Neural Transm.* 2007 Jan;114(1):77-92.
44. Fuxe K, Agnati LF, von Euler G, Benfenati F, Zoli M, Härfstrand A, et al. Receptor-receptor interactions and development of psychoactive drugs. In: Costa E, editor. *Neurochemical pharmacology.* Washington: Raven Press, New York; 1989. p. 211-27.
45. Fuxe K, Marcellino D, Woods AS, Giuseppina L, Antonelli T, Ferraro L, et al. Integrated signaling in heterodimers and receptor mosaics of different types of GPCRs of the forebrain: relevance for schizophrenia. *J Neural Transm.* 2009 Aug;116(8):923-39.
46. Ferre S, Baler R, Bouvier M, Caron MG, Devi LA, Durroux T, et al. Building a new conceptual framework for receptor heteromers. *Nat Chem Biol.* 2009 Mar;5(3):131-4.
47. Agnati LF, Ferre S, Genedani S, Leo G, Guidolin D, Filaferro M, et al. Allosteric modulation of dopamine D2 receptors by homocysteine. *J Proteome Res.* 2006 Nov;5(11):3077-83.
48. Agnati LF, Leo G, Genedani S, Andreoli N, Marcellino D, Woods A, et al. Structural plasticity in G-protein coupled receptors as demonstrated by the allosteric actions of homocysteine and computer-assisted analysis of disordered domains. *Brain Res Rev.* 2008 Aug;58(2):459-74.
49. Milligan G, Smith NJ. Allosteric modulation of heterodimeric G-protein-coupled receptors. *Trends Pharmacol Sci.* 2007 Dec;28(12):615-20.

50. May LT, Leach K, Sexton PM, Christopoulos A. Allosteric modulation of G protein-coupled receptors. *Annu Rev Pharmacol Toxicol.* 2007;47:1-51.
51. Canals M, Marcellino D, Fanelli F, Ciruela F, de Benedetti P, Goldberg SR, et al. Adenosine A2A-dopamine D2 receptor-receptor heteromerization: qualitative and quantitative assessment by fluorescence and bioluminescence energy transfer. *J Biol Chem.* 2003 Nov 21;278(47):46741-9.
52. Ciruela F, Burgueno J, Casado V, Canals M, Marcellino D, Goldberg SR, et al. Combining mass spectrometry and pull-down techniques for the study of receptor heteromerization. Direct epitope-epitope electrostatic interactions between adenosine A2A and dopamine D2 receptors. *Anal Chem.* 2004 Sep 15;76(18):5354-63.
53. Hillion J, Canals M, Torvinen M, Casado V, Scott R, Terasmaa A, et al. Coaggregation, cointernalization, and codesensitization of adenosine A2A receptors and dopamine D2 receptors. *J Biol Chem.* 2002 May 17;277(20):18091-7.
54. Kamiya T, Saitoh O, Yoshioka K, Nakata H. Oligomerization of adenosine A2A and dopamine D2 receptors in living cells. *Biochem Biophys Res Commun.* 2003 Jun 27;306(2):544-9.
55. Fuxe K, Ferre S, Canals M, Torvinen M, Terasmaa A, Marcellino D, et al. Adenosine A2A and dopamine D2 heteromeric receptor complexes and their function. *J Mol Neurosci.* 2005;26(2-3):209-20.
56. Fenu S, Pinna A, Ongini E, Morelli M. Adenosine A2A receptor antagonism potentiates L-DOPA-induced turning behaviour and c-fos expression in 6-hydroxydopamine-lesioned rats. *Eur J Pharmacol.* 1997 Feb 26;321(2):143-7.
57. Ferre S, Ciruela F, Woods AS, Lluís C, Franco R. Functional relevance of neurotransmitter receptor heteromers in the central nervous system. *Trends Neurosci.* 2007 Aug 8.
58. Ferre S, O'Connor WT, Fuxe K, Ungerstedt U. The striopallidal neuron: a main locus for adenosine-dopamine interactions in the brain. *J Neurosci.* 1993 Dec;13(12):5402-6.

59. Ferre S, von Euler G, Johansson B, Fredholm BB, Fuxe K. Stimulation of high-affinity adenosine A₂ receptors decreases the affinity of dopamine D₂ receptors in rat striatal membranes. *Proc Natl Acad Sci U S A*. 1991 Aug 15;88(16):7238-41.
60. Fuxe K, Marcellino D, Genedani S, Agnati L. Adenosine A_{2A} receptors, dopamine D₂ receptors and their interactions in Parkinson's disease. *Mov Disord*. 2007 Jul 6.
61. Tanganelli S, Sandager Nielsen K, Ferraro L, Antonelli T, Kehr J, Franco R, et al. Striatal plasticity at the network level. Focus on adenosine A_{2A} and D₂ interactions in models of Parkinson's Disease. *Parkinsonism Relat Disord*. 2004 Jul;10(5):273-80.
62. Ferre S, Ciruela F, Canals M, Marcellino D, Burgueno J, Casado V, et al. Adenosine A_{2A}-dopamine D₂ receptor-receptor heteromers. Targets for neuro-psychiatric disorders. *Parkinsonism Relat Disord*. 2004 Jul;10(5):265-71.
63. Fuxe K, Ferre S, Snaprud P, von Euler G, Johansson B, Fredholm BB. Antagonistic A_{2A}/D₂ receptor interactions as a basis for adenosine/dopamine interactions in the central nervous system. *Drug Dev Res*. 1993;28:374-80.
64. Woods AS, Ciruela F, Fuxe K, Agnati LF, Lluís C, Franco R, et al. Role of electrostatic interaction in receptor-receptor heteromerization. *J Mol Neurosci*. 2005;26(2-3):125-32.
65. Torvinen M, Kozell LB, Neve KA, Agnati LF, Fuxe K. Biochemical identification of the dopamine D₂ receptor domains interacting with the adenosine A_{2A} receptor. *J Mol Neurosci*. 2004;24(2):173-80.
66. Fuxe K, Agnati LF, von Euler G, Tanganelli S, O'Connor WT, Ferre S, et al. Neuropeptides, excitatory amino acid and adenosine A₂ receptors regulate D₂ receptors via intramembrane receptor-receptor interactions. Relevance for Parkinson's disease and schizophrenia. *Neurochem Int*. 1992 Mar;20 Suppl:215S-24S.
67. Morelli M, Wardas J. Adenosine A_{2A} receptor antagonists: potential therapeutic and neuroprotective effects in Parkinson's disease. *Neurotox Res*. 2001 Nov;3(6):545-56.

68. Schwarzschild MA, Agnati L, Fuxe K, Chen JF, Morelli M. Targeting adenosine A_{2A} receptors in Parkinson's disease. *Trends Neurosci.* 2006 Nov;29(11):647-54.
69. Stromberg I, Popoli P, Muller CE, Ferre S, Fuxe K. Electrophysiological and behavioural evidence for an antagonistic modulatory role of adenosine A_{2A} receptors in dopamine D₂ receptor regulation in the rat dopamine-denervated striatum. *Eur J Neurosci.* 2000 Nov;12(11):4033-7.
70. Hernandez-Lopez S, Tkatch T, Perez-Garci E, Galarraga E, Bargas J, Hamm H, et al. D₂ dopamine receptors in striatal medium spiny neurons reduce L-type Ca²⁺ currents and excitability via a novel PLC[β]₁-IP₃-calcineurin-signaling cascade. *J Neurosci.* 2000 Dec 15;20(24):8987-95.
71. Surmeier DJ, Ding J, Day M, Wang Z, Shen W. D₁ and D₂ dopamine-receptor modulation of striatal glutamatergic signaling in striatal medium spiny neurons. *Trends Neurosci.* 2007 May;30(5):228-35.
72. Salim H, Ferre S, Dalal A, Peterfreund RA, Fuxe K, Vincent JD, et al. Activation of adenosine A₁ and A_{2A} receptors modulates dopamine D₂ receptor-induced responses in stably transfected human neuroblastoma cells. *J Neurochem.* 2000 Jan;74(1):432-9.
73. Yang SN, Dasgupta S, Lledo PM, Vincent JD, Fuxe K. Reduction of dopamine D₂ receptor transduction by activation of adenosine A_{2a} receptors in stably A_{2a}/D₂ (long-form) receptor co-transfected mouse fibroblast cell lines: studies on intracellular calcium levels. *Neuroscience.* 1995 Oct;68(3):729-36.
74. Beaulieu JM, Gainetdinov RR, Caron MG. The Akt-GSK-3 signaling cascade in the actions of dopamine. *Trends Pharmacol Sci.* 2007 Apr;28(4):166-72.
75. Kull B, Ferre S, Arslan G, Svenningsson P, Fuxe K, Owman C, et al. Reciprocal interactions between adenosine A_{2A} and dopamine D₂ receptors in Chinese hamster ovary cells co-transfected with the two receptors. *Biochem Pharmacol.* 1999 Sep 15;58(6):1035-45.
76. Calabrese VP, Lloyd KA, Brancazio P, Cefali E, Martin P, Wall J, Jr., et al. N-0923, a novel soluble dopamine D₂ agonist in the treatment of parkinsonism. *Mov Disord.* 1998 Sep;13(5):768-74.

77. Chase TN. Striatal plasticity and extrapyramidal motor dysfunction. *Parkinsonism Relat Disord*. 2004 Jul;10(5):305-13.
78. Chase TN, Bibbiani F, Bara-Jimenez W, Dimitrova T, Oh-Lee JD. Translating A2A antagonist KW6002 from animal models to parkinsonian patients. *Neurology*. 2003 Dec 9;61(11 Suppl 6):S107-11.
79. Hakansson K, Galdi S, Hendrick J, Snyder G, Greengard P, Fisone G. Regulation of phosphorylation of the GluR1 AMPA receptor by dopamine D2 receptors. *J Neurochem*. 2006 Jan;96(2):482-8.
80. Hakansson K, Lindskog M, Pozzi L, Usiello A, Fisone G. DARPP-32 and modulation of cAMP signaling: involvement in motor control and levodopa-induced dyskinesia. *Parkinsonism Relat Disord*. 2004 Jul;10(5):281-6.
81. Ferre S, Fuxe K. Dopamine denervation leads to an increase in the intramembrane interaction between adenosine A2 and dopamine D2 receptors in the neostriatum. *Brain Res*. 1992 Oct 23;594(1):124-30.
82. Ferre S, Popoli P, Gimenez-Llort L, Rimondini R, Muller CE, Stromberg I, et al. Adenosine/dopamine interaction: implications for the treatment of Parkinson's disease. *Parkinsonism Relat Disord*. 2001 Jul;7(3):235-41.
83. Yang JN, Bjorklund O, Lindstrom-Tornqvist K, Lindgren E, Eriksson TM, Kahlstrom J, et al. Mice heterozygous for both A1 and A(2A) adenosine receptor genes show similarities to mice given long-term caffeine. *J Appl Physiol*. 2009 Feb;106(2):631-9.
84. Kanda T, Jackson MJ, Smith LA, Pearce RK, Nakamura J, Kase H, et al. Adenosine A2A antagonist: a novel antiparkinsonian agent that does not provoke dyskinesia in parkinsonian monkeys. *Ann Neurol*. 1998 Apr;43(4):507-13.
85. Pinna A, Fenu S, Morelli M. Motor stimulant effects of the adenosine A2A receptor antagonist SCH 58261 do not develop tolerance after repeated treatments in 6-hydroxydopamine-lesioned rats. *Synapse*. 2001 Mar 1;39(3):233-8.
86. Jenner P. Istradefylline, a novel adenosine A2A receptor antagonist, for the treatment of Parkinson's disease. *Expert Opin Investig Drugs*. 2005 Jun;14(6):729-38.

87. Bara-Jimenez W, Sherzai A, Dimitrova T, Favit A, Bibbiani F, Gillespie M, et al. Adenosine A(2A) receptor antagonist treatment of Parkinson's disease. *Neurology*. 2003 Aug 12;61(3):293-6.
88. Hauser RA, Hubble JP, Truong DD. Randomized trial of the adenosine A(2A) receptor antagonist istradefylline in advanced PD. *Neurology*. 2003 Aug 12;61(3):297-303.
89. Antonelli T, Fuxe K, Agnati L, Mazzoni E, Tanganelli S, Tomasini MC, et al. Experimental studies and theoretical aspects on A2A/D2 receptor interactions in a model of Parkinson's disease. Relevance for L-dopa induced dyskinesias. *J Neurol Sci*. 2006 Oct 25;248(1-2):16-22.
90. Carta AR, Kachroo A, Schintu N, Xu K, Schwarzschild MA, Wardas J, et al. Inactivation of neuronal forebrain A(2A) receptors protects dopaminergic neurons in a mouse model of Parkinson's disease. *J Neurochem*. 2009 Oct 8.
91. Ross GW, Abbott RD, Petrovitch H, Morens DM, Grandinetti A, Tung KH, et al. Association of coffee and caffeine intake with the risk of Parkinson disease. *Jama*. 2000 May 24-31;283(20):2674-9.
92. Ascherio A, Zhang SM, Hernan MA, Kawachi I, Colditz GA, Speizer FE, et al. Prospective study of caffeine consumption and risk of Parkinson's disease in men and women. *Ann Neurol*. 2001 Jul;50(1):56-63.
93. Ferre S, O'Connor WT, Snaprud P, Ungerstedt U, Fuxe K. Antagonistic interaction between adenosine A2A receptors and dopamine D2 receptors in the ventral striopallidal system. Implications for the treatment of schizophrenia. *Neuroscience*. 1994 Dec;63(3):765-73.
94. Beaulieu JM, Gainetdinov RR, Caron MG. Akt/GSK3 signaling in the action of psychotropic drugs. *Annu Rev Pharmacol Toxicol*. 2009;49:327-47.
95. Masri B, Salahpour A, Didriksen M, Ghisi V, Beaulieu JM, Gainetdinov RR, et al. Antagonism of dopamine D2 receptor/beta-arrestin 2 interaction is a common property of clinically effective antipsychotics. *Proc Natl Acad Sci U S A*. 2008 Sep 9;105(36):13656-61.
96. Tikh EI, Fenton RA, Chen JF, Schwarzschild MA, Dobson JG, Jr. Adenosine A1 and A2A receptor regulation of protein phosphatase 2A in the murine heart. *J Cell Physiol*. 2008 Jul;216(1):83-90.

97. Anden NE, Butcher SG, Corrodi H, Fuxe K, Ungerstedt U. Receptor activity and turnover of dopamine and noradrenaline after neuroleptics. *Eur J Pharmacol.* 1970;11(3):303-14.
98. Anden NE, Dahlstrom A, Fuxe K, Larsson K. Functional role of the nigro-neostriatal dopamine neurons. *Acta Pharmacol Toxicol (Copenh).* 1966;24(2):263-74.
99. Carlsson A, Lindqvist M. Effect Of Chlorpromazine Or Haloperidol On Formation Of 3methoxytyramine And Normetanephrine In Mouse Brain. *Acta Pharmacol Toxicol (Copenh).* 1963;20:140-4.
100. Seeman P. Dopamine receptors and the dopamine hypothesis of schizophrenia. *Synapse.* 1987;1(2):133-52.
101. Seeman P, Schwarz J, Chen JF, Szechtman H, Perreault M, McKnight GS, et al. Psychosis pathways converge via D2high dopamine receptors. *Synapse.* 2006 Sep 15;60(4):319-46.
102. Ogren SO, Rosen L, Fuxe K. The dopamine D2 antagonist remoxipride acts in vivo on a subpopulation of dopamine D2 receptors. *Neuroscience.* 1994 Jul;61(2):269-83.
103. Svensson TH. Dysfunctional brain dopamine systems induced by psychotomimetic NMDA-receptor antagonists and the effects of antipsychotic drugs. *Brain Res Brain Res Rev.* 2000 Mar;31(2-3):320-9.
104. Fuxe K, editor. Biological and pharmacological theories.discussion. The neuroleptics; 1970. S.Karger,Basel.
105. Diaz-Cabiale Z, Hurd Y, Guidolin D, Finnman UB, Zoli M, Agnati LF, et al. Adenosine A2A agonist CGS 21680 decreases the affinity of dopamine D2 receptors for dopamine in human striatum. *Neuroreport.* 2001 Jul 3;12(9):1831-4.
106. Rimondini R, Ferre S, Ogren SO, Fuxe K. Adenosine A2A agonists: a potential new type of atypical antipsychotic. *Neuropsychopharmacology.* 1997 Aug;17(2):82-91.
107. Andersen MB, Fuxe K, Werge T, Gerlach J. The adenosine A2A receptor agonist CGS 21680 exhibits antipsychotic-like activity in *Cebus apella* monkeys. *Behav Pharmacol.* 2002 Dec;13(8):639-44.

108. Groenewegen HJ. Organization of the afferent connections of the mediodorsal thalamic nucleus in the rat, related to the mediodorsal-prefrontal topography. *Neuroscience*. 1988 Feb;24(2):379-431.
109. Popken GJ, Bunney WE, Jr., Potkin SG, Jones EG. Subnucleus-specific loss of neurons in medial thalamus of schizophrenics. *Proc Natl Acad Sci U S A*. 2000 Aug 1;97(16):9276-80.
110. Popoli P, Betto P, Reggio R, Ricciarello G. Adenosine A2A receptor stimulation enhances striatal extracellular glutamate levels in rats. *Eur J Pharmacol*. 1995 Dec 12;287(2):215-7.
111. Marcellino D, Roberts DC, Navarro G, Filip M, Agnati L, Lluís C, et al. Increase in A2A receptors in the nucleus accumbens after extended cocaine self-administration and its disappearance after cocaine withdrawal. *Brain Res*. 2007 Apr 27;1143:208-20.
112. Weerts EM, Griffiths RR. The adenosine receptor antagonist CGS15943 reinstates cocaine-seeking behavior and maintains self-administration in baboons. *Psychopharmacology (Berl)*. 2003 Jul;168(1-2):155-63.
113. Knapp CM, Foye MM, Cottam N, Ciraulo DA, Kornetsky C. Adenosine agonists CGS 21680 and NECA inhibit the initiation of cocaine self-administration. *Pharmacol Biochem Behav*. 2001 Apr;68(4):797-803.
114. Filip M, Frankowska M, Zaniwska M, Przegalinski E, Müller CE, Agnati L, et al. Involvement of adenosine A2A and dopamine receptors in the locomotor and sensitizing effects of cocaine. *Brain Res*. 2006 Mar 10;1077(1):67-80.
115. Morgan D, Brebner K, Lynch WJ, Roberts DC. Increases in the reinforcing efficacy of cocaine after particular histories of reinforcement. *Behav Pharmacol*. 2002 Sep;13(5-6):389-96.
116. Soria G, Castane A, Ledent C, Parmentier M, Maldonado R, Valverde O. The lack of A2A adenosine receptors diminishes the reinforcing efficacy of cocaine. *Neuropsychopharmacology*. 2006 May;31(5):978-87.
117. Chiang MC, Lee YC, Huang CL, Chern Y. cAMP-response element-binding protein contributes to suppression of the A2A adenosine receptor promoter by mutant Huntingtin with expanded polyglutamine residues. *J Biol Chem*. 2005 Apr 8;280(14):14331-40.

118. Carlezon WA, Jr., Thome J, Olson VG, Lane-Ladd SB, Brodtkin ES, Hiroi N, et al. Regulation of cocaine reward by CREB. *Science*. 1998 Dec 18;282(5397):2272-5.
119. Mattson BJ, Bossert JM, Simmons DE, Nozaki N, Nagarkar D, Kreuter JD, et al. Cocaine-induced CREB phosphorylation in nucleus accumbens of cocaine-sensitized rats is enabled by enhanced activation of extracellular signal-related kinase, but not protein kinase A. *J Neurochem*. 2005 Dec;95(5):1481-94.
120. Schiffmann SN, Jacobs O, Vanderhaeghen JJ. Striatal restricted adenosine A2 receptor (RDC8) is expressed by enkephalin but not by substance P neurons: an in situ hybridization histochemistry study. *J Neurochem*. 1991 Sep;57(3):1062-7.
121. Schiffmann SN, Libert F, Vassart G, Vanderhaeghen JJ. Distribution of adenosine A2 receptor mRNA in the human brain. *Neurosci Lett*. 1991 Sep 16;130(2):177-81.
122. Schiffmann SN, Vanderhaeghen JJ. Adenosine A2 receptors regulate the gene expression of striatopallidal and striatonigral neurons. *J Neurosci*. 1993 Mar;13(3):1080-7.
123. Watts VJ. Molecular mechanisms for heterologous sensitization of adenylyl cyclase. *J Pharmacol Exp Ther*. 2002 Jul;302(1):1-7.
124. Vortherms TA, Nguyen CH, Bastepe M, Juppner H, Watts VJ. D2 dopamine receptor-induced sensitization of adenylyl cyclase type 1 is G α (s) independent. *Neuropharmacology*. 2006 Apr;50(5):576-84.
125. Vortherms TA, Watts VJ. Sensitization of neuronal A2A adenosine receptors after persistent D2 dopamine receptor activation. *J Pharmacol Exp Ther*. 2004 Jan;308(1):221-7.
126. Torvinen M, Marcellino D, Canals M, Agnati LF, Lluís C, Franco R, et al. Adenosine A2A receptor and dopamine D3 receptor interactions: evidence of functional A2A/D3 heteromeric complexes. *Mol Pharmacol*. 2005 Feb;67(2):400-7.
127. Nimchinsky EA, Hof PR, Janssen WG, Morrison JH, Schmauss C. Expression of dopamine D3 receptor dimers and tetramers in brain and in transfected cells. *J Biol Chem*. 1997 Nov 14;272(46):29229-37.

128. Rivera A, Cuellar B, Giron FJ, Grandy DK, de la Calle A, Moratalla R. Dopamine D4 receptors are heterogeneously distributed in the striosomes/matrix compartments of the striatum. *J Neurochem.* 2002 Jan;80(2):219-29.
129. Gerfen CR. The neostriatal mosaic: multiple levels of compartmental organization in the basal ganglia. *Annu Rev Neurosci.* 1992;15:285-320.
130. Graybiel AM. Neurotransmitters and neuromodulators in the basal ganglia. *Trends Neurosci.* 1990 Jul;13(7):244-54.
131. Joyce JN, Millan MJ. Dopamine D3 receptor antagonists as therapeutic agents. *Drug Discov Today.* 2005 Jul 1;10(13):917-25.
132. Schwartz JC, Diaz J, Pilon C, Sokoloff P. Possible implications of the dopamine D(3) receptor in schizophrenia and in antipsychotic drug actions. *Brain Res Brain Res Rev.* 2000 Mar;31(2-3):277-87.
133. Tran-Nguyen LT, Fuchs RA, Coffey GP, Baker DA, O'Dell LE, Neisewander JL. Time-dependent changes in cocaine-seeking behavior and extracellular dopamine levels in the amygdala during cocaine withdrawal. *Neuropsychopharmacology.* 1998 Jul;19(1):48-59.
134. Neisewander JL, Fuchs RA, Tran-Nguyen LT, Weber SM, Coffey GP, Joyce JN. Increases in dopamine D3 receptor binding in rats receiving a cocaine challenge at various time points after cocaine self-administration: implications for cocaine-seeking behavior. *Neuropsychopharmacology.* 2004 Aug;29(8):1479-87.
135. Sokoloff P, Le Foll B, Perachon S, Bordet R, Ridray S, Schwartz JC. The dopamine D3 receptor and drug addiction. *Neurotox Res.* 2001 Oct;3(5):433-41.
136. Vorel SR, Ashby CR, Jr., Paul M, Liu X, Hayes R, Hagan JJ, et al. Dopamine D3 receptor antagonism inhibits cocaine-seeking and cocaine-enhanced brain reward in rats. *J Neurosci.* 2002 Nov 1;22(21):9595-603.
137. Segal DM, Moraes CT, Mash DC. Up-regulation of D3 dopamine receptor mRNA in the nucleus accumbens of human cocaine fatalities. *Brain Res Mol Brain Res.* 1997 May;45(2):335-9.
138. Ferre S, Karcz-Kubicha M, Hope BT, Popoli P, Burgueno J, Gutierrez MA, et al. Synergistic interaction between adenosine A2A and glutamate

- mGlu5 receptors: implications for striatal neuronal function. *Proc Natl Acad Sci U S A*. 2002 Sep 3;99(18):11940-5.
139. Cabello N, Gandia J, Bertarelli DC, Watanabe M, Lluís C, Franco R, et al. Metabotropic glutamate type 5, dopamine D2 and adenosine A2a receptors form higher-order oligomers in living cells. *J Neurochem*. 2009 Jun;109(5):1497-507.
 140. Carriba P, Navarro G, Ciruela F, Ferre S, Casado V, Agnati L, et al. Detection of heteromerization of more than two proteins by sequential BRET-FRET. *Nat Methods*. 2008 Aug;5(8):727-33.
 141. Conn PJ, Battaglia G, Marino MJ, Nicoletti F. Metabotropic glutamate receptors in the basal ganglia motor circuit. *Nat Rev Neurosci*. 2005 Oct;6(10):787-98.
 142. Kachroo A, Orlando LR, Grandy DK, Chen JF, Young AB, Schwarzschild MA. Interactions between metabotropic glutamate 5 and adenosine A2A receptors in normal and parkinsonian mice. *J Neurosci*. 2005 Nov 9;25(45):10414-9.
 143. Coccorello R, Breyse N, Amalric M. Simultaneous blockade of adenosine A2A and metabotropic glutamate mGlu5 receptors increase their efficacy in reversing Parkinsonian deficits in rats. *Neuropsychopharmacology*. 2004 Aug;29(8):1451-61.
 144. Diaz-Cabiale Z, Vivo M, Del Arco A, O'Connor WT, Harte MK, Muller CE, et al. Metabotropic glutamate mGlu5 receptor-mediated modulation of the ventral striopallidal GABA pathway in rats. Interactions with adenosine A(2A) and dopamine D(2) receptors. *Neurosci Lett*. 2002 May 17;324(2):154-8.
 145. Kinney GG, Burno M, Campbell UC, Hernandez LM, Rodriguez D, Bristow LJ, et al. Metabotropic glutamate subtype 5 receptors modulate locomotor activity and sensorimotor gating in rodents. *J Pharmacol Exp Ther*. 2003 Jul;306(1):116-23.
 146. Fuxe K, Agnati L, Franco R, Roberts DC, Tanganelli S, Bader M, et al. Genomics and dopamine D2 receptor mediated molecular mechanisms of cannabinoid abuse and cocaine addiction. EU ADDTOMICS.proposal N 004863, LifeSciHealth-I,OJ 2003/C164,FP6-2003; 2003 Contract No.: Document Number].

147. Fuxe K, Ferre S, Woods A, Rivera A, Hoistad M, Franco R, et al., editors. Novel strategies for the treatment of Parkinson's Disease. Focus on receptor-receptor interactions in the basal ganglia. . Monitoring Molecules in Neuroscience; 2003; Stockholm. Karolinska University Press.
148. Kearn CS, Blake-Palmer K, Daniel E, Mackie K, Glass M. Concurrent stimulation of cannabinoid CB1 and dopamine D2 receptors enhances heterodimer formation: a mechanism for receptor cross-talk? *Mol Pharmacol.* 2005 May;67(5):1697-704.
149. Glass M, Felder CC. Concurrent stimulation of cannabinoid CB1 and dopamine D2 receptors augments cAMP accumulation in striatal neurons: evidence for a Gs linkage to the CB1 receptor. *J Neurosci.* 1997 Jul 15;17(14):5327-33.
150. Hermann H, Marsicano G, Lutz B. Coexpression of the cannabinoid receptor type 1 with dopamine and serotonin receptors in distinct neuronal subpopulations of the adult mouse forebrain. *Neuroscience.* 2002;109(3):451-60.
151. Julian MD, Martin AB, Cuellar B, Rodriguez De Fonseca F, Navarro M, Moratalla R, et al. Neuroanatomical relationship between type 1 cannabinoid receptors and dopaminergic systems in the rat basal ganglia. *Neuroscience.* 2003;119(1):309-18.
152. Pickel VM, Chan J, Kearn CS, Mackie K. Targeting dopamine D2 and cannabinoid-1 (CB1) receptors in rat nucleus accumbens. *J Comp Neurol.* 2006 Mar 20;495(3):299-313.
153. Uchigashima M, Narushima M, Fukaya M, Katona I, Kano M, Watanabe M. Subcellular arrangement of molecules for 2-arachidonoyl-glycerol-mediated retrograde signaling and its physiological contribution to synaptic modulation in the striatum. *J Neurosci.* 2007 Apr 4;27(14):3663-76.
154. Andersson M, Usiello A, Borgkvist A, Pozzi L, Dominguez C, Fienberg AA, et al. Cannabinoid action depends on phosphorylation of dopamine- and cAMP-regulated phosphoprotein of 32 kDa at the protein kinase A site in striatal projection neurons. *J Neurosci.* 2005 Sep 14;25(37):8432-8.

155. Beltramo M, de Fonseca FR, Navarro M, Calignano A, Gorriti MA, Grammatikopoulos G, et al. Reversal of dopamine D(2) receptor responses by an anandamide transport inhibitor. *J Neurosci*. 2000 May 1;20(9):3401-7.
156. Borgkvist A, Marcellino D, Fuxe K, Greengard P, Fisone G. Regulation of DARPP-32 phosphorylation by Delta(9)-tetrahydrocannabinol. *Neuropharmacology*. 2007 Jul 6.
157. Marcellino D, Carriba P, Filip M, Borgkvist A, Frankowska M, Bellido I, et al. Antagonistic cannabinoid CB1/dopamine D2 receptor interactions in striatal CB1/D2 heteromers. A combined neurochemical and behavioral analysis. *Neuropharmacology*. 2008 Apr;54(5):815-23.
158. Carriba P, Ortiz O, Patkar K, Justinova Z, Stroik J, Themann A, et al. Striatal Adenosine A(2A) and Cannabinoid CB(1) Receptors Form Functional Heteromeric Complexes that Mediate the Motor Effects of Cannabinoids. *Neuropsychopharmacology*. 2007 Mar 14.
159. Ferrer B, Gorriti MA, Palomino A, Gornemann I, de Diego Y, Bermudez-Silva FJ, et al. Cannabinoid CB1 receptor antagonism markedly increases dopamine receptor-mediated stereotypies. *Eur J Pharmacol*. 2007 Mar 22;559(2-3):180-3.
160. Lastres-Becker I, Cebeira M, de Ceballos ML, Zeng BY, Jenner P, Ramos JA, et al. Increased cannabinoid CB1 receptor binding and activation of GTP-binding proteins in the basal ganglia of patients with Parkinson's syndrome and of MPTP-treated marmosets. *Eur J Neurosci*. 2001 Dec;14(11):1827-32.
161. Piomelli D. The molecular logic of endocannabinoid signalling. *Nat Rev Neurosci*. 2003 Nov;4(11):873-84.
162. Andersson M, Terasmaa A, Fuxe K, Stromberg I. Subchronic haloperidol increases CB(1) receptor binding and G protein coupling in discrete regions of the basal ganglia. *J Neurosci Res*. 2005 Oct 15;82(2):264-72.
163. Hurley MJ, Mash DC, Jenner P. Expression of cannabinoid CB1 receptor mRNA in basal ganglia of normal and parkinsonian human brain. *J Neural Transm*. 2003 Nov;110(11):1279-88.
164. D'Souza DC, Abi-Saab WM, Madonick S, Forselius-Bielen K, Doersch A, Braley G, et al. Delta-9-tetrahydrocannabinol effects in schizophrenia:

- implications for cognition, psychosis, and addiction. *Biol Psychiatry*. 2005 Mar 15;57(6):594-608.
165. Leweke FM, Giuffrida A, Koethe D, Schreiber D, Nolden BM, Kranaster L, et al. Anandamide levels in cerebrospinal fluid of first-episode schizophrenic patients: impact of cannabis use. *Schizophr Res*. 2007 Aug;94(1-3):29-36.
 166. Vlachou S, Nomikos GG, Panagis G. WIN 55,212-2 decreases the reinforcing actions of cocaine through CB1 cannabinoid receptor stimulation. *Behav Brain Res*. 2003 May 15;141(2):215-22.
 167. Fattore L, Martellotta MC, Cossu G, Mascia MS, Fratta W. CB1 cannabinoid receptor agonist WIN 55,212-2 decreases intravenous cocaine self-administration in rats. *Behav Brain Res*. 1999 Oct;104(1-2):141-6.
 168. Gines S, Hillion J, Torvinen M, Le Crom S, Casado V, Canela EI, et al. Dopamine D1 and adenosine A1 receptors form functionally interacting heteromeric complexes. *Proc Natl Acad Sci U S A*. 2000 Jul 18;97(15):8606-11.
 169. Toda S, Alguacil LF, Kalivas PW. Repeated cocaine administration changes the function and subcellular distribution of adenosine A1 receptor in the rat nucleus accumbens. *J Neurochem*. 2003 Dec;87(6):1478-84.
 170. Franco R, Lluís C, Canela EI, Mallol J, Agnati L, Casado V, et al. Receptor-receptor interactions involving adenosine A1 or dopamine D1 receptors and accessory proteins. *J Neural Transm*. 2007 Jan;114(1):93-104.
 171. Ciruela F, Casado V, Mallol J, Canela EI, Lluís C, Franco R. Immunological identification of A1 adenosine receptors in brain cortex. *J Neurosci Res*. 1995 Dec 15;42(6):818-28.
 172. George SR, Lee SP, Varghese G, Zeman PR, Seeman P, Ng GY, et al. A transmembrane domain-derived peptide inhibits D1 dopamine receptor function without affecting receptor oligomerization. *J Biol Chem*. 1998 Nov 13;273(46):30244-8.

173. Ferre S, Popoli P, Gimenez-Llort L, Finnman UB, Martinez E, Scotti de Carolis A, et al. Postsynaptic antagonistic interaction between adenosine A1 and dopamine D1 receptors. *Neuroreport*. 1994 Dec 30;6(1):73-6.
174. Ferre S, Torvinen M, Antoniou K, Irenius E, Civelli O, Arenas E, et al. Adenosine A1 receptor-mediated modulation of dopamine D1 receptors in stably cotransfected fibroblast cells. *J Biol Chem*. 1998 Feb 20;273(8):4718-24.
175. Franco R, Casado V, Ciruela F, Saura C, Mallol J, Canela EI, et al. Cell surface adenosine deaminase: much more than an ectoenzyme. *Prog Neurobiol*. 1997 Jul;52(4):283-94.
176. Ciruela F, Saura C, Canela EI, Mallol J, Lluís C, Franco R. Adenosine deaminase affects ligand-induced signalling by interacting with cell surface adenosine receptors. *FEBS Lett*. 1996 Feb 19;380(3):219-23.
177. Torvinen M, Gines S, Hillion J, Latini S, Canals M, Ciruela F, et al. Interactions among adenosine deaminase, adenosine A(1) receptors and dopamine D(1) receptors in stably cotransfected fibroblast cells and neurons. *Neuroscience*. 2002;113(3):709-19.
178. Ferre S, Popoli P, Tinner-Staines B, Fuxe K. Adenosine A1 receptor-dopamine D1 receptor interaction in the rat limbic system: modulation of dopamine D1 receptor antagonist binding sites. *Neurosci Lett*. 1996 Apr 19;208(2):109-12.
179. Popoli P, Gimenez-Llort L, Pezzola A, Reggio R, Martinez E, Fuxe K, et al. Adenosine A1 receptor blockade selectively potentiates the motor effects induced by dopamine D1 receptor stimulation in rodents. *Neurosci Lett*. 1996 Nov 8;218(3):209-13.
180. Rimondini R, Ferre S, Gimenez-Llort L, Ogren SO, Fuxe K. Differential effects of selective adenosine A1 and A2A receptor agonists on dopamine receptor agonist-induced behavioural responses in rats. *Eur J Pharmacol*. 1998 Apr 24;347(2-3):153-8.
181. Ferre S, O'Connor WT, Svenningsson P, Bjorklund L, Lindberg J, Tinner B, et al. Dopamine D1 receptor-mediated facilitation of GABAergic neurotransmission in the rat strioentopeduncular pathway and its modulation by adenosine A1 receptor-mediated mechanisms. *Eur J Neurosci*. 1996 Jul;8(7):1545-53.

182. Popoli P, Ferre S, Pezzola A, Reggio R, Scotti de Carolis A, Fuxe K. Stimulation of adenosine A1 receptors prevents the EEG arousal due to dopamine D1 receptor activation in rabbits. *Eur J Pharmacol.* 1996 Jun 3;305(1-3):123-6.
183. Marcellino D, Ferre S, Casado V, Cortes A, Le Foll B, Mazzola C, et al. Identification of dopamine D1-D3 receptor heteromers. Indications for a role of synergistic D1-D3 receptor interactions in the striatum. *J Biol Chem.* 2008 Sep 19;283(38):26016-25.
184. Bezard E, Ferry S, Mach U, Stark H, Leriche L, Boraud T, et al. Attenuation of levodopa-induced dyskinesia by normalizing dopamine D3 receptor function. *Nat Med.* 2003 Jun;9(6):762-7.
185. Bordet R, Ridray S, Carboni S, Diaz J, Sokoloff P, Schwartz JC. Induction of dopamine D3 receptor expression as a mechanism of behavioral sensitization to levodopa. *Proc Natl Acad Sci U S A.* 1997 Apr 1;94(7):3363-7.
186. Schwartz JC, Diaz J, Bordet R, Griffon N, Perachon S, Pilon C, et al. Functional implications of multiple dopamine receptor subtypes: the D1/D3 receptor coexistence. *Brain Res Brain Res Rev.* 1998 May;26(2-3):236-42.
187. Le Foll B, Frances H, Diaz J, Schwartz JC, Sokoloff P. Role of the dopamine D3 receptor in reactivity to cocaine-associated cues in mice. *Eur J Neurosci.* 2002 Jun;15(12):2016-26.
188. Staley JK, Mash DC. Adaptive increase in D3 dopamine receptors in the brain reward circuits of human cocaine fatalities. *J Neurosci.* 1996 Oct 1;16(19):6100-6.
189. Hummel M, Unterwald EM. D1 dopamine receptor: a putative neurochemical and behavioral link to cocaine action. *J Cell Physiol.* 2002 Apr;191(1):17-27.
190. Surmeier DJ, Song WJ, Yan Z. Coordinated expression of dopamine receptors in neostriatal medium spiny neurons. *J Neurosci.* 1996 Oct 15;16(20):6579-91.
191. Fuxe K. Dopamine receptor agonists in brain research and as therapeutic agents. *Trends in Neurosciences.* 1979;2:1-4.

192. Quarta D, Ciruela F, Patkar K, Borycz J, Solinas M, Lluís C, et al. Heteromeric nicotinic acetylcholine-dopamine autoreceptor complexes modulate striatal dopamine release. *Neuropsychopharmacology*. 2007 Jan;32(1):35-42.
193. Li XM, Zoli M, Finnman UB, Le Novère N, Changeux JP, Fuxe K. A single (-)-nicotine injection causes change with a time delay in the affinity of striatal D2 receptors for antagonist, but not for agonist, nor in the D2 receptor mRNA levels in the rat substantia nigra. *Brain Res*. 1995 May 8;679(1):157-67.
194. Lee FJ, Pei L, Liu F. Disruption of the dopamine transporter-dopamine D2 receptor interaction in schizophrenia. *Synapse*. 2009 Aug;63(8):710-2.
195. Gong S, Zheng C, Doughty ML, Losos K, Didkovsky N, Schambra UB, et al. A gene expression atlas of the central nervous system based on bacterial artificial chromosomes. *Nature*. 2003 Oct 30;425(6961):917-25.

Figure 1- Fuxe et al.

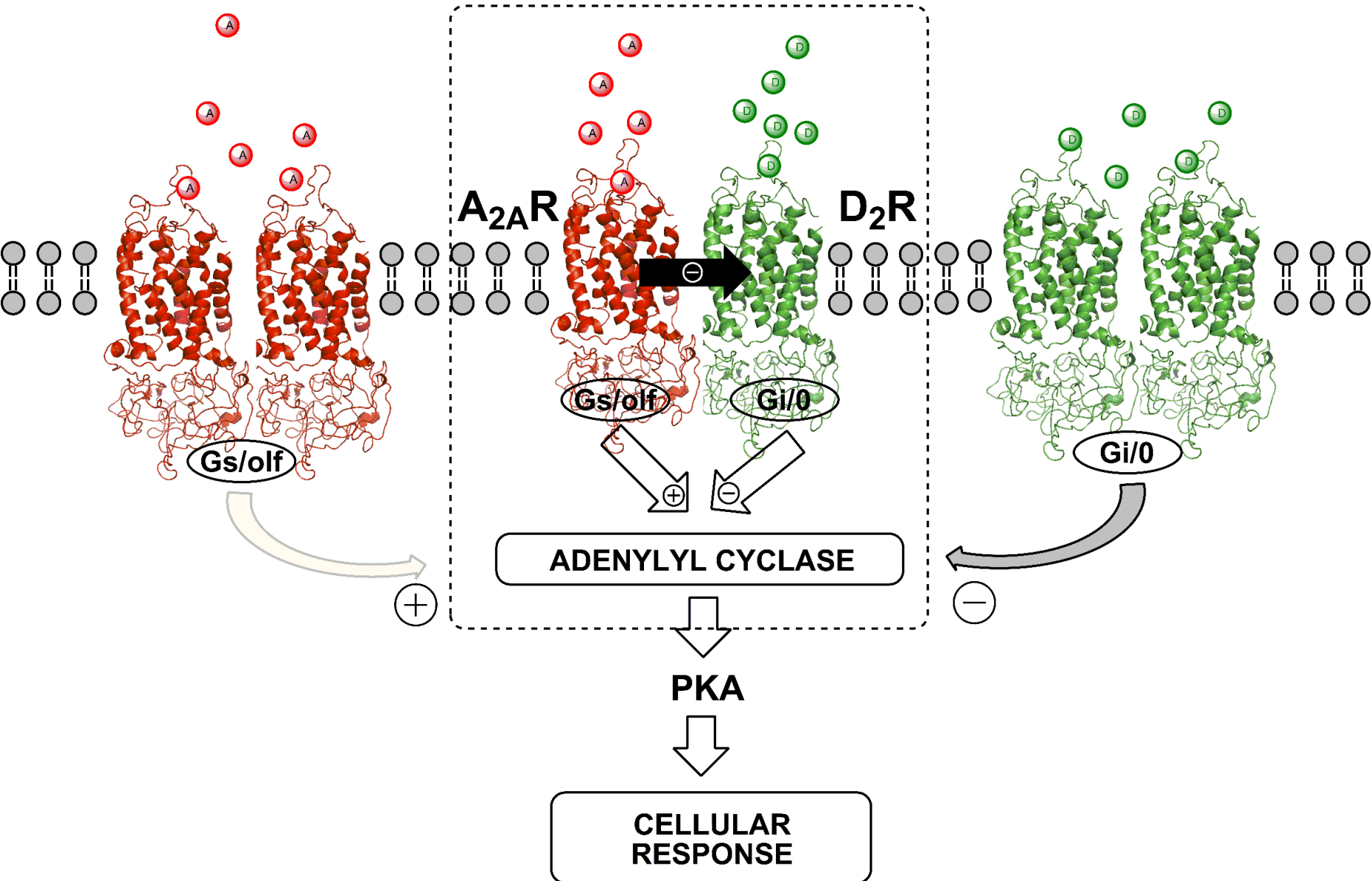
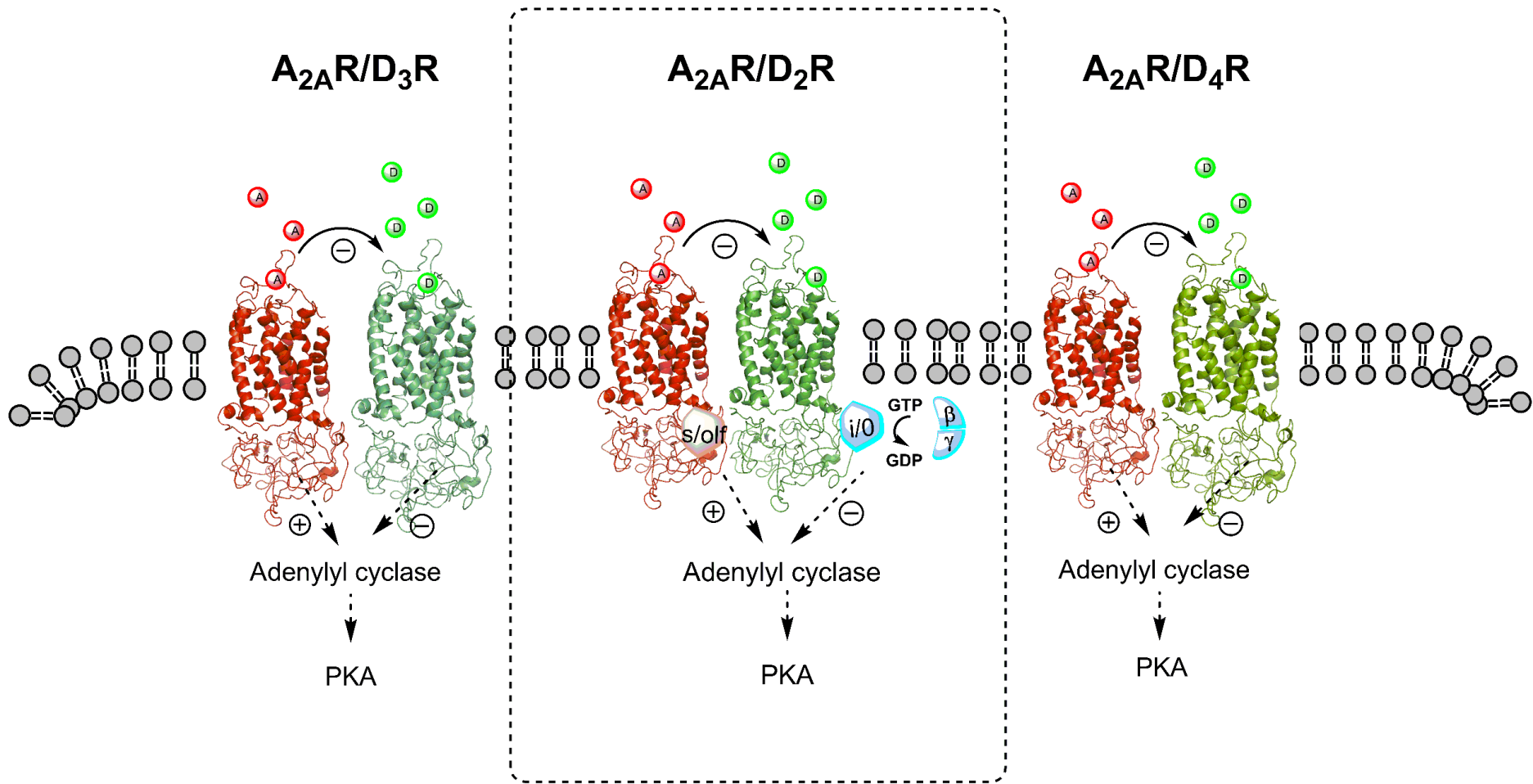


Figure 2- Fuxe et al.



D₃R-IC3: 216-**KQRRRKRI**-223

D₂R-IC3: 215-**VLRRRRKRVN**-224

D₄R-IC3: 377-**TRRRRRRAKI**-385

A_{2A}R-CT: 370-**SAQEpSQGNT**-378

A_{2A}R-CT: 388-**HELKGVCPPEPGLDDPLAQDGAGVS**-412

Figure 3- Fuxe et al.

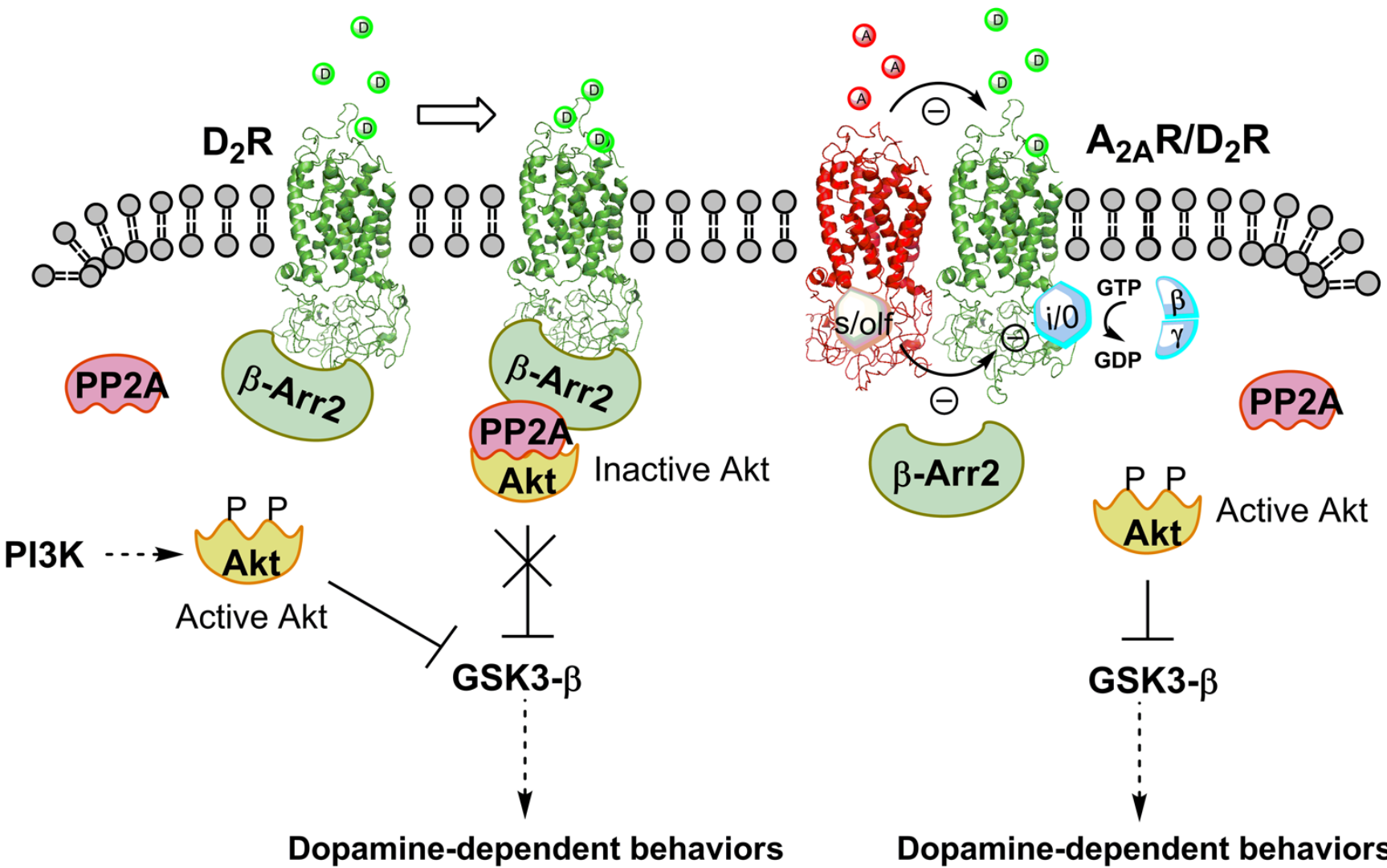


Figure 4- Fuxe et al

