



## Skull shape evolution in durophagous carnivorans

Journal:	<i>Evolution</i>
Manuscript ID:	12-0796.R1
Manuscript Type:	Original Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Figueirido, Borja; Facultad de Ciencias, Universidad de Málaga, Departamento de Ecología y Geología, Área de Paleontología Tseng, Zhijie; Natural History Museum of Los Angeles County, Department of Vertebrate Paleontology Martín-Serra, Alberto; Facultad de Ciencias, Universidad de Málaga, Ecología y Geología, Área de Paleontología
Keywords:	Evolutionary convergence, Constraint, Phenotype, Morphometrics, Durophagy, Carnivora

## Skull shape evolution in durophagous carnivorans

Borja Figueirido<sup>1,2\*</sup>, Zhijie Jack Tseng<sup>3</sup> and Alberto Martín-Serra<sup>2</sup>

<sup>1</sup> Department of Ecology and Evolutionary Biology, Brown University, Providence, RI 02912, U.S.A. E-mail: [Francisco\\_Figueirido\\_Castillo@brown.edu](mailto:Francisco_Figueirido_Castillo@brown.edu).

<sup>2</sup> Departamento de Ecología y Geología de la Facultad de Ciencias, Universidad de Málaga, 29071-Málaga, Spain. E-mail: [Borja.figueirido@uma.es](mailto:Borja.figueirido@uma.es); [almarse@uma.es](mailto:almarse@uma.es)

<sup>3</sup> Department of Vertebrate Paleontology, Natural History Museum of Los Angeles County, 900 Exposition Boulevard, Los Angeles, California 90007, USA. E-mail: [jack.tseng@alumni.usc.edu](mailto:jack.tseng@alumni.usc.edu)

**Short-title for page headings:** Evolution in durophages

*\*Author for correspondence*

Borja Figueirido

Departamento de Ecología y Geología de la Facultad de Ciencias de la Universidad de Málaga, 29071-Málaga, Spain

E-mail: [Borja.figueirido@uma.es](mailto:Borja.figueirido@uma.es)

Telephone: +34 952 13 18 56

1  
2  
3 In this paper we investigate convergent evolution towards durophagy in carnivoran skull  
4 shape using geometric morphometrics in a sample of living and extinct species.  
5  
6

7 Principal components analysis indicate that, in spite of the different dietary resources  
8 consumed by durophages –i.e., bone-crackers and bamboo-feeders– both groups of  
9 carnivorans share portions of skull phenotypic spaces. We identify by discriminant  
10 analyses a shared set of adaptations towards durophagy in the skull of carnivores.  
11  
12  
13

14 However, ancestral states indicate that although durophages reached similar phenotypes,  
15 the evolutionary pathways that they followed are different depending upon the family **to**  
16 **which they belong**. Furthermore, while the carnivoran cranium more closely reflects the  
17 nature of the resources consumed –i.e., soft or hard and tough items– the mandible  
18 shows particular feeding adaptations –i.e., bamboo or bone. This finding supports the  
19 interpretation that the mandible has more evolutionary plasticity than the cranium,  
20 which is more limited to evolve towards a particular feeding adaptation. However, we  
21 find that the shapes of the cranium and the mandible are highly integrated for the whole  
22 order Carnivora. Published studies of teratological cats and dogs indicate that the role of  
23 internal constraints in shaping this pattern of integration is absent or weak and malleable  
24 by selection.  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39

40  
41 **Key words:** Evolutionary convergence, morphometrics, phenotype, durophagy,  
42

43  
44 Carnivora  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

## INTRODUCTION

### *The problem*

Convergent evolution is a central topic in evolutionary biology (Madsen et al. 2001; Nevo 2001; Winter and Oxnard 2001; Wroe and Milne 2007; Futuyma 2010). Despite this, interpretations on its potential causes are not clear-cut (e.g. Stayton 2008; Losos 2011; Olson 2012) and most contemporary surveys recognize two different points of view for explaining of the evolutionary convergence of traits: internalist and externalist explanations (e.g., Wake 1991). From an externalist point of view, convergent evolution is exclusively an evidence of adaptation due to the powerful action of natural selection (e.g., Schlueter 2000; Conway-Morris 2003; Blackledge and Gillespie 2004; Grenier and Greenberg 2005; Langerhans et al. 2006; Stayton 2008). Under this perspective of understanding phenotypic evolution, convergent traits are optimal solutions to repeated environmental problems (Losos 2011). In contrast, the internalist point of view argues that convergence is the result of internal constraints, which hamper the production of phenotypic variability upon which natural selection can operate (e.g., Alberch and Gale 1985; Maynard-Smith et al. 1985; Alberch 1989; Wake 1991; Schwenk 1995; Schwenk and Wagner 2003, 2004; Jaekel and Wake 2007; Figueirido et al. 2011a).

Although both adaptation and constraint give account of order and discreteness in phenotypic spaces (Gould 2002), the two concepts have become divorced due to a dichotomous attitude about them (Schwenk and Wagner 2004). Good examples of these “rival traditions” in evolutionary morphology are the seminal articles on this topic published recently by Losos (2011) and Olson (2012). Both reviews indicate the need

1  
2  
3 for integrative studies of the effects of natural selection and constraints using different  
4  
5 kinds of approaches for understanding phenotypic evolution.  
6  
7

8 In this article, we investigate the evolutionary convergence towards durophagy  
9  
10 in the skull of mammalian carnivores and discuss the role of natural selection and  
11  
12 different kinds of constraints in shaping this pattern. The order Carnivora is an excellent  
13  
14 choice for this study because its evolution represents one of the most spectacular cases  
15  
16 of repeated and independent evolution of similar morphologies on a limited range of  
17  
18 ecologies (Van Valkenburgh 2007; Wroe and Milne 2007; Wroe et al. 2007; Wroe  
19  
20 2008). In fact, convergent patterns in the evolution of carnivoran skull shape have been  
21  
22 reported by a number of researchers, either towards hypercarnivory (e.g., Van  
23  
24 Valkenburgh 1991; Holliday and Stepan 2001; Van Valkenburgh 2007; Wroe and  
25  
26 Milne 2007; Goswami et al. 2011; Figueirido et al. 2011a), bone-cracking (Van  
27  
28 Valkenburgh et al. 2003, 2007; Palmqvist et al. 2011; Tseng and Wang 2011) or even  
29  
30 herbivory (Figueirido et al. 2010b). However, it is worth noting that convergent trends  
31  
32 towards durophagy are still unexplored. Furthermore, durophagous carnivorans open the  
33  
34 possibility of testing if two distantly related groups of carnivorans –e.g., living pandas  
35  
36 and hyaenas, which belong to suborders Caniformia and Feliformia, respectively–  
37  
38 adapted to feed on extremely different resources –i.e., bamboo and bones, respectively–  
39  
40 converge in skull biomechanics as a consequence of the required performance to feed  
41  
42 on hard and tough foods. In order to test this hypothesis, we applied landmark-based  
43  
44 methods of geometric morphometrics in a large sample of living and extinct  
45  
46 representatives of the order Carnivora. We define convergent evolution as the repeated  
47  
48 and independent evolution of similar traits towards the same environmental regimes  
49  
50 (Futuyma 2010). Therefore, we include under evolutionary convergence both parallel  
51  
52 and convergent evolution, because many authors cast doubts on whether there is a clear-  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 cut theoretical distinction between these two evolutionary phenomena, which are  
4 frequently envisaged as extremes of a continuum (e.g., Meyer 1999; Gould 2002;  
5 Desutter-Grandcolas et al. 2005; Arendt and Reznick 2008; Hall 2007; Abouheif 2008).  
6  
7  
8  
9

### 10 *Hard and tough food*

11  
12  
13 We classify as durophagous carnivorans those species that usually feed on bones  
14 and bamboo. Bone and bamboo are hard materials because they have high values of  
15 yield strength ( $\sigma$ ) –i.e., a large force is required to produce a material failure.  
16  
17 Furthermore, they are also tough because they have high values of toughness ( $J_c$ ) –i.e.,  
18 a high capacity to absorb a large amount of energy before breaking (e.g., Wegst &  
19 Ashby 2004). The combination of these parameters relative to the Young's Modulus  
20 (i.e., elasticity) unequivocally indicates that both biomaterials are almost identical in  
21 hardness and toughness (see Fig. 1A,B). As a consequence, osseous tissue –i.e., calcium  
22 phosphate in the hydroxylapatite chemical arrangement– is a hard organic material  
23 (Schmidt-Nielsen 1984) and the physical and mechanical properties of bamboo are  
24 comparable to those of hard materials such as low-carbon steel and glass-reinforced  
25 plastics, which leads to its frequent use in industry for constructing scaffolds and as a  
26 reinforcement for cement, rubber, thermoplastic, and even aluminum (Low et al. 2005).  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42

### 43 *Bone-crackers and bamboo-feeders as durophagous carnivorans*

44  
45  
46 Bone-crackers are considered strict hypercarnivores because they feed primarily  
47 on limb bones, with high nutritional content of blood and fat-rich marrow tissues (e.g.,  
48 Kruk 1972; Palmqvist et al. 1999, 2011; Van Valkenburgh 2007). As indicated by fossil  
49 data, the bone-cracking ecomorph evolved independently at least four times during the  
50 Cenozoic: in Hyaenidae, Percrocutidae†, and twice within Canidae –in the  
51 Aelurodontina and Borophagina subtribes of subfamily Borophaginae†– (e.g., Van  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Valkenburgh 1999; Wang 1999; Van Valkenburgh 2007; Tseng and Wang 2011). In  
4  
5 contrast, bamboo-feeders are considered as strict herbivores feeding almost entirely on  
6  
7 bamboo (e.g., Gittleman 1994; Figueirido et al. 2010). Similarly, data from the fossil  
8  
9 record suggest that the bamboo-feeding ecomorph evolved independently in two living  
10  
11 species of carnivorans: the giant panda (*Ailuropoda melanoleuca*) within Ursidae and  
12  
13 the red or lesser panda (*Ailurus fulgens*) within Ailuridae (e.g., Flynn et al. 2000, 2005).  
14  
15 However, despite that both species feed entirely on bamboo they also differ in their  
16  
17 feeding behavior. The giant panda usually eat on bamboo leaves and trunks depending  
18  
19 upon the season (from March to July they feed almost entirely on trunks in the wild;  
20  
21 Schaller et al. 1985). However, they remove the outer, green, smooth and waxy layer of  
22  
23 the trunks before chewing or biting because this is their less nutritive part (Hansen et al.  
24  
25 2010). Although the red panda, feed almost entirely on leaves supplemented by arboreal  
26  
27 fruits and bamboo shoots (Reid et al. 1991; Wei et al. 1999) sometimes they also feed  
28  
29 on peeled trunks. Furthermore, it is worth noting that both pandas are the only  
30  
31 carnivorans adapted to feed regularly on leaves –i.e., a fibrous material with high  
32  
33 contents in lignin, cellulose and hemicellulose. Other herbivorous carnivorans like the  
34  
35 kinakajou (*Potos flavus*) or the Andean bear (*Tremarctos ornatus*) mostly feed regularly  
36  
37 on fruits and shoots but not on leaves (Figueirido et al. 2010b; Figueirido et al. 2011b,  
38  
39 Figueirido et al. 2012).

40  
41  
42 Therefore, we considered both bamboo-feeders and bone-crackers as durophages  
43  
44 because they feed regularly on hard and tough materials such are bamboo and bones,  
45  
46 respectively.

47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

*Anatomical and functional traits shared by bone-crackers and bamboo-feeders*

Previous researchers have noted qualitatively an external morphological resemblance between the skulls of bamboo-feeders and bone-crackers (e.g., Davis 1964) which have led to analyze both groups of carnivorans jointly in functional studies (e.g., Christiansen and Wroe 2007). In fact, a brief review of the literature indicates that both types of durophages share a number of morphological traits in the skull that could be considered potential adaptations to feed on hard and tough foods. These traits include a robust craniodental morphology with a raised and dome-like frontal region of the cranium, enlarged areas for the attachment of masticatory muscles, well-developed frontal sinuses, enlarged premolars, and microstructurally reinforced tooth enamel (Dong 2008; Figueirido et al. 2012; Joeckel 1998; Stefen 1999, 2001; Stefen and Rensberger 2002; Tanner et al. 2008; Tseng 2009). All of these morphological traits have been interpreted as adaptations to exert extremely high bite forces for chewing and biting on hard materials (Fig. 1C) and dissipate the stresses generated (Sacco and Van Valkenburgh 2004; Christiansen and Adolfssen 2005; Christiansen and Wroe 2007; Figueirido and Soibelzon 2010; Figueirido et al. 2010a, 2010b, 2011a, 2011b, 2012; Oldfield et al. 2012). However, several functional differences between bamboo feeders and bone-crackers also exist. For example, bamboo-feeders have a higher position of the condylar process of the mandible (Fig. 2) relative to the tooth row than bone-crackers which allow the former to have a slight translation movement of the lower jaw. However, although this action is an important movement for chewing and processing the fibrous bamboo leaves, the translation movement in the giant panda is very limited (only 5.4mm of translation) as a consequence of having a transverse cylindrical mandibular articulation forcing the jaw action mainly to a simple hinge movement vertically (Davis 1964).

1  
2  
3 Another morpho-functional difference between both groups of canivorans is that  
4  
5 although both have an exceptional ability for exerting large bite forces (Fig. 1C) they  
6  
7 could be using this ability in a different way. In fact, while bone-crackers are  
8  
9 particularly adapted to exert high peak forces to cracking bones during punctual periods  
10  
11 of time and to resist the stresses generated, bamboo-feeders could be adapted to resist  
12  
13 fatigue as a result of constant chewing applying sub maximal forces over protracted  
14  
15 periods. However, the fact that the giant panda torn off the stripped outer layer of  
16  
17 bamboo trunks by means of a twisting movement of the fore foot coupled with a lateral  
18  
19 turning of the head by an active bite (Davis 1964) indicates that most probably giant  
20  
21 pandas are adapted to resist both: high peak forces during punctual periods of time and  
22  
23 fatigue during constant chewing. In fact, although both pandas invest more time  
24  
25 chewing on bamboo leaves and stalks than cracking bamboo trunks (at least the red  
26  
27 panda), hyenas also feed on flesh on a regular basis and fracture limb bones less  
28  
29 frequently for accessing their medullary contents (e.g., Kruk 1972; Ewer 1973;  
30  
31 Palmqvist et al. 1999, 2011). From a biomechanical point of view, in both cases what  
32  
33 matters is the force that is exerted during those activities that demand a greater  
34  
35 resistance of the skull against the stresses generated by elevated loads (i.e., as those  
36  
37 produced during bamboo/bone-cracking). In other words, although these loads are only  
38  
39 exerted occasionally, there is safety factor which implies that the skulls of pandas and  
40  
41 hyenas must be adapted for withstanding the maximal loads exerted during chewing and  
42  
43 biting.  
44  
45  
46  
47  
48  
49

50 In sum, bone-crackers and bamboo-feeders feed regularly on extremely hard and  
51  
52 tough materials (Fig. 1A,B). Accordingly, both groups of carnivorans seem to share a  
53  
54 set of skull traits related with the ability for exerting exceptionally large bite forces (Fig.  
55  
56 1C) and to dissipate the stresses generated. As a consequence, these shared traits  
57  
58  
59  
60

1  
2  
3 between both groups of carnivorans could be interpreted as potential adaptations  
4  
5 towards durophagy. However, in spite of this morphological resemblance and presumed  
6  
7 functional similarity between bone-crackers and bamboo feeders there is a lack of  
8  
9 ecomorphological studies of skull shape evolution towards durophagy in a  
10  
11 comprehensive way. This is particularly the main objective of this article.  
12  
13

## 14 MATERIAL AND METHODS

### 15 *Data collection and geometric morphometrics*

16  
17  
18 We collected 299 mandibles and 322 crania of adult individuals belonging to 57 species  
19  
20 –45 living and 12 extinct (†)– of all living terrestrial families of the order Carnivora  
21  
22 plus the extinct family Percrocutidae (see Table 1). We collected data from only adult  
23  
24 individuals –as indicated by closed basilar synchondroses and complete tooth eruption.  
25  
26 Mandible and cranial shapes were recovered with a set of landmarks representing  
27  
28 functional key features in the carnivoran skull (Fig. 2). Landmarks were digitized into  
29  
30 two-dimensional Cartesian coordinates ( $x,y$ ) on high-resolution digital images using  
31  
32 TPSdig V. 2.11 (Rohlf 2008). Digital images were collected using a tripod and following  
33  
34 a standardized protocol for avoiding lens distortion and parallax. Later, interlandmark  
35  
36 distances were modeled by means of an outline, in order to obtain clearer shape  
37  
38 transformation models in subsequent multivariate analysis. All the specimens were  
39  
40 aligned using Procrustes superimposition procedure and projected onto the tangent  
41  
42 space (Dryden and Mardia 1998).  
43  
44  
45  
46  
47  
48

### 49 *Testing the influence of phylogeny and allometry in skull shape*

50  
51 In order to test the presence of phylogenetic signal in our data and to assess for  
52  
53 phylogenetic patterning in multivariate analyses, a phylogenetic consensus tree was  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 assembled with Mesquite (Maddison and Maddison 1997) using published sources (see  
4  
5 Fig. 3).  
6  
7

8 We used a permutation approach developed by Laurin (2004), extended for  
9  
10 multivariate analysis by Klingenberg and Gidaszewski (2010) and applied to shape data  
11  
12 by Gidaszewski et al. (2009), Figueirido et al. (2010) and Klingenberg et al. (2012) to  
13  
14 simulate the null hypothesis of complete absence of phylogenetic signal in mandible  
15  
16 and cranial shapes. The 56 observed mean species shapes were randomly distributed as  
17  
18 the tips of the phylogeny in 10,000 permutations. For each permutation, tree length (i.e.,  
19  
20 the sum of the squared Procrustes distances between ancestral and descendant shapes  
21  
22 for all branches) was computed. If the resulting tree length computed for each  
23  
24 permutation was greater than the one obtained with the original data, the null hypothesis  
25  
26 of absence of phylogenetic structure in our data was rejected. A *P*-value was used for  
27  
28 assessing the presence of phylogenetic signal in shape (Klingenberg and Gidaszewski  
29  
30 2010).  
31  
32  
33  
34

35 A multivariate regression analysis (Monteiro 1999) of shape (i.e., using  
36  
37 Procrustes coordinates-Pco) on size (i.e., using Centroid size-Cs) was performed for  
38  
39 testing the influence of allometry. Independent contrast analysis (Felsenstein 1985) was  
40  
41 applied to take into account the phylogenetic relationships of the species under study  
42  
43 (e.g., Harvey and Pagel 1991; MacLeod 2001). Therefore, the contrast for shape (Pco)  
44  
45 was regressed on the contrast for size (Cs). The statistical significance was tested with a  
46  
47 permutation test against the null hypothesis of complete independence of shape on size  
48  
49 (Drake and Klingenberg 2008).  
50  
51  
52

53  
54 ***The phenotypic spaces and their histories of phylogenetic occupation***  
55  
56  
57  
58  
59  
60

1  
2  
3 The distribution of durophages in the phenotypic space was explored by Principal  
4 Components Analysis (PCA) using the covariance matrix. As PCA finds orthogonal  
5 axis of maximal variance, it is therefore a suitable method for exploring the phenotypic  
6 variation of mandible and cranium shapes.  
7  
8  
9  
10

11  
12 To investigate the phylogenetic history of the phenotypic space occupation, we  
13 first reconstructed the ancestral states of cranium and mandible shapes using the  
14 squared-change parsimony method (Maddison 1991) weighted by branch lengths (see  
15 Fig. 3). Subsequently, these shapes were plotted on the original phenotypic spaces and  
16 the branches of tree were connected (Klingenberg and Ekau 1996; Rholf 2002; Polly et  
17 al. 2008; Gidazweski et al. 2009; Klingenberg and Gidaszewski 2010; Figueirido et al.  
18 2010; Klingenberg 2012) to obtain mandible and cranium “phylomorphospaces”. This  
19 approach provided us a unique opportunity to explore the history of phylogenetic  
20 occupation of phenotypic spaces. Furthermore, we explored the evolutionary trajectories  
21 of shape transformation towards durophagy by investigating the shape changes from  
22 ancestral state reconstructions to the tips of the phylogeny in durophagous taxa.  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36

### 37 *Exploring shared morphological traits in the skull of durophagous carnivores*

38  
39  
40 Given that PC axes reflect the greatest variance in the whole dataset and the  
41 specimens are ordered on these axes according to the major patterns of shape variation,  
42 PCA is not an appropriate method for discriminating a priori defined groups of  
43 specimens. In order to separate durophages from other carnivores included in the  
44 sample, we computed a linear discriminant analysis (DA) from cranium and mandible  
45 shapes. Similarly, the morphological differences between bone-crackers and bamboo-  
46 feeders were also explored using DA. The reliability of the discrimination was assessed  
47 by the “leave-one-out” cross-validation method (e.g., Timm 2002) given the high  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 dimensionality of our data. Subsequently, the statistical significance of pairwise  
4  
5 differences in mean shapes was assessed with a permutation test (10,000 in our case)  
6  
7 using the Mahalanobis distances (MD) between groups.  
8  
9

10  
11 ***Investigating patterns of covariation between mandible and cranium shapes in***  
12  
13 ***durophagous carnivorans***  
14

15  
16 Patterns of covariation between cranium and mandible shapes were explored using two-  
17  
18 block partial least squares (2B-PLS) analysis (Rohlf and Corti 2000; Zelditch et al.  
19  
20 2004; MacLeod 2006). Separate Procrustes fits for each block –i.e., mandible and  
21  
22 cranium shapes– were performed for analyzing the shape of the two configurations of  
23  
24 landmarks (Klingenberg 2009). Also, the *RV* coefficient of Escoufier (1973) was  
25  
26 computed as a scalar measure of the strength of the association between mandible and  
27  
28 cranium shapes. Statistical significance was tested with a permutation test against the  
29  
30 null hypothesis of complete absence of covariation between blocks (Klingenberg 2009).  
31  
32 In addition, the *RV* coefficient of a second PLS analysis from the contrasts for both  
33  
34 blocks of variables was computed for exploring if covariation between mandible and  
35  
36 cranium shapes was due to phylogenetic patterning. Statistical significance of the *RV*  
37  
38 coefficient was tested by a permutation test (10,000 in our case) against the null  
39  
40 hypothesis of independence between both block of variables.  
41  
42  
43  
44

45  
46 All the morphometric procedures, including multivariate analyses, were performed with  
47  
48 MorphoJ software package (Klingenberg 2011).  
49

50  
51 **RESULTS**  
52

53  
54 ***The influence of phylogeny and allometry on skull shape***  
55  
56  
57  
58  
59  
60

1  
2  
3 The permutation test indicated a strong phylogenetic signal in both cranium size and  
4 shape (LogCs: *Tree length* = 2.39619,  $P < 0.0001$ ; Pco: *Tree length* = 0.2144,  
5  
6  
7  $P < 0.0001$ ) and mandible size and shape (LogCs: *Tree length* = 2.76301,  $P < 0.0001$ ; Pco:  
8  
9  
10 *Tree length* = 0.25306,  $P < 0.0001$ ).

11  
12 The multivariate regression of Pco on LogCs was statistically significant for  
13 both crania and mandibles ( $n = 322$ ; cranium:  $P < 0.0001$ ;  $n = 299$ ; mandible:  $P = 0.001$ ),  
14  
15 with a percentage of shape explained for by size differences of 8.609% for the crania  
16  
17 and 1.557 % for the mandible. The contrast for shape (Pco) against the contrast for size  
18  
19 (LogCs) yielded a clear significant association for both the cranium ( $n = 56$ ;  $P = 0.001$ )  
20  
21 and the mandible ( $n = 55$ ;  $P = 0.010$ ), with percentages of shape explained for by size  
22  
23 differences of 14.666% and 5.562%, respectively. Therefore, since allometry is a  
24  
25 significant source of shape variation in our sample, the regression residuals of shape on  
26  
27 size were computed to eliminate the predicted component of shape variation due to size  
28  
29 differences (Klingenberg 2009). These regression residuals were used as strict shape  
30  
31 variables free of allometric effects in all subsequent multivariate analyses. It is worth  
32  
33 noting that it would be ideal to compute a pooled-within species regression analysis  
34  
35 rather than an conventional multivariate regression (at least in those cases were all the  
36  
37 species share the same regression slopes). However, the fact that some species in our  
38  
39 sample are only represented by one or two individuals, particularly in the case of fossil  
40  
41 taxa, precludes us from performing the pooled within-species regression analysis.  
42  
43 Furthermore, the slopes within species were different which also precludes us from  
44  
45 obtaining residuals from this regression analysis.  
46  
47  
48  
49  
50  
51  
52

53 ***The phenotypic spaces***  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 The phenotypic spaces derived from the PCA of the covariance matrix for the  
4 residuals of the 322 crania are shown in Figures 4A, B. Given that the first three  
5 Principal Components (PCs) represent a reasonable amount of the total shape variation  
6 (>72% of variance explained), we only present here these three axes. The first PC (Fig.  
7 4A) differentiates clearly the dolichocephalic crania of the family Canidae –with  
8 negative scores– from the crania of the other species included in the sample –scoring  
9 positively (Fig. 4C). In contrast, the second PC (Fig. 4A) describes a shape gradient that  
10 goes from the crania of the family Felidae –with positive scores– to the crania of the  
11 other specimens included in the sample –scoring negatively– according with those  
12 morphologies showed in Figure 4D. The third PC (Fig. 4B) accounts for shape changes  
13 from the cranium of durophages –taking positive scores– to crania of other species –  
14 with negative scores (Fig. 4E).

15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30 The PCA computed from the covariance matrix for the residuals of the 299  
31 mandibles included in the sample yielded 20 PCs. Again, the first three axes explained  
32 >70% of the original variance, and therefore, we only show here the results derived  
33 from them. The first PC (Fig. 4F) accounts for the shape changes that take place from  
34 the mandibles of the kinkajou (*P. flavus*) –with positive scores– to those of other  
35 species –with extreme negative scores some representatives of the family Canidae (Fig.  
36 4H). Conversely, the second PC (Fig. 4F) relates mainly with a morphological gradient  
37 that goes from the mandibles of caniform carnivorans –with extreme negative scores for  
38 the mandibles of Ailuridae and Ursidae– to the ones of feliforms, showing extreme  
39 positive scores, with those shapes depicted in Figure 4I. Finally, the third PC accounts  
40 for shape changes from the mandible of durophagous carnivorans, taking negative  
41 scores in this third axis (Fig. 4G), to those of other carnivores –scoring positively– with  
42 the morphological traits shown in Figure 4J.

### *The phylomorphospaces and the inferred evolutionary pathways towards durophagy*

The phylogenetic history of the phenotypic space occupation depicted from the first and second PCs of both crania and mandibles (Figs. 4A, F) resulted in the phylomorphospaces shown in Figures 5A, C. In general, many terminal branches of the tree are relatively short whereas the internal branches appear to be long. This indicates that closely related species possess similar cranial shapes (Gidaszewski et al. 2009). Furthermore, branches tended to be quite uncrossed in these phylomorphospaces, which indicates a low degree of convergence in the shapes recovered by the first two eigenvectors. Therefore, since the shapes of the species belonging to the same family were often found in the same areas of these morphospaces, a high degree of phylogenetic structure is implied. However, the phylogenetic history of the phenotypic space occupation depicted from the second and third PCs (Fig. 4B, G) of both anatomical structures (Fig. 5B, D) indicates the presence of evolutionary convergence. In fact, a visual inspection of these graphs allows appreciating that the internal branches of the tree are relatively short compared to the terminal ones. This suggests that some closely related species showed clearly different cranial and mandibular shapes (Gidaszewski et al. 2009). Moreover, in both plots there are many sharp changes of direction in the evolutionary trajectories, with many crossed branches leading to a “messy” appearance (Fig. 5B, D).

It is worth noting that the reconstructed evolutionary trajectories of skull shape transformation are different depending on the family that which durophages belong (Fig. 5E). However, as revealed by the phenotypic spaces (Fig. 4B,G) and their respective phylomorphospaces (Fig. 5B, D), durophages have attained similar morphological traits, which suggests their convergent adaptation to feeding on hard/tough tissues (i.e., bone and bamboo). In contrast, other morphological traits have

1  
2  
3 evolved independently as specific features of bone-crackers and bamboo-feeders (Fig.  
4  
5 5E).

### 8 *The morphological traits of the skull shared by durophages*

9  
10  
11 In order to quantify those traits shared by durophages that were identified in the  
12 phenotypic spaces, a DA was computed to identify those morphological traits that best  
13 distinguish between durophages and other carnivores. The discrimination of both  
14 groups, using cranium shape, was highly significant (MD = 4.7468;  $P < 0.0001$ ; >95%  
15 of correct classifications [CC]; Fig. 6A). Durophages are characterized by having a deep  
16 cranium with a large sagittal crest, downward positioned orbits, a short and deep snout,  
17 and reinforced postglenoid processes (Fig. 6C; see also Fig. 2D). The discrimination  
18 between both groups using mandible shape was also significant (MD = 3.1686;  $P <$   
19 0.0001; ~90% CC; Fig. 6E) indicating that, compared to other carnivores, the mandible  
20 of durophages has a large coronoid, a more dorsally positioned condyle, and a more  
21 concave and deeper corpus (Fig. 6G).

22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37 In order to identify those morphological traits that best distinguish between  
38 bamboo-feeding and bone-cracking carnivorans, a second DA was computed between  
39 both groups. The discrimination between bone-crackers and bamboo-feeders using  
40 cranium shape was significant [MD = 13.9643;  $P < 0.0001$ ; >95% CC; Fig. 6B], which  
41 allows recognition of a set of cranial traits indicative of these specific feeding behaviors  
42 (Fig. 6D). Specifically, bamboo-feeders have a more developed occiput anteriorly,  
43 larger post-carnassial molars, more horizontal zygomatic arches, more anteriorly placed  
44 orbits and a larger postglenoid process relative to bone-cracker carnivores (see Fig. 2).  
45  
46  
47 In contrast, bone-crackers are characterized by having well developed carnassials and  
48 pre-carnassial molars, as well as more frontalized orbits and a posteriorly developed  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 occiput (Fig. 6D). Similarly, the discrimination between both feeding groups using  
4  
5 mandible shape was highly significant ( $MD = 9.8617$ ;  $P < 0.0001$ ; 100% CC; Fig. 6F).  
6  
7 The results obtained show that the mandibles of bamboo-feeders are larger and with a  
8  
9 more anteriorly oriented coronoid, a more dorsally positioned condyle, are deeper below  
10  
11 the postcarnassial molars and shows a more developed grinding dentition relative to  
12  
13 bone-crackers. In contrast, bone cracking carnivores are characterized by having a well  
14  
15 developed anterior dentition (i.e., premolars) (Fig. 6H).  
16  
17  
18

### 19 *Patterns of covariation between cranium and mandible shape in durophages*

20  
21  
22 The 2B-PLS analysis performed for testing the covariance between crania and  
23  
24 mandibles is shown in Figure 7. The morphological covariation between cranium and  
25  
26 mandible shape was statistically significant ( $RV = 0.663$ ;  $P < 0.0001$ ). Similarly, the 2B-  
27  
28 PLS computed from the contrast of mandible and cranium shapes provided statistically  
29  
30 significant results ( $RV = 0.599$ ;  $P < 0.0001$ ).  
31  
32  
33

34 From a total of nine significant pairs of PLS axes, we only show here the results  
35  
36 of the first three, because they jointly account for >98% of the total shape covariance.  
37  
38 The first pair of axes (85.9 % of the total covariance) mainly explained changes from  
39  
40 dolichocephalic crania and mandibles to brachycephalic ones (Figs. 7A, B). The second  
41  
42 pair of axes (9.1% of explained covariance) was also significant ( $P < 0.0001$ ) and they  
43  
44 mainly explain changes related to the shape of the neurocranium and zygomatic arches  
45  
46 (among the cranium) plus the orientation of the coronoid process and the corpus (among  
47  
48 the mandible) (Figs. 7C, D). Finally, the third pair of PLS axes, although it explains a  
49  
50 low proportion of the total covariance (only 3.5%), is still significant ( $P < 0.0001$ ) and  
51  
52 mainly separates the skull of durophagous carnivorans from those of other species  
53  
54 included in the sample (Fig. 7E, F).  
55  
56  
57  
58  
59  
60

## DISCUSSION

### *Durophagous carnivorans are confined in the phenotypic space*

Our results demonstrate that despite the different resources consumed by durophagous carnivores, bone-crackers and bamboo-feeders share the same regions of phenotypic skull space (Figs. 4B, G). This demonstrates that both groups of carnivorans share a set of anatomical traits in their skulls (Figs. 4E, J), which leads us to hypothesize that they could represent morphological adaptations towards “durophagy”. However, we obtained a significant phylogenetic signal in both mandible and cranium shapes, which casts doubt on considering these traits as solely the outcome of natural selection. The study of phylomorphospaces and the inferred evolutionary trajectories provide interesting clues on the role of natural selection in shaping these traits.

### *The evolutionary routes to reach adaptations towards durophagy*

Both the reconstructed phylomorphospaces and the inferred hypothetical evolutionary pathways of skull shape transformation towards durophagy indicate that natural selection is not sufficient for explaining the shared anatomical traits among bone-crackers and bamboo-feeders. In fact, although durophages reached similar –but not identical– skull morphologies (Fig. 5E) and show similar deviations from their sister groups, their evolutionary pathways differed depending on the phylogenetic legacy of each particular family. In sum, the results of PCA indicate that the traits shared between both carnivoran groups are strongly influenced by both phylogenetic inheritance (including here different types of constraints; Schwenk 2003) and natural selection. However, further investigation based in Bayesian approaches (e.g., Slater et al. 2012) could clarify more finely to which model of evolution fits the evolutionary patterns described here.

### *Morphological adaptations of durophagous carnivorans*

Given that the groups of “durophagous” carnivores and “other taxa” are both represented by different families belonging to two different suborders, the results obtained in DA allow us to identify a shared suite of morphological features among durophages that are relatively independent of the phylogenetic legacy of each particular family. Such features could be *a priori* considered as potential adaptations towards durophagy. However, convergence *per se* is not indicative of adaptation (Stayton 2008; Losos 2011). In fact, evolutionary convergence of traits can occur without the implicit action of natural selection favoring them within a particular selective environment (see Losos [2011] and references therein). Therefore, since phenotypes may evolve convergently in similar environments, even if not directly favored by natural selection, independent arguments are necessary for testing the adaptative nature of the traits shared by durophages.

Functional studies of trait evolution could give clues to test our adaptation hypothesis, as selection operates on the functional consequences of traits (Arnold 1983). There is compelling evidence from skull biomechanics in mammals (Fig. 2B) that all these traits enhance the ability to produce the large bite forces required for feeding on hard/tough foods and for resisting the high compressive loads generated during chewing and biting (e.g., Ewer 1973). For example, a well developed sagittal crest (Fig. 2D) indicates a large area of attachment for the temporalis muscle (e.g., Ewer 1973) and temporalis volume correlates well with bite force (e.g., Christiansen and Adolfsen 2005; Christiansen and Wroe 2007; Wroe et al. 2005). Similarly, large distances between the condyle and both the coronoid –i.e., insertion area of the temporalis– and angular –i.e., insertion area of the masseter– processes of the mandible (Fig. 2D) enhance the input levers of the temporalis and masseter muscles, respectively (Fig. 2B), which increases

1  
2  
3 mechanical advantage and bite force (Biknevicius and Van Valkenburgh 1996).  
4  
5 Similarly, a shortened **facial skeleton** (Fig. 2D) diminishes the output moment arms at  
6  
7 different points of the dentition (Fig. 2B) and also increases bite force (Turnbull 1976;  
8  
9 Biknevicius and Van Valkenburgh 1996; Christiansen and Wroe 2007). Enlarged  
10  
11 **postglenoid** processes result in a deep glenoid fossa (Fig. 2D), which precludes  
12  
13 dislocating the lower jaw when large bite forces are exerted at closing angles (Fig. 2A)  
14  
15 (Ewer 1973, Werdelin, 1989). A deep frontal region of the skull is the consequence of  
16  
17 having large paranasal sinuses, which are involved in dissipating the high compressive  
18  
19 loads generated during chewing (dotted lines in Fig. 2B) (Werdelin 1989; Joeckel 1998;  
20  
21 Tanner et al. 2008 ). Similarly, a deep mandibular corpus correlates **to** a large  
22  
23 proportion of cortical bone, which is responsible for dissipating compressive forces  
24  
25 (Biknevicius and Ruff 1992). Therefore, **since** these anatomical features of the skull  
26  
27 enhance the biomechanical performance required to feed on hard/tough foods, it seems  
28  
29 reasonable that these phenotypes represent convergent adaptations towards durophagy.  
30  
31  
32  
33

34  
35 ***The cranium of durophages reflects the “nature” of the resources consumed and***  
36  
37 ***their mandible the “type” of food***  
38

39  
40 Although the DA performed for separating durophagous carnivores from other  
41  
42 taxa provided significant results using both the cranium and the mandible, the  
43  
44 discrimination **using** cranial shape was better than the one with mandible shape  
45  
46 (compare the overlapping areas –in grey– of the histograms depicted in **Figs. 6A, E**).  
47  
48 **The crania of bone-crackers and bamboo-feeders are more similar in shape than their**  
49  
50 **mandible. In fact, this was already noted from the PCA** (compare the areas occupied by  
51  
52 durophages in Figs. **4B, G**). This result was unexpected, as previous investigations  
53  
54 indicated that lower jaw shape has more evolutionary plasticity towards particular  
55  
56 feeding adaptations than cranial shape (Barone 1986). The reason is that while mandible  
57  
58  
59  
60

1  
2  
3 morphology mainly reflects feeding behavior, cranium shape is more constrained by  
4  
5 conflicting functional demands (e.g., feeding, olfaction, vision and brain processing;  
6  
7 Figueirido and Soiblezon 2010; Figueirido et al. 2010a,b, 2011a,b). Furthermore, the  
8  
9 cranium of carnivores is made up of a set of bones (i.e., ~33) but the mandible is only  
10  
11 composed of the dentary bone. As a consequence, the lower jaw should be a structure  
12  
13 easier to modify towards durophagy than the cranium (Figueirido et al. 2012). However,  
14  
15 the results obtained here point to the opposite direction.  
16  
17

18  
19 The second round of DA performed to discriminate between bamboo-feeders  
20  
21 and bone-eaters gave clues about this counterintuitive result. Here, we obtained a better  
22  
23 discrimination between both groups of durophages using mandible shape than using  
24  
25 cranium shape (compare the histograms depicted in Figs. 6B, F). This result confirmed  
26  
27 previous statements on the higher evolutionary plasticity of the carnivoran mandible  
28  
29 compared to the cranium, as the latter seems to be more limited to evolve towards the  
30  
31 particular feeding behavior of a durophage. As a result, while there are two different  
32  
33 mandible solutions to be a durophage, depending upon the resources consumed –i.e.,  
34  
35 bamboo or bone– there is only one possible solution for the cranium. As a consequence,  
36  
37 cranial shape reflects the nature of the resources consumed –i.e., hard or soft tissues–  
38  
39 but mandible morphology more finely reflects the particular type of food –i.e., bone or  
40  
41 bamboo. For this reason, durophagous carnivorans converge more in cranial shape than  
42  
43 in lower jaw shape.  
44  
45  
46  
47

48  
49 ***Durophagous carnivorans share patterns of covariation between the shapes of crania***  
50  
51 ***and mandibles***  
52

53  
54 The fact that both structures reflect different aspects of feeding behavior in  
55  
56 durophages opened the possibility to test if the cranium and the mandible were  
57  
58  
59  
60

1  
2  
3 morphologically integrated. Furthermore, if cranial shape is more limited to evolve  
4  
5 towards durophagy than mandible shape, we should expect a lower variation in their  
6  
7 crania than in their mandibles. We analyzed patterns of covariation between cranium  
8  
9 and mandible shape using PLS-analysis for exploring quantitatively this hypothesis.  
10

11  
12 In spite of the fact that the cranium and the mandible of durophages reflect  
13  
14 different aspects of feeding behavior –i.e., nature and type of the resources consumed–  
15  
16 our results clearly demonstrate that cranial and lower jaw shape are highly integrated in  
17  
18 carnivores. This result was in part expected because other authors have identified the  
19  
20 same two modules coordinating shape variability in the cranium –i.e., neurocranium and  
21  
22 **facial skull**; Drake and Klingenberg (2010)– and in the mandible –i.e., ramus and  
23  
24 corpus; Meloro et al. (2011). Furthermore, from a functional perspective this result was  
25  
26 also expected, because the ramus is mainly connected with the neurocranium by the  
27  
28 same main muscles involved in mastication and the corpus fits with the **facial skeleton**  
29  
30 by the occlusion of upper and lower teeth.  
31  
32  
33  
34

35  
36 Durophagous carnivorans also share patterns of covariation (Figs. 7E, F), most  
37  
38 probably as a result of the functional demands to feed on tough foods. **It is obviously**  
39  
40 **inconceivable to have a “strong” cranium equipped to withstand high bite forces and a**  
41  
42 **“weak” mandible without these biomechanical adaptations.** Furthermore, for those  
43  
44 morphological aspects recovered from the third pair of PLS-axes –which are those traits  
45  
46 that best distinguish the pattern of durophagous covariation– cranial shape shows much  
47  
48 less variation than lower jaw shape (Fig. 7E; *box-plots*), which confirms that the former  
49  
50 has a more limited evolvability. However, this interpretation is mainly functional and  
51  
52 again, the presence of a strong phylogenetic signal in cranium and mandible shape cast  
53  
54 doubts on **whether** this pattern is exclusively an evidence of adaptation or reflects a  
55  
56 constraint.  
57  
58  
59  
60

1  
2  
3 *Teratological cats and dogs provide evidence that covariation in lower jaw and*  
4  
5 *cranial shape is externally governed*  
6  
7

8           The study of some cat –e.g., exotic short-hair cats– and dog –e.g., English  
9 bulldogs– breeds with a substantially longer lower jaw relative to the upper jaw (Fox  
10 1963; Schlueter et al. 2009) constitutes a complementary approach to test if the pattern  
11 of covariation reported between cranial and lower jaw shape is externally or internally  
12 governed. These breeds were already considered as teratological phenotypes by Charles  
13 Darwin (1875), who identified the English bull-dog as a monstrosity fixed by man’s  
14 selection. Experiments on artificial selection provide a valuable experimental test for  
15 assessing whether phenotypes not observed in nature are the result of selection or  
16 constraint (Losos 2011; Olson 2012), as they contribute information about which  
17 phenotypes can and cannot be produced by normal development (e.g., Alberch 1989;  
18 Galis et al. 2006; Olson 2012).  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32

33           Brachycephalic breeds are known in the veterinary jargon as “undershot bite”.  
34 The “undershot bite” or brachycephalic condition is considered the result of having a  
35 modified achondroplastic cranium with a extremely shorted snout and palatal region,  
36 which results in a wide face and causes the mandible to overhang beyond the maxilla  
37 (Stockard 1941; Fox 1963). All of these morphological traits are the breed standard in  
38 brachycephalic dogs (American Kennel Club 2006), which constitutes a documented  
39 record of the selection regime applied by breeders (Young and Bannasch 2006). The  
40 genetic cause of the brachycephalic condition –at least in dogs– is a mutation in the  
41 retro-gene coding for fibroblast growth factor 4 (*FGFR4*) (Parker et al. 2009), which  
42 causes an abnormal pattern in cartilage formation. In contrast, the opposite condition,  
43 the “overshot bite” or the “the parrot-mouth” seen in several dogs –e.g., in German  
44 Shepards (Fox 1963) and cattle have never been selected in any specific breed. In fact,  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 this abnormal phenotype, when present, entails a set of injuries caused by the occlusion  
4  
5 of the mandibular teeth in the soft tissues of the palatal region (Fox 1963). However,  
6  
7 even although not being selected by breeders, this deformity seems to appear regularly  
8  
9 in some dog breeds (see Stockard 1941; Fox 1963) and it is corrected, whenever  
10  
11 possible, during the first months of individual's life.  
12  
13

14  
15 Both abnormal conditions seen in **domesticated animals** indicate that these  
16  
17 phenotypes are developmentally possible, which points to the hypothesis that  
18  
19 phenotypes lacking integrated mandibles and crania are not the result of an internal  
20  
21 constraint. In fact, this was already noted by Fox (1963) referring to Stockard (1941):  
22  
23 *“By crossbreeding different types of dogs (dolicocephalic, mesocephalic, and*  
24  
25 *brachycephalic) a wide spectrum of hybrid variations have been produced and it was*  
26  
27 *concluded that the length of the upper jaw is inherited independently of the lower jaw*  
28  
29 *and vice versa”*.  
30  
31

32  
33 Therefore, both phenotypes which **do not have** integrated mandibles and crania  
34  
35 are ontogenetically possible. Consequently, there is no reason to invoke the presence of  
36  
37 developmental constraints to shape the pattern of covariation between the shapes of the  
38  
39 mandible and the cranium (Fig. 8). Thus, two possible evolutionary scenarios could  
40  
41 explain the pattern of integration: (i) that this pattern is internally –developmentally–  
42  
43 governed in wild species, but in the domestic ones the pleiotropic effects that integrate  
44  
45 both structures have disappeared because artificial selection entailed both directional  
46  
47 and stabilizing selection in lower jaw and cranial shapes, respectively (see Wagner,  
48  
49 1996; Wagner and Altenberg, 1996; Wagner et al., 2007); and (ii) that the observed  
50  
51 pattern of integration is externally governed, and hence, it is only the outcome of natural  
52  
53 selection. Under the latter scenario, both abnormal phenotypes of cats and dogs would  
54  
55 be eliminated in the wild by natural selection at adult stages, as they result in several  
56  
57  
58  
59  
60

1  
2  
3 disorders in both dogs –e.g., dental anomalies, palatitis or otocephaly; Fox (1963)– and  
4  
5 cats –e.g., abnormal dislocation of the ventral nasal concha, horizontally rotated upper  
6  
7 canine teeth and steeply oriented nasolacrimal drainage system; Schlueter et al. (2009)–  
8  
9 which makes their survivorship almost impossible without veterinary care.

10  
11 Distinguishing between these two hypotheses is a difficult task with the actual data. In  
12  
13 any case, it is clear from the pattern of integration obtained here that the role of  
14  
15 developmental constraints is absent or weak, and malleable by selection.  
16  
17

## 18 19 CONCLUSIONS

20  
21 We have demonstrated that in spite of the extreme differences in the type of  
22  
23 resources consumed by durophagous carnivorans, bone-crackers and bamboo-feeders  
24  
25 share a set of unique traits. Furthermore, as these shared traits enhance the fitness of  
26  
27 taxa under this environmental regime, we interpret them as adaptations to feed on  
28  
29 tough/hard foods. Therefore, the external action of natural selection in shaping these  
30  
31 traits is clear. However, the adaptations towards durophagy were reached independently  
32  
33 in different carnivoran families that departed from quite different ancestral  
34  
35 morphologies, which forced them to follow different evolutionary pathways to reach a  
36  
37 similar phenotypic solution. Therefore, both the particular phylogenetic legacy of each  
38  
39 durophagous family and the action of natural selection led to a pattern of incomplete  
40  
41 convergence (Herrel 2004; Stayton 2008) in skull shape evolution towards durophagy.  
42  
43 Furthermore, the cranium of carnivores is more limited to evolve towards durophagy  
44  
45 than the mandible, perhaps because cranial morphology is a compromise between  
46  
47 different functions while lower jaw shape is only involved in food acquisition and  
48  
49 processing, or maybe because the cranium is composed of more bones than the  
50  
51 mandible. In any case, this limitation is evidenced by the lower variation of the cranium  
52  
53 compared to the mandible, and has led to a higher degree of evolutionary convergence  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 in cranial shape than in lower jaw shape towards durophagy. While the mandible  
4 reflects the particular trophic adaptations of durophages –bamboo-feeding or bone-  
5 cracking– the cranium only reflects the nature of the items consumed –i.e., soft or hard  
6 tissues. In spite of the fact that cranial and lower jaw shape reflect different aspects of  
7 feeding behavior, both anatomical structures are highly integrated in the carnivoran  
8 skull. Previous studies of teratological cats and dogs subject to strong artificial selection  
9 indicated that non-integrated phenotypes are ontogenetically possible. This indicates  
10 that, if the pattern of integration between the shapes of the cranium and the mandible in  
11 carnivorans is the result of an internal constraint, it can be eliminated by strong artificial  
12 selection. The alternative explanation is that this constraint does not exist and the  
13 pattern of integration is merely due to natural selection. Therefore, we would expect an  
14 overlapping set of selective forces acting on both the cranium and the mandible, in  
15 addition to other non-masticatory demands on the cranium. Future analyses, for  
16 example the generation of biomechanical models that vary according to the major axes  
17 of morphological change highlighted in our morphometric analyses, will further clarify  
18 the functional basis not only for the differences in skull shape between durophages and  
19 other carnivorans, but also between bamboo and bone specialists.

20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41 The results obtained in this paper point to understanding evolutionary  
42 convergence as the combined result of natural selection and constraint. We do not  
43 expect evolutionary convergence of traits in the same environmental regime without the  
44 implicit action of natural selection (except those cases of chance, exaptations or  
45 correlations on another characters; see Losos [2011] and references therein), but its  
46 action has to be envisaged –in our view– upon a limited range of phenotypic variants  
47 imposed by different types of constraints –genetic, developmental, or structural. All of  
48 these potential constraints could be grouped under “phylogenetic constraints”. Although  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 this term has a low operational value, it is a useful heuristic concept (Schwenk 1994).  
4  
5 Therefore, we refer to phylogenetic constraints as those aspects derived from the early  
6  
7 definition of body plans determined early in lineage phylogenies within determined  
8  
9 biomaterials and under highly canalized developmental routes (e.g., Seilacher 1979;  
10  
11 Maynard-Smith 1985; Gould and Lewontin 1979; Gould 1989; Wagner 1988; Schluter  
12  
13 1988; Ligon 1993; Thomas and Reif 1993; Schwenk 1995; Cheverud 1982, 1985;  
14  
15 Wagner 1996), which strongly biases the production of future phenotypic variants.  
16  
17

### 18 19 **ACNOWLEDGEMENTS**

20  
21 We thank E. Westwig, Judy Galkin (AMNH, New York), P. Jenkins, L. Tomsset and  
22  
23 Roberto Portella (NHM, London) for kindly providing us access to the specimens under  
24  
25 their care. We are also grateful to Christine M. Janis, Paul Palmqvist, Juan A. Pérez-  
26  
27 Claros and two anonymous reviewers whose comments and suggestions greatly  
28  
29 improved the rigor of the manuscript contents. BF would like to dedicate this paper to  
30  
31 Miquel De Renzi who gave him the underlying logic for understanding monsters. This  
32  
33 study was supported by Fulbright Postdoctoral Grant FU2009-0184 (to B.F.).  
34  
35  
36  
37

### 38 39 **REFERENCES**

- 40  
41 Abouheif, E. (2008). Parallelism as the pattern and process of mesoevolution. *Evol.*  
42  
43 *Dev.* 10, 3–5.  
44  
45  
46  
47 Alberch, P., and E. A. Gale. 1985. A developmental analysis of an evolutionary trend—  
48  
49 digital reduction in amphibians. *Evolution* 39:8–23.  
50  
51  
52 Alberch, P. 1989. The logic of monsters: evidence for internal constraint in development  
53  
54 and evolution. *Geobios* 12:21-57.  
55  
56  
57 American Kennel Club 2006 *The complete dog book*. New York, NY: Ballantine Books.  
58  
59  
60

1  
2  
3 Arendt, J., and D. Reznick. 2008. Convergence and parallelism reconsidered: what have  
4 we learned about the genetics of adaptation? *Trends Ecol. Evol.* 23, 26–32.  
5  
6

7  
8 Arnold, S. J. 1983. Morphology, performance and fitness. *Am. Zool.* 23:347–361.  
9

10  
11 Barone, R. 1986. *Anatomie Compare des Mammifères Domestiques*, vol. 1. Vigot, Paris.  
12

13  
14 Bardeleben, C., R.L. Moore, and R.K. Wayne. 2005. A molecular phylogeny of the  
15 Canidae based on six nuclear loci. *Mol. Phylogenet. Evol.* 37, 815–831.  
16  
17

18  
19 Blackledge, T. A. and, R. G Gillespie. 2004. Convergent evolution of behavior in an  
20 adaptive radiation of Hawaiian web-building spiders. *Proc. Natl. Acad. Sci. USA* 101,  
21 16228-16233.  
22  
23  
24

25  
26 Bicknevicus, A.R., and B. Van Valkenburgh. 1996. Design for killing: craniodental  
27 adaptations of predators. In: *Carnivore Behavior, Ecology and Evolution*, vol. 2  
28 (J.L.Gittleman, ed.), pp. 393–428. Cornell University Press, Ithaca.  
29  
30  
31

32  
33 Biknevicus, A.R., and C.B Ruff. 1992. The structure of the mandibular corpus and its  
34 relationship to feeding behaviors in extant carnivorans. *J. Zool. (Lond.)* 228: 479–507.  
35  
36  
37

38  
39 Cheverud, J.M. 1982. Phenotypic, genetic, and environmental morphological  
40 integration in the cranium. *Evolution* 36: 499–516.  
41  
42  
43

44  
45 Cheverud, J.M. 1985. Quantitative genetics and developmental constraints on evolution  
46 by selection. *J. Theor. Biol.* 110: 155–172.  
47  
48  
49

50  
51 Christiansen P, and J.S. Adolfssen. 2005. Bite forces, canine strength and skull  
52 allometry in carnivores (Mammalia, Carnivora). *J Zool Lond* 266, 133–151.  
53  
54

55  
56 Christiansen, P., and S. Wroe. 2007. Bite forces and evolutionary adaptations to feeding  
57 ecology in carnivores. *Ecology* 88: 347–358.  
58  
59  
60

1  
2  
3 Conway-Morris, S. 2003. Life's solution: inevitable humans in a lonely universe.  
4  
5 Cambridge Univ. Press, Cambridge , UK .  
6

7  
8 Darwin, C. 1875. The Variation of Animals and Plants under Domestication, Vol. 1,  
9  
10 John Murray, London, UK.  
11

12  
13 Davis, D.D. 1964. The giant Panda. A morphological study on evolutionary  
14  
15 mechanisms. Fieldiana Zool. 3: 1–339.  
16

17  
18 Desutter-Grandcolas, L., F. Legendre, P. Grandcolas, T. Robillard, and J. Murienne.  
19  
20 2005. Convergence and parallelism: is a new life ahead of old concepts? Cladistics  
21  
22 21:51–61.  
23

24  
25  
26 Dong, W. 2008. Virtual cranial endocast of the oldest giant panda (*Ailuropoda microta*)  
27  
28 reveals great similarity to that of its extant relative. *Naturwissenschaften* 95:1079–1083.  
29

30  
31 Drake, A.G., and C.P. Klingenberg. 2008. The pace of morphological change: historical  
32  
33 transformation of skull shape in St Bernard dogs. *Proc. R. Soc. B* 275: 71–76.  
34

35  
36 Drake, A. G., and C. P. Klingenberg. 2010. Large-scale diversification of skull shape in  
37  
38 domestic dogs: Disparity and modularity. *Am Nat* 175: 289–301.  
39

40  
41 Dryden, I.L., and K. Mardia. 1998. *Statistical Shape Analysis*. Wiley, Chichester.  
42

43  
44 Escoufier, Y. 1973. Le traitement des variables vectorielles. *Biometrics* 29:  
45  
46 751–760.  
47  
48

49  
50 Ewer, R.F. 1973. *The Carnivores*. Cornell University Press, New York.  
51

52  
53 Eizirik, E., W. J. Murphy, K.-P. Koepfli, W. E. Johnson, J. W. Dragoo, R. K. Wayne,  
54  
55 and S. J. O'Brien. 2010. Pattern and timing of diversification of the mammalian order  
56  
57  
58  
59  
60

1  
2  
3 Carnivora inferred from multiple nuclear gene sequences. *Mol. Phylogenet. Evol.*  
4  
5 56:49–63.  
6

7  
8 Felsenstein, J.J. 1985. Phylogenies and the comparative method. *Am. Nat.* 125: 1–15.  
9

10  
11 Figueirido, B., and L. H. Soibelzon. 2010. Inferring palaeoecology in extinct  
12  
13 tremarctine bears (Carnivora, Ursidae) using geometric morphometrics. *Lethaia*, 43:  
14  
15 209–222.  
16

17  
18 Figueirido, B., J.A. Pérez-Claros, V. Torregrosa, A. Martín-Serra, and P. Palmqvist.  
19  
20 2010a. Demythologizing *Arctodus simus*, the ‘short-faced’, long-legged and predaceous  
21  
22 bear that never was. *J. Vert. Paleontol.* 30: 262–275.  
23

24  
25 Figueirido, B., F. J. Serrano-Alarcón, G. J. Slater, and P. Palmqvist. 2010b. Shape at the  
26  
27 cross-roads: homoplasy and history in the evolution of the carnivoran skull towards  
28  
29 herbivory. *Journal of Evolutionary Biology*, 23: 2579–2594.  
30  
31

32  
33 Figueirido, B., N. MacLeod, J. Krieger, M. De Renzi, J. A. Pérez-Claros, and P.  
34  
35 Palmqvist. 2011a. Constraint and adaptation in the evolution of carnivoran skull shape.  
36  
37 *Paleobiology*, 37:490-518.  
38

39  
40 Figueirido, B., P. Palmqvist, J.A. Pérez-Claros, and W. Dong. 2011b. Cranial shape  
41  
42 transformation in the evolution of the giant panda (*Ailuropoda melanoleuca*).  
43  
44 *Naturwissenschaften* 98, 107–116.  
45  
46

47  
48 Figueirido, B., F. J. Serrano-Alarcón, and P.Palmqvist. 2012. Geometric morphometrics  
49  
50 shows differences and similarities in skull shape between the red and giant pandas.  
51  
52 *Journal of Zoology*, 286: 293–302.  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Finarelli, J. A. and J.J. Flynn. 2006. Ancestral state reconstruction of body size in the  
4  
5 Caniformia (Carnivora, Mammalia): The effects of incorporating data from the fossil  
6  
7 record. *Syst Biol* 55: 301-313.  
8

9  
10 Finarelli, J.S., and J.J. Flynn. 2009. Brain-size evolution and sociality in Carnivora. *P.*  
11  
12 *Natl. Acad. Sci. USA* 106: 9345-9349.  
13

14  
15 Flynn, J.J., Nedbal, M., Dragoo, J. and Honeycutt, L. 2000. Whence the red panda?  
16  
17 *Mol. Phylogenet. Evol.* 17: 190–199.  
18

19  
20 Flynn, J.J., J.A. Finarelli, S. Zehr, J. Hsu and M. Nedbal. 2005. Molecular Phylogeny of  
21  
22 the Carnivora (Mammalia): assessing the impact of increased sampling on resolving  
23  
24 enigmatic relationships. *Syst. Biol.* 54: 317–337.  
25  
26

27  
28 Fox, M. L. 1963. Developmental abnormalities of the canine skull. *Can J Comp Med*  
29  
30 *Vet Sci.* 27: 219–222.  
31

32  
33 Futuyma, D.J. 2010. Evolutionary constraint and ecological consequences. *Evolution*  
34  
35 64: 1865-1884.  
36

37  
38 Galis, F., T. J. M. Van Dooren, H. Feuth, S. Ruinard, A. Witkam and, M. J. Steigenga.  
39  
40 2006. Extreme selection in humans against homeotic transformations of cervical  
41  
42 vertebrae. *Evolution* 60: 2643–2654.  
43  
44

45  
46 Gaubert P., and G. Veron. 2003. Exhaustive sample set among Viverridae reveals the  
47  
48 sister-group of felids: the Linsangs as a case of extreme morphological convergence  
49  
50 within Feliformia. *P Roy Soc B-Biol Sci* 270: 2523-2530.  
51  
52

1  
2  
3 Gidaszewski, N.A., M. Baylac, and C.P. Klingenberg. 2009. Evolution of sexual  
4 dimorphism of wing shape in the *Drosophila melanogaster* subgroup. *BMC Evol. Biol.*  
5 9: 110.  
6  
7

8  
9  
10 Gittleman, J.L. 1994. Are the Pandas Successful Specialists or Evolutionary Failures?  
11 *Bioscience* 44: 456–464.  
12

13  
14  
15 Gould, S.J. 1989. A developmental constraint in *Cerion*, with comments on the  
16 definition and interpretation of constraint in evolution. *Evolution* 43: 516–539.  
17

18  
19  
20 Gould, S.J. 2002. *The Structure of the Evolutionary Theory*. Harvard University Press,  
21 Harvard.  
22

23  
24  
25 Gould, S.J., and R. Lewontin. 1979. The spandrels of San Marco and the Panglossian  
26 paradigm: a critique of the adaptationist programme. *Proc. R. Soc. B* 205: 581–598.  
27

28  
29  
30 Goswami, A., Milne, N., and S. Wroe. 2011. Biting through constraints: cranial  
31 morphology, disparity and convergence across living and fossil carnivorous mammals.  
32 *Proc. Roy. Soc. B Biol. Sci.* 278, 1831–1839.  
33

34  
35  
36 Grenier, J. L., and R. Greenberg. 2005. A biogeographic pattern in sparrow bill  
37 morphology: Parallel adaptation to tidal marshes. *Evolution* 59:1588–1595.  
38

39  
40  
41 Hall, B.K. 2007. Homoplasy and homology: dichotomy or continuum? *J. Hum. Evol.*,  
42 52: 473–479.  
43

44  
45  
46 R. L. Hansen, M. M. Carr, C. J. Apanavicius, P. Jiang, H. A. Bissell, B. L. Gocinski, F.

47  
48  
49 Maury, M. Himmelreich, S. Beard, J. R. Ouellette, and A. J. Kouba. 2010. Seasonal

50  
51  
52 Shifts in Giant Panda Feeding Behavior: Relationships to Bamboo Plant Part

53  
54  
55 Consumption. *Zoo. Biol.*, 29: 470–483.  
56  
57  
58  
59  
60

1  
2  
3 Harvey, P. H., and M. D. Pagel. 1991. The comparative method in evolutionary biology.  
4  
5 Oxford Univ. Press, Oxford , UK .  
6

7  
8 Herrel, A., B. Vanhooydonck, and R. Van Damme. 2004. Omnivory in lacertid lizards:  
9  
10 adaptive evolution or constraint? *J. Evol. Biol.* 17: 974–984  
11

12  
13 Holliday, J.A., and S.J. Stepan. 2001. Evolution of hypercarnivory: the effect of  
14  
15 specialization on morphological and taxonomic diversity. *Paleobiology* 30: 108–128.  
16

17  
18 Jaekel, M., and D. B. Wake. 2007. Developmental processes underlying the evolution of  
19  
20 a derived foot morphology in salamanders. *Proc. Natl. Acad. Sci. USA* 104:20437–  
21  
22 20442.  
23

24  
25  
26 Joeckel, R.M. 1998. Unique frontal sinuses in fossil and living Hyaenidae (Mammalia,  
27  
28 Carnivora): description and interpretation. *Journal of Vertebrate Paleontology* 18: 627–  
29  
30 639.  
31

32  
33 Johnson, K. G., G. B. Schaller, and J. Hu. 1988. Comparative behavior of red and giant  
34  
35 pandas in the Wolong Reserve. China. *J. Mammal.*, 69: 552-564.  
36

37  
38 Johnson W., E. Eizirik, J. Pecon-Slatery, W. Murphy, A. Antunes, E. Teeling, and S  
39  
40 O’Brian. 2006. The late Miocene radiation of modern Felidae: a genetic assessment.  
41  
42 *Science*, 311:73–77.  
43

44  
45  
46 Klingenberg, C. P. 2009. Morphometric integration and modularity in configurations of  
47  
48 landmarks: Tools for evaluating a-priori hypotheses. *Evolution and Development*  
49  
50 11:405–421.  
51

52  
53  
54 Klingenberg, C. P. 2011. MorphoJ: an integrated software package for geometric  
55  
56 morphometrics. *Molecular Ecology Resources* 11: 353-357.  
57  
58  
59  
60

1  
2  
3 Klingenberg, C.P., and W. Ekau. 1996. A combined morphometric and phylogenetic  
4 analysis of an ecomorphological trend: pelagization in Antarctic fishes (Perciformes:  
5 Nototheniidae). *Biol. J. Linn. Soc.* 59: 143–177.  
6  
7

8  
9  
10 Klingenberg, C.P. and N.A. Gidaszewski. 2010. Testing and quantifying phylogenetic  
11 signals and homoplasy in morphometric data. *Syst. Biol.* 59: 245–261.  
12  
13

14  
15 Klingenberg, C. P., S. Dutke, S. Whelan, and M. Kim. 2012. Developmental plasticity,  
16 morphological variation and evolvability: a multilevel analysis of morphometric  
17 integration in the shape of compound leaves. *Journal of Evolutionary Biology*, 25: 115–  
18 129.  
19  
20  
21  
22  
23

24  
25 Koepfli K.P., S.M. Jenks, E. Eizirik, T. Zahirpour, B. Van Valkenburgh, and R.K.  
26 Wayne. 2006. Molecular systematics of the Hyaenidae: relationships of a relictual  
27 lineage resolved by a molecular supermatrix. *Mol. Phylogenet. Evol.*, 38: 603–620.  
28  
29  
30

31  
32 Koepfli, K.P., M.E. Gompper, E. Eizirik, C.C. Ho, L. Linden, J.E. Maldonado, and R.K.  
33 Wayne. 2007. Phylogeny of the Procyonidae (Mammalia: Carnivora): molecules,  
34 morphology and the great American interchange. *Mol. Phylogenet. Evol.*, 43: 1076–  
35 1095.  
36  
37  
38  
39  
40

41  
42 Koepfli, K.P., K.A. Deere, G.J. Slater, C. Begg, K. Begg, L. Grassman, M. Lucherini,  
43 G. Veron, and R.K. Wayne. 2008. Multigene phylogeny of the Mustelidae: resolving  
44 relationships, tempo and biogeographic history of a mammalian adaptive radiation  
45 *BMC Biol.*, 6: 10.  
46  
47  
48  
49  
50

51  
52 Krause J., T. Unger, A. Noçon, A.S. Malaspinas, S.O. Kolokotronis, M. Stiller, L.  
53 Soibelzon, H. Spriggs, P.H. Dear, A.W. Briggs, S.C. Bray, S.J. O'Brien, G. Rabeder, P.  
54 Matheus, A. Cooper, M. Slatkin, S. Pääbo, M. Hofreiter. 2008. Mitochondrial genomes  
55  
56  
57  
58  
59  
60

- 1  
2  
3 reveal an explosive radiation of extinct and extant bears near the Miocene-Pliocene  
4  
5 boundary. *BMC Evol Biol* 2008, 8:220.  
6  
7  
8 Kruk, H. 1972. *The spotted hyena: A study of predation and social behavior*. Univ. of  
9  
10 Chicago Press, Chicago. 335pp.  
11  
12  
13 Langerhans, R. B., J. H. Knouft, and J. B. Losos. 2006. Shared and unique features of  
14  
15 diversification in Greater Antillean *Anolis* ecomorphs. *Evolution* 60:362–369.  
16  
17  
18 Laurin, M. 2004. The evolution of body size, Cope’s rule and the origin of amniotes.  
19  
20 *Syst. Biol.* 53: 594–622.  
21  
22  
23 Ligon J.D. 1993. The role of phylogenetic history in the evolution of contemporary  
24  
25 avian mating and parental care systems. *Curr Ornithol* 10: 1-46.  
26  
27  
28  
29 Losos, J. 2011. Convergence, adaptation and constraint. *Evolution* 65: 1827-1840.  
30  
31  
32 Low, I.M., Z.Y. Che, and B.A. Latella . 2006. Mapping the structure, composition and  
33  
34 mechanical properties of bamboo. *J. Materials Research* 21:1969-1976.  
35  
36  
37 MacLeod, N. 2001. The role of phylogeny in quantitative paleobiological data analysis.  
38  
39 *Paleobiology* 27: 226–240.  
40  
41  
42 MacLeod, N., 2006, Data blocks and partial least squares analysis: *Palaeontological*  
43  
44 *Association Newsletter*, v. 63, p. 36–48.  
45  
46  
47  
48 Maddison, W.P. 1991. Squared-change parsimony reconstructions of ancestral states for  
49  
50 continuous-valued characters on a phylogenetic tree. *Syst. Zool.* 40: 304–314.  
51  
52  
53 Maddison, W. P. and D.R. Maddison. 2011. *Mesquite: a modular system for*  
54  
55 *evolutionary analysis*. Version 2.75 <http://mesquiteproject.org>  
56  
57  
58  
59  
60

1  
2  
3 Madsen, O., M. Scally, C. J. Douady, D. J. Kao, R. W. DeBry, R. Adkins, H. M.

4 Amrine, M. J. Stanhope, W. W. De Jong, and M. S. Springer. 2001. Parallel adaptive  
5 radiations in two major clades of placental mammals. *Nature* 409:610–614.  
6  
7

8  
9  
10 Maynard-Smith, J., R. Burian, S. Kauffman, P. Alberch, J. Campbell, B. Goodwin, R.  
11 Lande, D. Raup, and L. Wolpert. 1985. Developmental constraints and evolution. *Q.*  
12 *Rev. Biol.* 6:265–287.  
13  
14

15  
16  
17 Meloro C., Raia P., Carotenuto F., Cobb S. (2011) Phylogenetic signal, function and  
18 integration in the subunits of the carnivoran mandible. *Evolutionary Biology* 38: 465-  
19 475.  
20  
21  
22  
23

24  
25 Meyer, A. 1999. Homology and homoplasy: the retention of genetic programmes. In  
26 homology: 141–157. Bock, G. (Ed.). Chichester: Wiley and Son.  
27  
28

29  
30 **Monteiro, L.R. 1999. Multivariate regression models and geometric morphometrics: the**  
31 **search for causal factors in the analysis of shape. *Syst. Biol.* 48: 192–199.**  
32  
33

34  
35  
36 Nevo, E. 2001. Evolution of genome–phenome diversity under environmental stress.  
37 *Proc. N.Y. Acad. Sci.* 98:623–6240.  
38  
39

40  
41 Nowak, R. M., and E. P. Walker. 1999. Walker's mammals of the world. 6th ed. JHU  
42 Press.  
43  
44

45  
46 **Oldfield, C. C., C. R. McHenry, P. D. Clausen, U. Chamoli, W. C. H. Parr, D. Stynder,**  
47 **and S. Wroe. 2012. Finite element analysis of ursid cranial mechanics and the**  
48 **prediction of feeding behaviour in the extinct giant *Agriotherium africanum*. *J.Zool.***  
49 **(Lond), 286: 163-170.**  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Olson, M. E. 2012. The developmental renaissance in adaptationism. *Trends in Ecology*  
4 and *Evolution* 27(5): 278–287  
5  
6

7  
8 Palmqvist, P., A. Arribas, and B. Martínez-Navarro. 1999. Ecomorphological analysis  
9 of large canids from the lower Pleistocene of southeastern Spain. *Lethaia*, 32: 75-88.  
10  
11

12  
13 Palmqvist, P., Martínez-Navarro, B., Pérez-Claros, J.A., Torregrosa, V., Figueirido, B.,  
14 Jiménez-Arenas, J.M., Patrocinio Espigares, M., Ros-Montoya, S. and, De Renzi, M.  
15 2011. The giant hyena *Pachycrocuta brevirostris*: Modelling the bone-cracking behavior  
16 of an extinct carnivore. *Quaternary International*, 243: 61-79.  
17  
18  
19  
20  
21

22  
23 Patou M.L., P.A. Mclenachan, C.G Morley, A. Couloux, C. Cruaud, A.P. Jennings,  
24 and G. Veron. 2009. Molecular phylogeny of the Herpestidae (Mammalia, Carnivora)  
25 with a special emphasis on the Asian Herpestes. *Mol. Phylogenet. Evol.* 53: 69-80.  
26  
27  
28  
29

30  
31 Polly, P.D. 2001. Paleontology and the comparative method: ancestral node  
32 reconstructions versus observed node values. *Am. Nat.* 157: 596–609.  
33  
34

35  
36 Polly, P.D. 2008. Adaptive Zones and the Pinniped Ankle: a 3D Quantitative Analysis  
37 of Carnivoran Tarsal Evolution. In: *Mammalian Evolutionary Morphology: A Tribute to*  
38 *Frederick S. Szalay* (E.Sargis and M.Dagosto, eds), pp. 165–194. Springer, Dordrecht,  
39 The Netherlands.  
40  
41  
42  
43  
44

45 Reid, D.G., J. Hu, and Y. Huang. 1991. Ecology of the red panda in the Wolong  
46 Reserve, China. *J. Zool. (Lond.)*, 225: 347–364.  
47  
48

49  
50  
51 Rohlf, F.J. 2002. Geometric morphometrics and phylogeny. In: *Morphology, Shape, and*  
52 *Phylogeny* (N.MacLeod and P.L.Forey, eds), pp. 175–193. Taylor and Francis, London.  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Rohlf, F.J. 2008. TpsDig, ver. 2.11. Digitize landmarks and outlines. Department of  
4 Ecology and Evolution, State University of New York at Stony Brook [computer  
5 program and documentation].  
6  
7  
8

9  
10 Sacco, T. and Van Valkenburgh, B. 2004. Ecomorphological indicators of feeding  
11 behaviour in bears (Carnivora: Ursidae). *J. Zool. (Lond.)* 263: 41–54.  
12  
13

14  
15 Schaller G.B., H. Jinchu, P. Wenshi, and Z. Jing. 1985. *The giant pandas of Wolong.*  
16  
17  
18  
19 Chicago: University of Chicago Press.

20  
21 Schluter, D. 1988. Estimating the form of natural selection on a quantitative trait.  
22  
23 *Evolution* 42:849–861.  
24

25  
26 Schluter, D. 2000. *The ecology of adaptive radiation.* Oxford Univ. Press, Oxford , UK  
27

28  
29 Schlueter, D. 2009. Evidence for ecological speciation and its alternative. *Science* 323:  
30  
31 737–741.  
32

33  
34 Schmidt-Nielsen, K. 1984. *Scaling: Why is animal size so important?* Cambridge:  
35  
36 Cambridge University Press.  
37

38  
39 Schwenk, K. 1995. A utilitarian approach to evolutionary constraint. *Zoology* 98: 251–  
40  
41 262.  
42

43  
44 Schwenk, K., and G. P. Wagner. 2003. Constraint. Pp. 52-61. In: *Key Words and*  
45  
46 *Concepts in Evolutionary Developmental Biology.* B. K. Hall and W. M. Olson (eds.).  
47  
48 Harvard University Press, Cambridge.  
49

50  
51  
52 Schwenk, K., and G. P. Wagner (2004) The relativism of constraints on phenotypic  
53  
54 evolution. Pp. 390-408. In: *Phenotypic Integration: Studying the Ecology and Evolution*  
55  
56  
57  
58  
59  
60

1  
2  
3 of Complex Phenotypes. M. Pigliucci and K. Preston (eds.). Oxford Univ. Press,  
4  
5 Oxford.

6  
7  
8 Slater, G.J., L.J. Harmon, and M. Alfaro. 2012. Integrating fossils with molecular  
9  
10 phylogenies improves inference of trait evolution. *Evolution* 66-12: 3931–3944.

11  
12  
13 Stayton, C.T. 2006. Testing hypotheses of convergence with multivariate data:  
14  
15 morphological and functional convergence among herbivorous lizards. *Evolution* 60:  
16  
17 824–841.

18  
19  
20  
21 Stayton, C. T. 2008. Is convergence surprising? An examination of the frequency of  
22  
23 convergence in simulated datasets. *J. Theor. Biol.* 252:1–14.

24  
25  
26 Stockard. C. R. 1941. *The Genetic and Endocrine Basis for Differences in Form and*  
27  
28 *Behavior.* Wistar Institute, Philadelphia.

29  
30  
31 Tanner, J. B., E. R. Dumont, S. T. Sakai, Lundrigan, B. L., and K. E. Holekamp. 2008.  
32  
33 Of arcs and vaults: the biomechanics of bone-cracking in spotted hyenas (*Crocuta*  
34  
35 *crocuta*). *Biological Journal of the Linnean Society*, 95: 246–255.

36  
37  
38  
39 Tedford, R.H., X. Wang, B.E. Taylor. 2009. Phylogenetic systematics of the North  
40  
41 American fossil Caninae. *Bull. Am. Mus. Nat. Hist* 325, 1–218.

42  
43  
44 Thomas, R. D. K., and W. E. Reif. 1993. The skeleton space: a finite set of organic  
45  
46 designs. *Evolution* 47, 341–360.

47  
48  
49 Timm, N. H. 2002, *Applied multivariate analysis.* New York, Springer.

50  
51  
52 Tseng, Z.J. 2009. Cranial function in a late Miocene *Dinocrocuta gigantea* (Mammalia:  
53  
54 Carnivora) revealed by comparative finite element analysis. *Biological Journal of the*  
55  
56 *Linnean Society* 96:51-67.  
57  
58  
59  
60

- 1  
2  
3 Tseng, Z. J. 2012. Connecting Hunter–Schreger Band microstructure to enamel  
4 microwear features: New insights from durophagous carnivores. *Acta Palaeontologia*  
5 *Polonica* 57: 473–484.  
6  
7  
8  
9  
10 Tseng, Z.J., and X. Wang. 2011. Do convergent ecomorphs evolve through convergent  
11 morphological pathways? Cranial shape evolution in fossil hyaenids and borophagine  
12 canids (Carnivora, Mammalia). *Paleobiology* 37:470-489.  
13  
14  
15  
16  
17 Tseng Z.J., and W. Binder. 2010. Mandibular biomechanics of *Crocuta crocuta*, *Canis*  
18 *lupus*, and the late Miocene *Dinocrocuta gigantea* (Carnivora, Mammalia). *Zool. J.*  
19 *Linn. Soc-Lond*, 158: 683-696.  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60
- Turnbull, W.D. 1976. Mammalian masticatory apparatus. *Fieldiana: Geol.* 18: 152–356.
- Van Valkenburgh, B. 1991. Iterative evolution of hypercarnivory in canids (Mammalia: Carnivora): evolutionary interactions among sympatric predators. *Paleobiology* 17: 340–362.
- Van Valkenburgh, B. 1999. Major patterns in the history of carnivorous mammals. *Annu. Rev. Earth Planet. Sci.* 27: 463–493.
- Van Valkenburgh, B. 2007. Déjà vu: the evolution of feeding morphologies in the Carnivora. *Integr. Comp. Biol.* 47: 147–163.
- Van Valkenburgh, B., T. Sacco, and X. Wang. 2003. Pack hunting in Miocene borophagine dogs: evidence from craniodental morphology and body size. *Bull. Am. Mus. Nat. Hist.* 279: 147–162.
- Wagner, G.P. 1988. The influence of variation and of developmental constraints on the rate of multivariate phenotypic evolution. *J. Evol. Biol.* 1: 45–66.

1  
2  
3 Wagner, G.P. 1996. Homologues, natural kinds and the evolution of modularity. Am.  
4  
5 Zool. 36: 36-43.  
6

7  
8 Wagner, G.P. and Altenberg, L. 1996. Complex adaptations and the evolution of  
9  
10 evolvability. Evolution, 50: 967-976.  
11

12  
13 Wagner, G.P., Pavlicev, M. and Cheverud, J.M. 2007. The road to modularity. Nat.  
14  
15 Rev. Genet. 8: 921-931.  
16

17  
18 Wake, D. 1991. Homoplasy: The Result of Natural Selection, or Evidence of Design  
19  
20 Limitations? Am. Nat. 138:543-567.  
21

22  
23 Wang, X., R. H. Tedford, and B. E. Taylor. 1999. Phylogenetic systematics of the  
24  
25 Borophaginae (Carnivora: Canidae). Bulletin of the American Museum of Natural  
26  
27 History, 243: 1-391  
28

29  
30  
31 Wegst, U. G. K. and M. F. Ashby. 2004. The mechanical efficiency of natural materials,  
32  
33 Philosophical Magazine, 21: 2167-2186.  
34

35  
36  
37 Wei, F., Z. Feng, Z. Wang, A. Zhou, and J. Hu. 1999. Use of the nutrients in bamboo by  
38  
39 the red panda (*Ailurus fulgens*). J. Zool. (Lond.), 248: 535–541.  
40

41  
42 Werdelin, L. 1989. Constraint and adaptation in the bone-cracking canid *Osteoborus*  
43  
44 (Mammalia: Canidae). Paleobiology 15: 387–401.  
45

46  
47 Werdelin, L. and N. Solounias. 1991. The Hyaenidae: taxonomy, systematic and  
48  
49 evolution. Fossils Strata 30: 1-104.  
50

51  
52 Winter, W., de, and C. E. Oxnard. 2001. Evolutionary radiations and convergences in  
53  
54 the structural organization of mammalian brains. Nature 409:710–714.  
55  
56  
57  
58  
59  
60

1  
2  
3 Wroe, S. 2008. High-resolution 3-D computer simulation of feeding behaviour in  
4  
5 marsupial and placental lions. *J. Zool.* 274:332-339.  
6  
7

8 Wroe, S. and N. Milne. 2007. Convergence and remarkably consistent constraint in the  
9  
10 evolution of carnivore skull shape. *Evolution* 61: 1251–1260.  
11  
12

13 Wroe, S., C. McHenry, and J. Thomason. 2005. Bite club: Comparative bite force in big  
14  
15 biting mammals and the prediction of predatory behaviour in fossil taxa. *Proc. R. Soc. B*  
16  
17 272: 619-625.  
18  
19

20 Wroe, S., P. Clausen, C. R. McHenry, K. Moreno, and E. Cunningham. 2007. Computer  
21  
22 simulation of feeding behaviour in the thylacine and dingo: a novel test for convergence  
23  
24 and niche overlap. *Proc. R. Soc. B* 274: 2819–2828.  
25  
26  
27

28 Zelditch, M.L., Swinderski, D.L., Sheets, H.D. and Fink, W.L. 2004. *Geometric*  
29  
30 *Morphometrics for Biologist: A Primer*. Elsevier Academic Press, New York/London.  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**Table 1.** Sample size of crania and mandibles used in this study. Species highlighted in light grey correspond to bone-crackers and those highlighted in dark grey correspond to bamboo-feeders. We considered among the bone-cracking specialists: *Dinocrocuta gigantea*† (family Percrocutidae; Werdelin and Solunias 1991) following Tseng (2009) and Tseng and Binder (2010); *Crocuta crocuta*, *Hyaena hyaena*, *Parahyaena brunea* and *Adcrocuta eximia*† (family Hyaenidae) following Nowak (1991) and Tseng and Wang (2011); *Borophagus secundus*†, *Epicyon saevus*† and *Aelurodon spp.* † (family Canidae, subfamily Borophaginae†) following Wang et al. (1999) and Tseng and Wang (2011). As bamboo-feeders we classified the red or lesser panda *A. fulgens* (family Ailuridae) and the giant panda *A. melanoleuca* (family Ursidae) following Johnson et al. (1988). Daggers denote extinct taxa.

FAMILY	SPECIES (Abbreviations)	N (mandible)	N (cranium)
URSIDAE	<i>Ailurpoda melanoleuca</i> (Ame)	13	15
URSIDAE	<i>Tremarctos ornatus</i> (To)	5	7
URSIDAE	<i>Ursus arctos</i> (Ua)	12	16
URSIDAE	<i>Ursus americanus</i> (Um)	5	8
FELIDAE	<i>Acinonyx jubatus</i> (Aju)	3	2
FELIDAE	<i>Felis serval</i> (Fser)	1	1
FELIDAE	<i>Felis temnicki</i> (Ftem)	1	1
FELIDAE	<i>Felis tigrina</i> (Ftig)	1	1
FELIDAE	<i>Lynx lynx</i> (Lly)	2	9
FELIDAE	<i>Neofelis nebulosa</i> (Nneb)	3	9
FELIDAE	<i>Puma concolor</i> (Pcon)	4	9
FELIDAE	<i>Panthera leo</i> (Pleo)	4	6
FELIDAE	<i>Panthera onca</i> (Ponc)	5	10
FELIDAE	<i>Panthera pardus</i> (Ppar)	5	3
FELIDAE	<i>Panthera tigris</i> (Ptig)	1	1
FELIDAE	<i>Panthera uncia</i> (Punc)	1	1
EUPLERIDAE	<i>Cryptoprocta ferox</i> (Cfer)	3	3
HERPESTIDAE	<i>Crossarcus obscurus</i> (Cros)	1	1
HERPESTIDAE	<i>Herpestes ichenumon</i> (Her)	3	4
VIVERRIDAE	<i>Genetta genetta</i> (Gen)	5	5
AILURIDAE	<i>Ailurus fulgens</i> (Aful)	12	12
PROCYONIDAE	<i>Bassaricyon medius</i> (Baa)	3	3
PROCYONIDAE	<i>Bassariscus sumichastri</i> (Bas)	3	3
PROCYONIDAE	<i>Procyon lotor</i> (Plot)	8	7

1				
2				
3	PROCYONIDAE	<i>Potos flavus</i> (Pfla)	10	10
4	MUSTELIDAE	<i>Eira barbara</i> (Eir)	8	8
5	MUSTELIDAE	<i>Gulo gulo</i> (Gul)	7	8
6	MUSTELIDAE	<i>Meles meles</i> (Mel)	6	9
7	MUSTELIDAE	<i>Mellivora capensis</i> (Mell)	5	5
8	MUSTELIDAE	<i>Taxidea taxus</i> (Tax)	5	5
9				
10	HYAENIDAE	<i>Crocuta crocuta</i> (Ccr)	9	9
11	HYAENIDAE	<i>Hyaena hyaena</i> (Hhy)	11	8
12	HYAENIDAE	<i>Hyaena brunnea</i> (Hyb)	9	8
13				
14	HYAENIDAE	<i>Ictitherium sp.</i> (Ict) †	1	3
15	HYAENIDAE	<i>Adcrocuta eximia</i> (Adc) †	-	3
16				
17	HYAENIDAE	<i>Hyaenictitherium wongi</i> (Hyc) †	1	4
18	HYAENIDAE	<i>Dinocrocuta gigantea</i> (Din) †	1	2
19				
20	CANIDAE	<i>Alopex lagopus</i> (Ala)	10	10
21	CANIDAE	<i>Cuon alpinus</i> (Cal)	11	8
22	CANIDAE	<i>Chrysocyon brachyurus</i> (Cbr)	5	1
23	CANIDAE	<i>Canis latrans</i> (Cla)	8	10
24	CANIDAE	<i>Canis lupus</i> (Clu)	12	10
25	CANIDAE	<i>Canis mesomelas</i> (Cme)	10	10
26	CANIDAE	<i>Canis aureus</i> (Cau)	8	9
27	CANIDAE	<i>Lycan pictus</i> (Lpic)	9	10
28	CANIDAE	<i>Speothos venaticus</i> (Sve)	3	3
29	CANIDAE	<i>Vulpes corsac</i> (Vco)	4	10
30	CANIDAE	<i>Vulpes velox</i> (Vve)	8	1
31	CANIDAE	<i>Vulpes vulpes</i> (Vvul)	14	9
32				
33	CANIDAE	<i>Borophagus secundus</i> (Bsec) †	1	4
34	CANIDAE	<i>Epicyon saevus</i> (Esa) †	1	1
35				
36	CANIDAE	<i>Tomarctus hipophaga</i> (Tom) †	1	1
37	CANIDAE	<i>Aelurodon kilpatricki</i> (Akil) †	-	1
38	CANIDAE	<i>Aelurodon stirtoni</i> (Astir) †	1	1
39	CANIDAE	<i>Aelurodon ferox</i> (Afer) †	5	1
40				
41	CANIDAE	<i>Desmocyon thompsoni</i> (Des) †	2	1
42	CANIDAE	<i>Carpocyon compressus</i> (Car) †	1	0
43				
44				
45				
46				
47				
48				
49				
50				
51				
52				
53				
54				
55				
56				
57				
58				
59				
60				

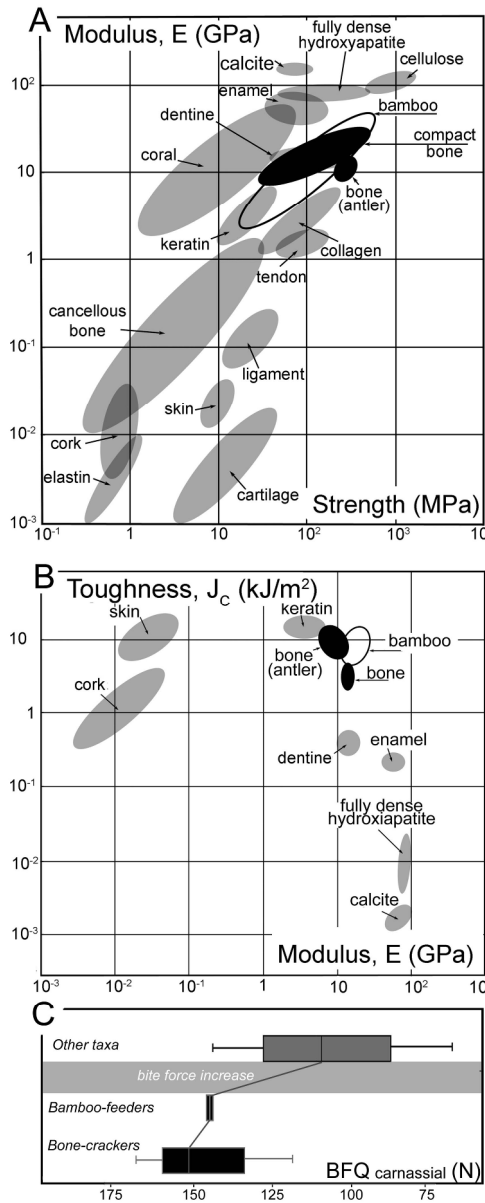


Figure 1. Material properties of bone and bamboo and bite force in bone-cracking and bamboo-feeding carnivorans. A, Bivariate plot showing Young’s Modulus ( $E$ ; measured in GPa) on the strength ( $\sigma$ ; measured in MPa) of bone and bamboo relative to other natural materials (redrawn from Wegst and Ashby [2004]). B, Bivariate plot depicting the toughness of bone and bamboo ( $J_c$ ; measured in KJ/m<sup>2</sup>) on Young’s Modulus ( $E$ ; measured in GPa) relative to other natural materials (redrawn from Wegst and Ashby [2004]). C, Box-plot showing the bite force quotient (bite force for each species in relation to its body mass) at the carnassials (BFQcarnassial) estimated by the dry skull method in the living bone-crackers, bamboo-feeders and other taxa included in our sample. The vertical line inside each box is the median. Box length is the interquartile range (IQR) and shows the difference between the 75th and 25th percentiles (data from Christiansen and Wroe 2007). Horizontal bars enclose values of 5–95%.

177x439mm (300 x 300 DPI)

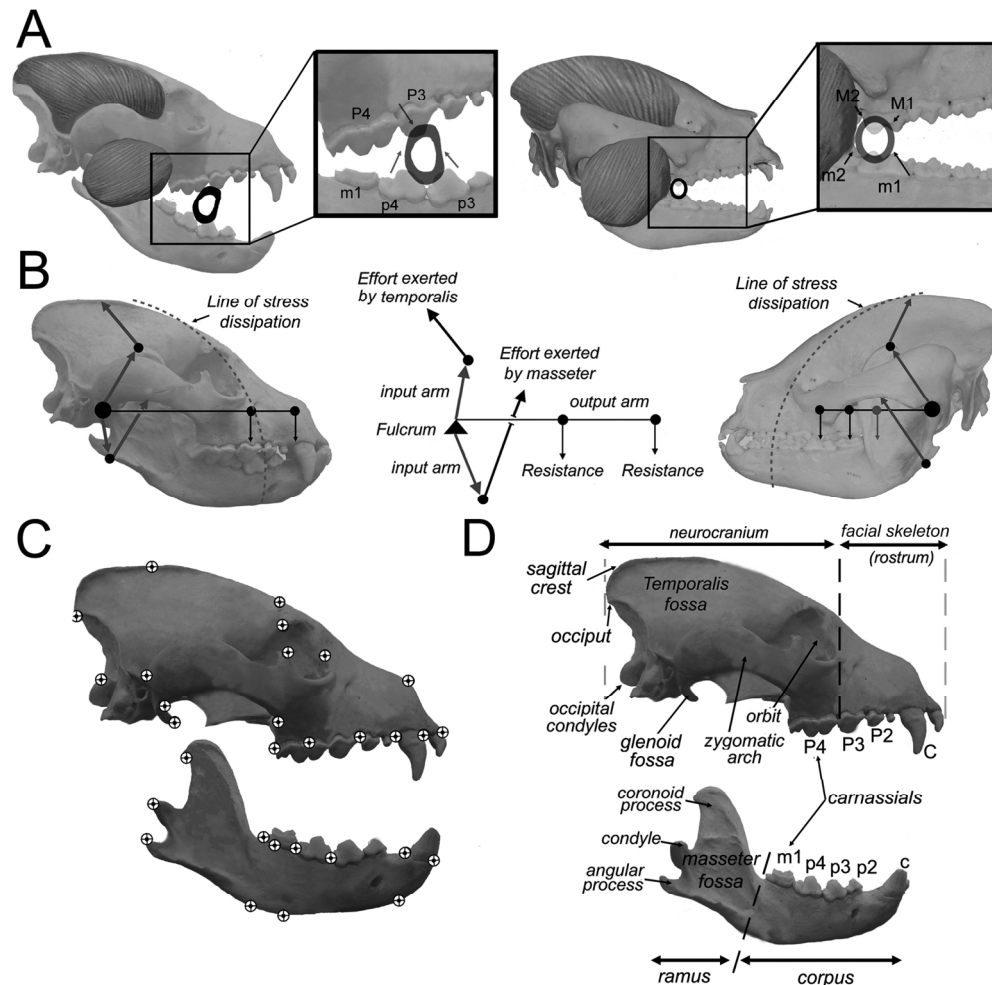


Figure 2. Skull biomechanics in durophagous carnivorans. A, Main muscles and dentition involved in the mastication practice, exemplified in a skull of spotted hyena (*C. crocuta*) and in a skull of giant panda (*A. melanoleuca*) with an hypothetical cross-section of a bone and bamboo stem, respectively. B, Masticatory apparatus modelled as a third class lever system showing the combined effort of the temporalis and masseter muscles as a function of their input arms, applied in a point between the fulcrum (temporomandibular joint) and the resistance exerted at the main teeth involved in processing food. The line of stress dissipation is a circle drawn with its centre at the fulcrum and with a radius of length from the fulcrum to P3 in the case of spotted hyaena and to M1 in the case of the giant panda (Werdelin 1989). C, Landmarks used in the morphometric analysis of skull function. D, Morphological key aspects in function recovered in the morphometric analyses. Abbreviations: C, upper canine; M, upper molar; P, upper premolar; c, lower canine; m, lower molar; p, lower premolar. The numbers indicate the position of each type of teeth. Skulls not to scale.

163x162mm (300 x 300 DPI)

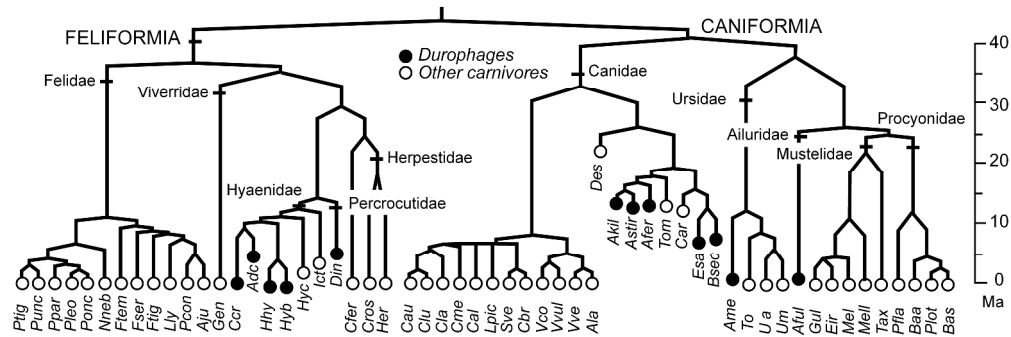


Figure 3. Phylogenetic tree used in this study. The main topology of the tree (i.e., interfamilial relationships) was arranged following Flynn et al. (2005), with the only exception of the unresolved polytomy for the family Viverridae. In this case, we follow Gaubert and Veron (2003), Finarelli and Flynn (2009) and Eizirik et al. (2010) and we considered the Viverridae (s.s.) as the stem group to Hyaenidae plus Herpestidae. To assess the intrafamilial relationships, we followed different published sources: Johnson et al. (2006) for the family Felidae; Koepfli et al. (2006) for Hyaenidae; Koepfli et al. (2007) for the family Procyonidae; Koepfli et al. (2008) for Mustelidae; Patou et al. (2009) for the relationships of Herpestidae; Krause et al. (2008) for Ursidae; and Bardeleben et al. (2005), Wang et al. (1999) and Tedford et al. (2009) for Canidae. The extinct family Percrocutidae was considered as a sister group of Hyaenidae following Werdelin and Solunias (1991) as were as the phylogenetic relationships of "stem" Hyaenidae (*Ictitherium*, *Hyaenictitherium*, *Adcrocuta*). We incorporated branch lengths as million years before present following the procedure described in Finarelli and Flynn (2006). See Table 1 for species abbreviations.

1257x416mm (96 x 96 DPI)

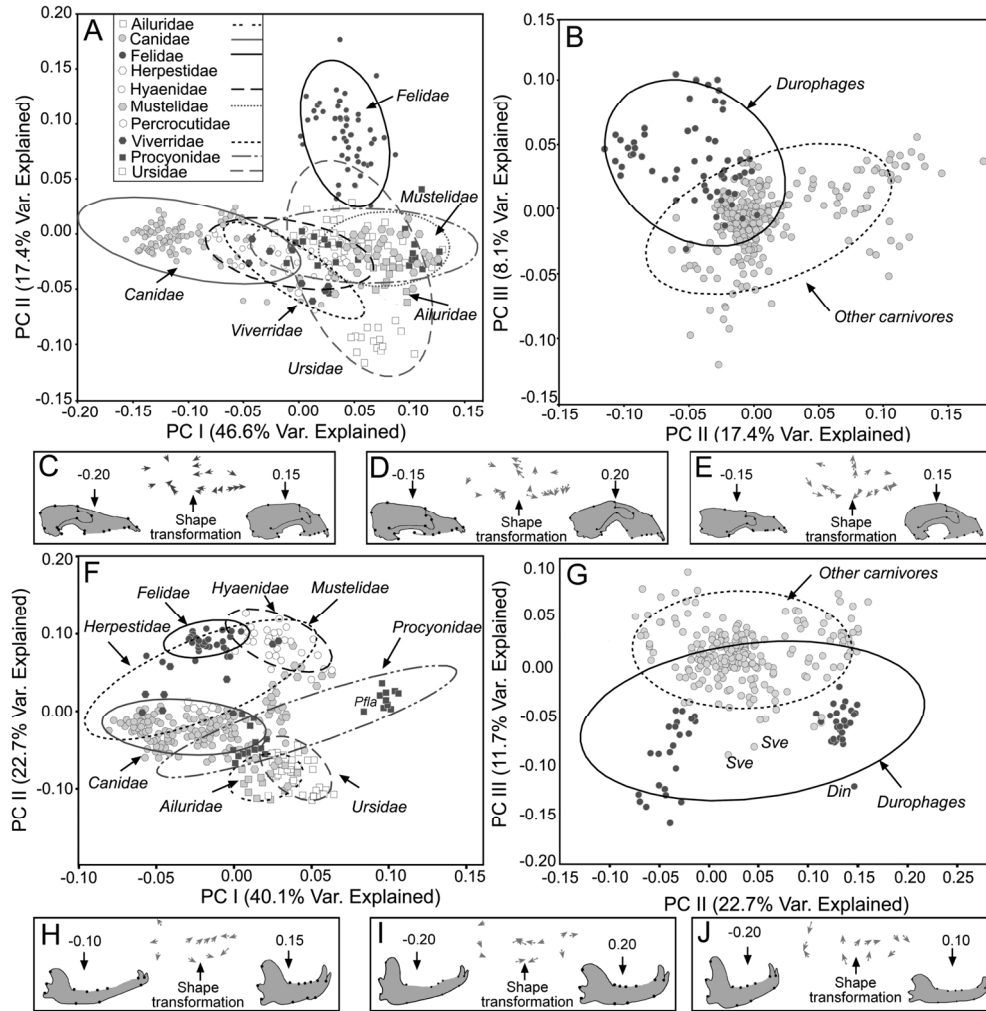


Figure 4. The phenotypic spaces derived from skull shape. A, Pairwise plot derived from the first two PCs of cranial analysis. B, Pairwise plot obtained from PCII and PCIII of cranial analysis. Shape variation accounted for by PCI (C), PCII (D) and PCIII (E) of cranial analysis. F, Pairwise plot depicted from the first two PCs of mandible analysis. D, Pairwise plot derived from PCII and PCIII of mandible analysis. Shape variation accounted for by PCI (H), PCII (I) and PCIII (J) of mandible analysis. The 95% confidence ellipses are also shown. For species abbreviations, see Table 1.

162x165mm (300 x 300 DPI)

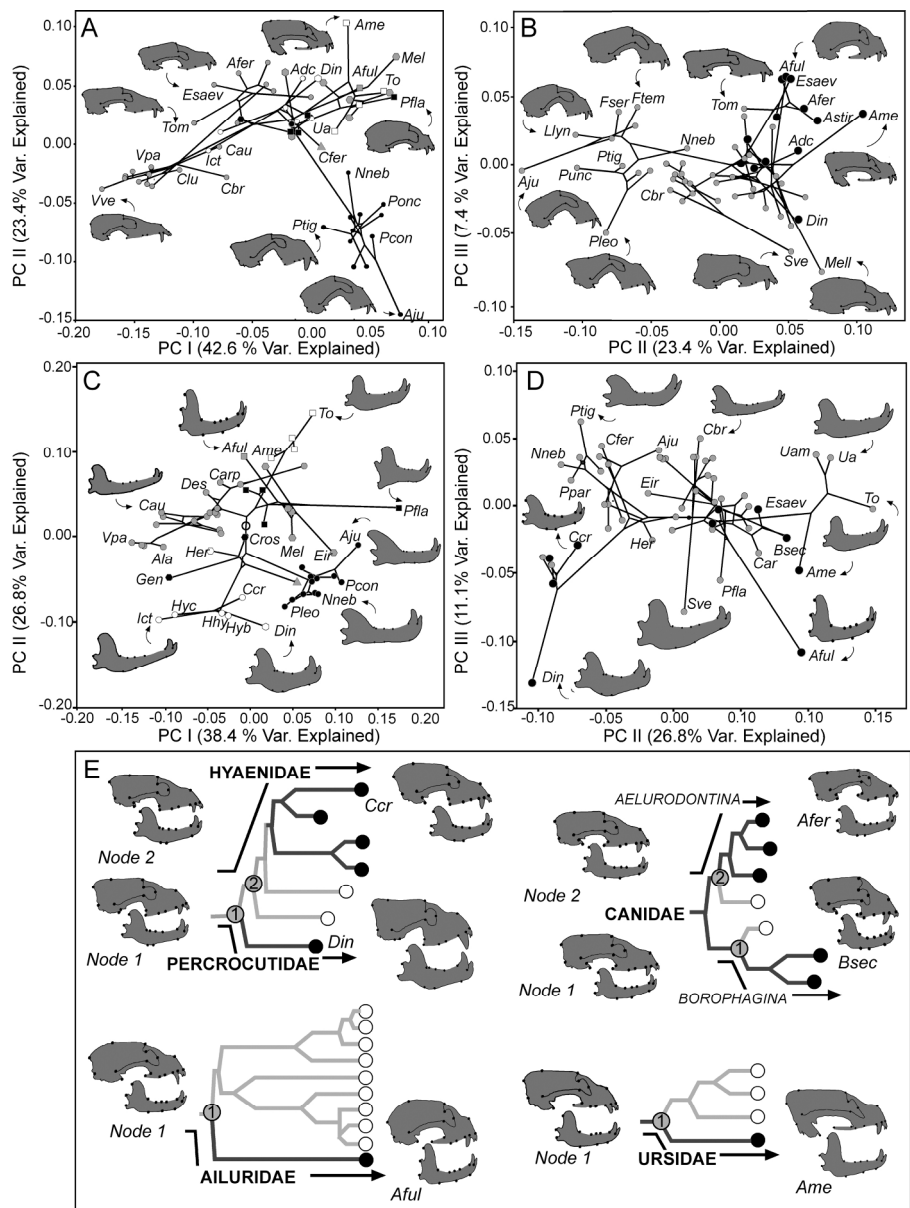


Figure 5. Phylomorphospaces and evolutionary trajectories of skull shape transformation. A, Projection of the phylogenetic tree topology onto the phenotypic space depicted from the first two PCs of cranium analysis. B, projection of the phylogenetic tree onto the phenotypic space depicted from PCII and PCIII of cranium analysis. C, Projection of the phylogenetic tree topology onto the phenotypic space depicted from the first two PCs of mandible analysis. D, projection of the phylogenetic tree onto the phenotypic space depicted from PCII and PCIII of mandible analysis. E, hypothetical evolutionary pathways followed by durophages, represented as the shape change from the reconstructed skull shape of the ancestors for each family to living species. Symbols as in Figure 4.

212x282mm (300 x 300 DPI)

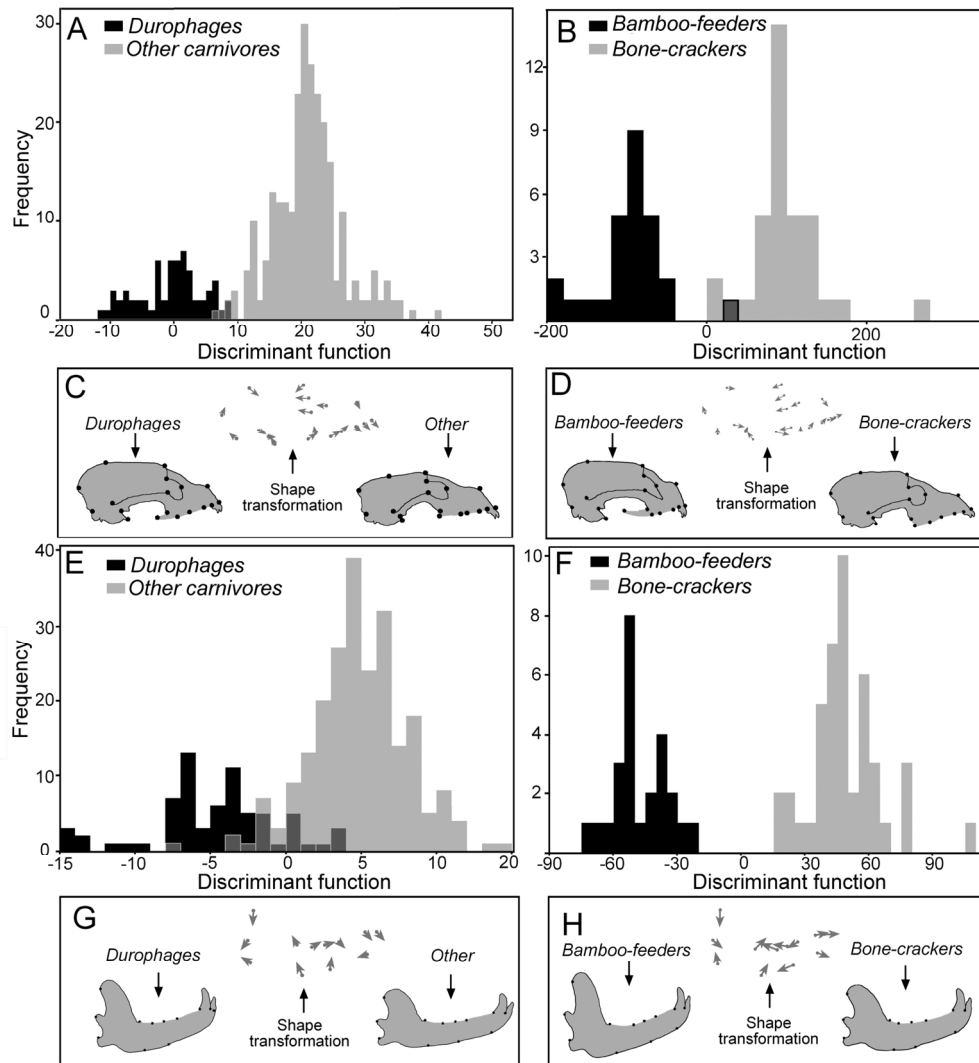


Figure 6. Discriminant analyses. A, Histogram showing the distribution of the specimens according to their scores on the discriminant function adjusted from cranial morphology to separate between durophages and other carnivorans. B, Histogram showing the distribution of the specimens according to their scores on the discriminant function adjusted from cranial morphology to separate between bone-crackers and bamboo-feeders. C, morphological variation accounted for by the discriminant function adjusted from cranial morphology to separate between durophages and other carnivorans. D, morphological variation accounted for by the discriminant function adjusted from cranial morphology to separate between bone-crackers and bamboo-feeders. E, Histogram showing the distribution of the specimens according to their scores on the discriminant function adjusted from mandible morphology to separate between durophages and other carnivorans. F, Histogram showing the distribution of the specimens according to their scores on the discriminant function adjusted from mandible morphology to separate between bone-crackers and bamboo-feeders. G, morphological variation accounted for by the discriminant function adjusted from mandible morphology to separate between durophages and other carnivorans. H, morphological variation accounted for by the discriminant function adjusted from mandible morphology to separate between bone-crackers and bamboo-feeders. Dark-grey bars represent the overlapping areas between groups.

164x174mm (300 x 300 DPI)

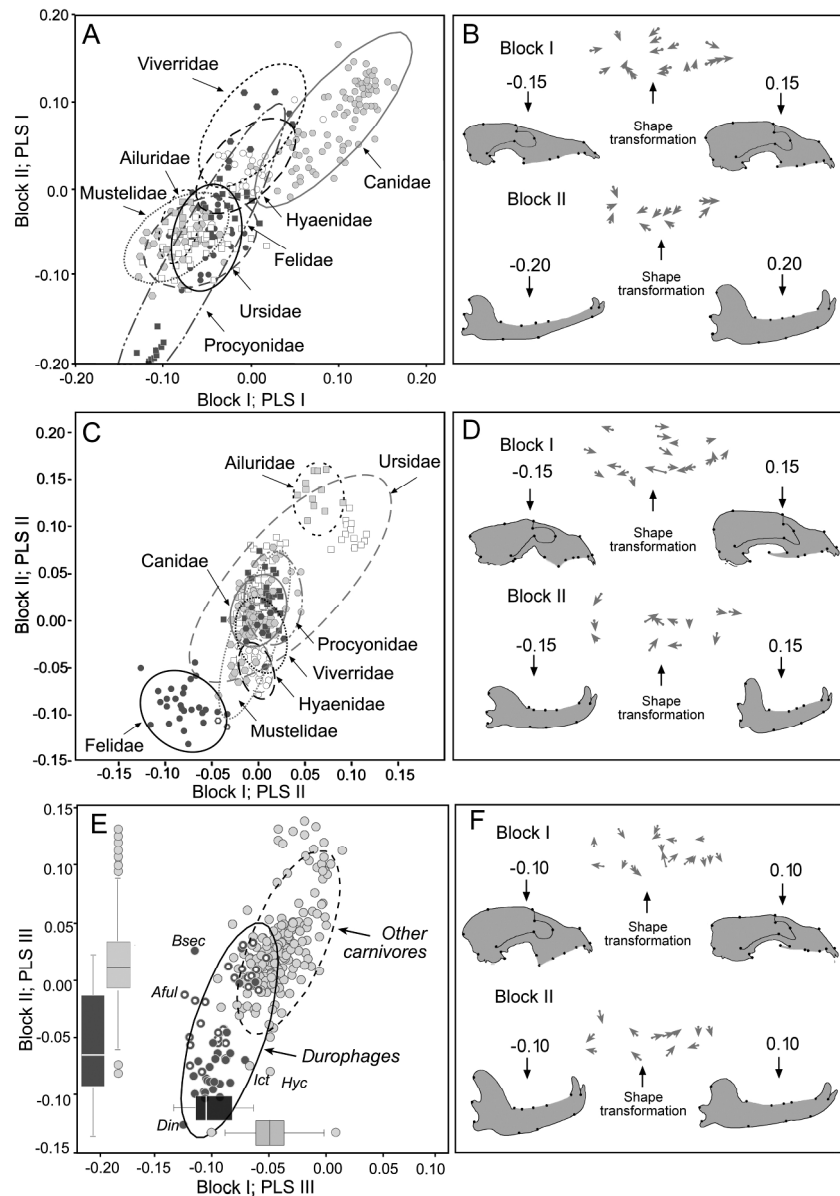


Figure 7. PLS-analysis performed between cranium (Block I) and mandible (Block II) shape. A, Pairwise plot derived from the first pair of PLS axes. B, Shape covariation between both blocks of variables accounted for by the first pair of PLS-axes. C, Pairwise plot derived from the second pair of PLS axes. D, Shape covariation between both blocks of variables accounted for by the second pair of PLS-axes. E, Pairwise plot derived from the third pair of PLS axes. Box-plots indicate the range variation of the species scores within this third of PLS axes in both blocks of variables. The vertical line inside each box is the median. Box length is the interquartile range (IQR) and shows the difference between the 75th and 25th percentiles. Horizontal bars enclose values of 5–95%. F, Shape covariation between both blocks of variables accounted for by the third pair of PLS-axes. Symbols as in Figure 4.

228x327mm (300 x 300 DPI)

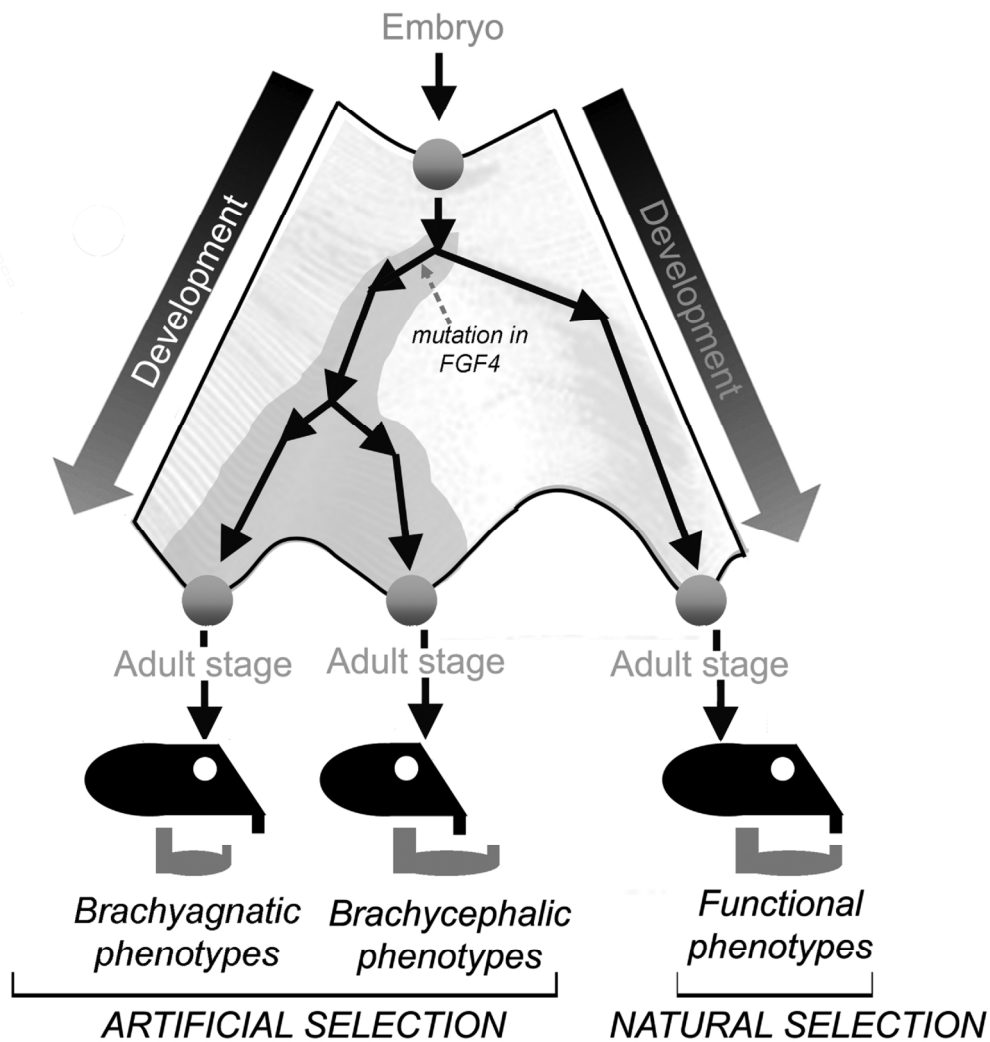


Figure 8. Hypothesis of ontogeny supported for the high integrated pattern of lower jaw and cranial shape depicted in the context of the epigenetic landscape of Waddington (1956). We propose here the mutation in the retro-gene coding for fibroblast growth factor 4 (FGFR4) as the factor that induces a change in the developmental pathway from functional to teratological phenotypes (see text for details) fixed by man's selection.

167x175mm (300 x 300 DPI)

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60