

1 Pharmacogenomics in Drug Induced Liver Injury

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14 **Abstract:** Drug-induced liver injury (DILI) is a severe adverse effect. The majority of
15 DILI cases are idiosyncratic and several mechanisms have been postulated to explain
16 why some subjects develop DILI with drugs that are safe for the majority of individuals.
17 Major mechanisms proposed for DILI are based on the production of reactive
18 metabolites, immune-mediated hepatotoxicity, a “danger signal” hypothesis and/or
19 alterations in mitochondrial function. These mechanisms are compatible with the
20 hypothesis for genetic variability in drug metabolism or bioactivation and are a major
21 determinant for DILI. In this review we summarize present knowledge on underlying
22 mechanisms, and clinical expression as well as genetic and non-genetic factors that
23 modulate the risk of developing DILI. With regard to DILI pharmacogenomics, we
24 summarize current evidence on the role of polymorphisms in genes coding for the drug-
25 metabolizing enzymes CYP1A2, CYP2C9, CYP2C19, CYP2D6, CYP2E1, CYP3A4,
26 CYP3A5, NAT2, GSTM1, GSTT1, UGT1A1, UGT1A3, UGT1A9
27 and UGT2B7. Conclusive evidence for association with DILI risk has been obtained for
28 non-mutated *CYP2E1*, slow *NAT2* and slow *GSTM1* genotypes. For the rest of the genes
29 additional pharmacogenomics and toxicogenomics studies are required. We identify
30 potential sources of heterogeneity in studies carried out so far as well as new genetic
31 targets which require further investigation.

32 **Keywords:** Pharmacogenomics, toxicogenomics, hepatotoxicity, intolerance, liver injury.
33

34 INTRODUCTION

35 Drug-induced liver injury (DILI) is a major concern for the pharmaceutical industry and
36 regulatory agencies as it remains the single leading cause of stopping drugs for further
37 development and drug withdrawal once on the market [1-3]. Nearly one thousand
38 medicines used in clinical practice have been shown to induce hepatotoxicity in medical
39 literature [4]. Two large prospective Registries in Spain and the USA intended to capture
40 DILI incidences regardless of causative agent have shown that the main compounds

41 incriminated in DILI were anti-bacterial agents, non-steroidal anti-inflammatory drugs
42 (NSAIDs) and anticonvulsants [5, 6]. In both Registries, amoxicillin-clavulanate was the
43 leading agent responsible for DILI.

44 The incidence of DILI remains largely undetermined in the general population. In a
45 population-based study in France, the frequency of DILI was prospectively estimated to be
46 about 14 per 100,000 inhabitants per year [7]. Further, in roughly 1% of medical inpatients
47 DILI develops during the course of hospitalization [8].

48 An obvious ‘dose effect’ is the hall-mark of intrinsic “dose-dependent” adverse drug
49 reactions. Acetaminophen (paracetamol) is, in practical terms, the only marketed drug
50 whose capacity to produce liver injury is closely related to the dose consumed.

51 In contrast, *idiosyncratic* DILI, which represents the bulk of cases, is typically non-
52 predictable. Idiosyncratic reactions cannot usually be predicted from preclinical studies.
53 They are further classified as immunoallergic (hypersensitivity features present), or
54 metabolic, although this is a more arbitrary division and overlap between categories can
55 occur. Although the classical view of idiosyncrasy in DILI refers to a lack of a clear dose
56 relationship, it has been recalled that even in the most extreme form of unpredictable
57 adverse drug reactions (allergic), a dose threshold is required to trigger the immune
58 response, [9]. Actually, a recent study used two comprehensive pharmaceutical databases
59 to examine the relationship between daily doses of the top 200 brands and the top 200
60 generic medications by prescription volume in the United States during the year 2005 and
61 reported frequency of their hepatic adverse events. By categorizing medications into <10
62 mg/day, 11-49 mg/day, and >50 mg/day groups, a statistically significant relationship was
63 observed between daily dose of oral medicines and reports of liver failure, liver
64 transplantation and death caused by DILI. In a second part of the study the authors examined
65 serious DILI cases reported to the Swedish Adverse Drug Reactions Advisory Committee
66 (1970-2004) and found that most of the 598 cases of idiosyncratic DILI (77%) occurred in
67 patients taking drugs at a daily dose of at least 50 mg [10]. Additionally, it has been pointed
68 out that the majority of compounds that either have been withdrawn from the market, or
69 have received a black box warning due to hepatotoxicity were prescribed at daily doses
70 greater than 50 mg/day. These data suggest that there is some dose-response relationship in
71 idiosyncratic DILI as well. Therefore, the classic distinction between predictable and
72 unpredictable adverse hepatic reactions to drugs is worthy of revision as the concept is
73 overly simplified [11].

74 **MECHANISMS**

75 Development of DILI is a complex, multi-step process that requires the interplay

76 between the toxic potential of the drug, and genetic and acquired factors coupled with
77 failure in the adaptive processes. Thus, most patients tolerate the drug without adverse
78 liver effects or there is a background of mild, and often transient, asymptomatic liver injury
79 (i.e. statins, isoniazid), indicating an adaptation to the drug and further tolerance.
80 Therefore, the combination of susceptibility factors coupled with drugs that (due to
81 variation in handling between different phases of drug metabolism, detoxification, and
82 transport) reach a threshold for exposure to drug or toxic metabolites, enhances the risk of
83 idiosyncratic hepatotoxicity. There are nowadays several independent (yet not mutually
84 excluding) hypotheses to explain idiosyncratic DILI. No one may satisfactory accomplish
85 all complex circumstances in which DILI occurs.

86 The reactive metabolite hypothesis states that most DILI instances would result from
87 the production of reactive metabolites [12-16], with capacity to initiate the damage once a
88 critical threshold has been crossed. In support of this hypothesis is the common
89 occurrence of centrilobular necrosis (the zone richest in CYPs) in severe acute DILI. This
90 concept in fact arose from the discovery that acetaminophen (APAP) induced
91 hepatotoxicity is caused by a reactive imidoquinone metabolite [17]. Most of this reactive
92 metabolite is detoxified by reaction with glutathione, and significant toxicity does not occur
93 until liver glutathione is substantially depleted. It is noteworthy that the bulk of our
94 knowledge of DILI derives from the experimental model of APAP hepatotoxicity [18]. But
95 whether clinical APAP liver injury is relevant to idiosyncratic hepatotoxicity has not been
96 fully established. One study has given support to the reactive metabolite hypothesis,
97 showing that among the 27 drugs frequently cited in adverse drug reaction studies, a
98 significantly higher proportion (59%) were metabolized by enzymes coded for by allelic
99 variants (“genotype”) associated with a poor metabolizer “phenotype” as compared with
100 22% of randomly selected drugs and 7% of the top-selling drugs [18] However, in general,
101 substantial increases in enzyme activity reflect environmental rather than genetic factors
102 [19].

103 Immune-mediated hepatotoxicity appears to involve the generation of reactive
104 metabolites that undergo covalent binding with hepatocytes/carrier proteins, also known
105 as “haptization” [20]. The covalent binding of the reactive metabolites to “self” proteins
106 results in the formation of neo-antigens that “mislead” the immune system into mounting
107 an immune response against hepatocytes. However the pattern of the immune response-to-
108 adduct is likely to vary among individuals [21, 22]. It is not clear as yet whether the
109 association of immune response with liver injury and reactive metabolite formation is
110 coincidental or consequential. Classic examples that seem to indicate that reactive
111 metabolite formation can be a first step to the development of an immune response are

112 some kind of drug-induced immunoallergic hepatitis. Several drugs (some of them no
113 longer on the market), including halothane, tienilic acid, dihydralazine and
114 anticonvulsants, induce immunoallergic hepatitis associated with antibodies antihapten
115 (neoantigen) or anti-native proteins such as CYPs. In these cases it is believed that the
116 drug is first metabolized into a reactive metabolite which binds to the enzyme that
117 generated it, resulting in the production of a neoantigen which, once presented to the
118 immune system, might trigger an immune response characterized by the production of
119 antibodies recognizing both the native and/or the modified protein. For example, most
120 patients who develop halothane-induced hepatotoxicity have antibodies against
121 trifluoroacetyl modified proteins
122 [23] and this implies that modification of protein by the reactive metabolite has led to an
123 immune response. Halothane is administered for a brief period of time, which is insufficient
124 in a single exposure to allow for the development of a full adaptive immune response, and
125 the observation that hepatotoxicity almost always occurs after multiple exposures [24] also
126 suggests a process of immune sensitization. Another example is the antihypertensive drug
127 dihydralazine that has caused, on rare occasions, immunoallergic hepatitis characterized by
128 the presence of anti-liver microsome (anti-LM) autoantibodies. Since these antibodies
129 found in the serum of the patients react with CYP1A2, it is suggested that dihydralazine is
130 biotransformed into a reactive metabolite, which covalently binds to cytochrome CYP1A2
131 and triggers an immunological response as a neoantigen [25].

132 Recent data give further support to the participation of acquired immune system (in a
133 background of genetic predisposition) in the hepatotoxicity induced by some drugs; a
134 genome-wide association study using 866,399 markers in 51 cases of flucloxacillin DILI
135 and 282 controls matched for sex and ancestry showed an association peak in the major
136 histocompatibility complex (MHC) region with the strongest association ($P = 8.7 \times 10^{-33}$)
137 seen for rs2395029[G], and other SNPs [26] all of which are a part of an extended
138 MHC 57.1 haplotype present in < 4% of Europeans, which is associated to hypersensitivity
139 to abacavir [27]. Also the DILIGEN group has confirmed [31] the significant association
140 previously reported between *DRB1*1501-DRB5*0101-DQB1*0602* haplotype and cho-
141 lestatic hepatitis related to amoxicillin-clavulanate [26].

142 A compelling hypothesis which attempts to unify several observations in DILI is the
143 danger signal hypothesis [28]. The rational basis for this hypothesis is the observation that
144 foreign proteins hardly induce an immune response in the absence of an adjuvant, and
145 the primary purpose of the adjuvant is to activate antigen-presenting cells (APCs) [29].
146 Adapting the danger signal hypothesis to DILI would mean that if a drug is metabolized
147 in the liver to form a reactive compound that covalently binds to proteins (haptenization),

148 this alone might be insufficient to trigger an immune reaction or could induce a non-
149 pathogenic immune response [20]. The danger that primes a genetically susceptible
150 adaptive immune system might include the background mild hepatic injury caused by the
151 reactive metabolites [20]. Thus, the ability of a reactive metabolite to cause cell damage
152 could be a determinant of whether a drug that forms reactive metabolites will be associated
153 with a significant incidence of idiosyncratic DILI [9]. It appears that endogenous
154 molecules that act as danger signals may bind to the same receptors that recognize
155 molecules on foreign pathogens, that is, toll-like receptors [30]. Other attractive co-
156 stimulatory signals could be infection or inflammatory conditions, such as HIV, or other
157 viral or bacterial infections. There are specific examples where this appears to be true;
158 however, there does not appear to be a clear and dominant pattern of factors such as
159 infection being associated with an increased risk of idiosyncratic DILI [29]. Subsequently,
160 hapten formation leading to major histocompatibility complex class II (MHC II)
161 presentation of haptened peptides by (APC) along with co-stimulation of APC by
162 “danger” signals promote helper T-cell activation (clonal antigen-recognizing cytotoxic
163 cell expansion) and B-cell mediated antibody production. In response to cellular
164 stress/death signal, innate immune system leads to the production of protective and/or
165 injurious cytokines [20]. The altered cytokine milieu of chronic viral disease can also
166 influence susceptibility to non-allergic toxicity and helps to explain the suggested
167 increased susceptibility of patients with HIV or chronic hepatitis B and C to isoniazid
168 hepatotoxicity [31]. Modulating innate immune response might be crucial in determining
169 the severity and extent of liver injury. Factors affecting the expression of protective
170 cytokines (genetic polymorphisms) or an underlying disease might promote liver toxicity.
171 The balance between protective cytokines (interleukin (IL)-10, IL-6, monocyte
172 chemoattractant protein (MCP)-1, MCP-2) and pro-inflammatory cytokines (interferon
173 (IFN)- γ , fas ligand (FasL), tumor-necrosis factor (TNF) in the liver may determine the
174 extent of organ damage and the mode of cell death (apoptosis and necrosis) [20].

175 Drug impairment of the mitochondrial function leading to decreased fat oxidation and/or
176 energy production which results in steatosis and cell death is the hallmark of several
177 instances of DILI [32, 33]. Opening the mitochondrial permeability transition pore, which
178 subsequently results in necrosis or apoptosis, either through drug-induced direct toxicity or
179 immune reactions, is another pathway of cell damage [34]. Several drugs had been found
180 to carry *in vivo* or *in vitro* mitochondrial hazards and recently a unifying hypothesis has been
181 proposed to explain the susceptibility for a number of drugs that target mitochondria and
182 cause DILI [35]. The mitochondrial hypothesis states that in the setting of underlying genetic
183 or acquired mitochondrial abnormalities, initially silent and accumulating organelles damage

184 then reach a critical threshold and abruptly trigger overt liver injury. This is consistent with
185 the fact, that the time to onset is typically delayed (by weeks or months) in idiosyncratic
186 DILI. The heterozygous superoxide dismutase 2 gene knockout (Sod2^{+/-}) mice provided
187 supporting evidence for this concept showing that the intraperitoneal administration of 30
188 mg/kg/day of troglitazone in this animal model for twenty-eight days caused liver injury,
189 characterized by increased serum ALT activity and hepatic necrosis [36]. However, these
190 results were irreproducible in a repeated dose toxicity study using the same animal model
191 treated orally with 300 mg/kg/day troglitazone for twenty- eight days, suggesting that the
192 mitochondrial damage alone might not be the main cause of the troglitazone-induced
193 idiosyncratic liver injury observed in humans [37]. Nevertheless, extensive genetic analyses
194 of samples from DILI patients would be needed to provide proof-of-concept [34].

195 **RISK FACTORS**

196 Risk factors influencing DILI are in most instances those affecting drug disposition and
197 metabolism. Incomplete information in case reports have precluded analyses of drug-
198 related, environmental risk factors and clinical presentation profiles, which however can
199 be obtained in large cohorts of patients with DILI [6, 38, 39]. Known risk factors are,
200 nevertheless, poorly predictive of hepatotoxicity in clinical practice. Furthermore, in many
201 instances of DILI no risk factor can be identified.

202 **CHEMICAL HAZARDS**

203 Chemical properties of some medicaments make them more prone to induce liver
204 damage but this is probably critical only for selected medications. For example, ebrotidine
205 (an H₂ receptor antagonist withdrawn from the market in Spain because of severe
206 hepatotoxicity) [40] shares with famotidine the same thiazole ring but in the side chain it
207 bears a 4-bromo-benzene ring which could explain the differences in liver safety between
208 both drugs [40]. Similarly, the quinolone trovafloxacin (withdrawn from the market in
209 Europe and with black box warning in the USA because of its hepatotoxic potential) bears
210 a unique difluorinated side chain that is not found in the other quinolones, with the
211 exception of temafloxacin (another drug withdrawn because of unacceptable toxicity) and
212 which renders these drugs highly lipophilic [41]. Clotiazepam and bentazepam are more
213 structurally similar to phenothiazines than to other benzodiazepines due to the thiophene
214 ring they bear [42] and in fact, unlike other benzodiazepines, several cases of moderate-to-
215 severe hepatitis, and even chronic liver injury, have been reported with both drugs [43].

216 On the other hand, certain compounds form acyl glucuronides, which are capable of
217 hydrolysis, intra-molecular acyl-migration and covalent binding to proteins [44]. It is
218 worth noting that 10 out of

219 47 drugs (alclofenac, bendazac, benoxaprofen, fenclofenac, ibufenac, indoprofen,
220 pirprofen, suprofen, ticrynafen and zomepirac) were removed from the US, UK and
221 Spanish markets between 1964 and 1993 due to their poor safety profile. They are
222 carboxylic acids and are metabolized primarily to acyl glucuronides in hu- mans. Several
223 environmental factors exist which appear to operate in determining individual
224 susceptibility.

225 **Age**

226 Elderly patients are the population most vulnerable to DILI. In particular, increased age
227 is a susceptibility factor for developing hepatotoxicity to antituberculous drugs [45]. Old
228 age has also been implicated in amoxicillin-clavulanate induced cholestasis [46, 47]. In
229 contrast, valproic acid and erythromycin hepatotoxicity are more common in childhood
230 [44]. Old age, rather than being a predisposing factor to DILI, seems to have an impact on
231 the phenotypic expression of toxic liver damage, strongly favouring the appearance of a
232 cholestatic pattern of injury [5, 38].

233 **Gender**

234 Women are believed to be more susceptible than men to most forms of DILI [7, 48-50]
235 For instance, autoimmune hepatitis triggered by drugs is seen almost exclusively in
236 women [24] and diclofenac hepatotoxicity has been reported more frequently in women
237 with osteoarthritis [44].

238 However, new epidemiological data have challenged this traditional belief, showing a
239 similar sex distribution in DILI cases [5, 51], with a higher prevalence of female gender
240 only at younger ages [5]. Yet female gender is considered to be a risk factor for develop-
241 ing fulminant liver failure [38] and the female sex accounted for 76% of patients
242 presenting with drug-induced acute liver failure in the USA who were transplanted [52].
243 As with age, gender may influence the clinical presentation of DILI as women are
244 overrepresented in the hepatocellular type of injury [5].

245 **Chronic Alcohol Consumption**

246 A history of alcohol intake was considered to be a general risk factor for idiosyncratic
247 DILI, and as such scores 1 + point in the standard clinical scale for the assignation of
248 causality developed by experts, the CIOMS (Council for International Organizations of
249 Medical Sciences) or RUCAM (Roussel Uclaf Causality Assessment Method) scale [53].
250 However, for most of the drugs capable of inducing hepatotoxicity, there is no evidence
251 for a role of alcohol in potentiating toxicity. Furthermore, in a recent report analyzing a
252 prospective cohort in the US any alcohol use in the preceding 12 months was a negative

253 predictor of severe DILI (odds ratio, 0.33; 95% confidence interval, 0.15– 0.76) [6]. In
254 practical terms, chronic alcohol intake increases the risk of developing liver fibrosis during
255 methotrexate therapy, and enhances acetaminophen hepatotoxicity by inducing CYP2E1,
256 (with generation of higher levels of the reactive metabolite and depletion of glutathione
257 stores), as well as susceptibility to liver damage from isoniazid, halothane and cocaine
258 [44].

259 **Concomitant Drugs**

260 Are capable of modulating the hepatotoxic potential of other drugs through CYP or
261 hepatic transport systems induction, inhibition or substrate competition. In addition,
262 concomitant medication with hepatotoxic potential may further increase the risk [54]. Con-
263 current use of anticonvulsants which induce drug metabolism greatly increases the risk of
264 hepatotoxicity due to valproate [44]. The concurrent use of isoniazid and rifampicin, a
265 potent microsomal inducer, enhances the risk of isoniazid hepatotoxicity [55].

266 **Underlying Disease States**

267 Controversy exists on whether preexisting liver disease is a susceptible factor for the
268 development of hepatotoxicity. The presence of chronic hepatitis B, hepatitis C infection
269 or co-infection with human immunodeficiency virus (HIV) increases the risk of isoniazid
270 hepatotoxicity or elevated transaminases on HAART (highly active antiretroviral therapy)
271 [56]. The increased susceptibility exhibited by HIV patients suggests a role for cytokine
272 imbalance [31]. Rifampicin is more hepatotoxic when used for control of itching in
273 patients with primary biliary cirrhosis [57].

274 Obesity, diabetes mellitus type 2 and insulin resistance are known risk factors for
275 steatohepatitis, and have also been shown, along with psoriasis, to increase the risk of
276 developing liver fibrosis during methotrexate therapy [58]. In the analysis of the first 300
277 cases of idiosyncratic hepatotoxicity included in the prospective DILIN study in the US
278 the presence of diabetes mellitus was an independent risk factor for severe DILI (odds
279 ratio, 2.69, 95% confidence interval, 1.14 – 6.45) [6].

280 **CLINICAL EXPRESSION**

281 The phenotypic expressions of DILI may present in several ways (clinical and
282 pathological) that resemble known forms of acute and chronic liver disease; the severity
283 ranges from sub-clinical elevations in liver enzyme concentrations to acute liver failure.
284 On the whole, drugs tend to induce acute hepatitis, cholestasis or a mixed condition [24]
285 (Table 1).

286 Liver histology (although not very specific and, at best, “compatible with”) is still the

287 ideal tool for defining the pattern of liver damage. However, since a liver biopsy specimen
288 is often not available, the pattern of drug-related liver injury is, from a practical
289 standpoint, classified according to laboratory data [59]. The classification scheme was
290 proposed by the Council for International Organizations of Medical Sciences (CIOMS) and
291 recently updated by the Food and Drug Administration Drug Hepatotoxicity Committee
292 (Table 2). Reference is made to the pattern of serum enzymes elevations to define whether
293 the hepatic injury is “hepatocellular”, “mixed” or “cholestatic”, which are defined by
294 calculation of the “R value”. The R value is calculated by dividing the alanine
295 aminotransferase (ALT) by the alkaline phosphatase (AP) using multiples of the upper limit
296 of the normal range for both values. The values used should be from the same day and
297 should be those from the initial blood test results. The reason is that an initial hepa-
298 tocellular pattern can be converted to a cholestatic-type pattern during the follow-up, which
299 may be related to the differences in the faster rate of improvement of ALT versus AP or
300 glutamyltranspeptidase following drug withdrawal [60]. Calculation of the type of liver
301 damage according to this rule correlates quite well with the underlying histological lesion,
302 has proven prognostic value, and is essential to apply the RUCAM scale.

303 The acute hepatocellular (cytotoxic, cytolytic) type of liver injury is the most common
304 expression of hepatotoxicity [6, 38] and is observed with many drugs (Table 1). Patients
305 with acute hepatocellular injury related to drugs are at risk of acute liver failure. The
306 presence of combined increases in ALT and bilirubin levels in DILI reflects a substantial
307 loss in hepatocellular function and potential for liver failure [61]. These parameters are
308 considered a specific indicator of severe hepatotoxic potential of a drug. In patients with
309 acute drug-induced hepatitis the presence of jaundice is the most significant predictor of
310 mortality/liver transplantation. The observation by Hyman Zimmerman, known as “Hy’s
311 rule” [24], predicts a mean mortality (or its surrogate marker, liver transplantation) of
312 10% (range 5-50%) for jaundiced patients with acute toxic hepatocellular damage. Hy’s
313 rule has been validated in large case series from Spain [38], Sweden [39] and the US [6],
314 in which analysis of pooled data showed that other variables such as old age, female gender
315 and AST levels were independently associated with a poor outcome [38, 39]. The late
316 Hyman Zimmerman also stated that prolonging exposure to the causative drug once
317 hepatotoxicity is initiated may favor a fulminant outcome [24]. Interestingly, in the
318 American Registry the median duration between first exposure to the implicated agent
319 and DILI recognition was significantly longer in severe cases than in mild/moderate cases
320 although the association was not statistically significantly longer in the multivariate
321 analysis [6]. Further analysis of the Swedish database has shown that there was also a
322 significant linear relationship between daily dose and the outcome of death/liver

323 transplantation (2%, 9.4%, and 13.2% in <10, 11-49, and >50 mg/day groups, respectively,
324 $P= 0.03$)[10].

325 Conversely, it has recently been suggested that eosinophilia accompanying DILI may be
326 associated with a better short-term prognosis [62]. Actually, none of the patients included
327 in the Spanish Registry with idiosyncratic DILI who died, evolved to liver failure, or
328 required a liver transplantation, had eosinophilia, whereas this feature was found in the
329 23% of the patients with milder outcomes [38]. Of note is that the height of transaminases
330 in acute hepatocellular DILI lacks prognosis significance. Indeed, a decrease in liver
331 enzymes after drug withdrawal in patients with severe DILI may reflect, rather than a
332 clinical improvement, a limited hepatic reserve associated with impending liver failure
333 [61]. Moreover, if acute injury is superimposed on underlying liver dis- ease, monitoring
334 liver function tests may be difficult to interpret. For example, if advanced cirrhosis is
335 present, the severity of liver injury may be underestimated by the height of the serum ALT
336 measurements [63]. which is predominantly centrilobular [24, 31]. In such cases director
337 more commonly immune-mediated damage to the cholangyocyte is probably caused by
338 toxic intermediates excreted into bile.

339 In mixed hepatic injury, the clinical and biological picture is intermediate between the
340 hepatocellular and the cholestatic pat- terns, and features of either type may predominate.
341 By definition, the ALT/AP ratio is between 2 and 5 (Table 2). When faced clini- cally with
342 a mixed hepatitis picture, the gastroenterologist should always suspect and seek a
343 potentially hepatotoxic medication, since this type of injury is far more characteristic of
344 drug-induced hepatotoxicity than of viral hepatitis [24]. Almost all drugs that produce
345 cholestatic injury are also capable of inducing a mixed pattern. Genotypic studies in patients
346 with DILI have shown the association of some HLA alleles with cholestatic/mixed
347 damage, which points to an allergic basis for this variety of hepatotoxicity [64].

348

349 Acute cholestatic injury is caused by many drugs, including cyclosporine anabolic and
350 contraceptive steroids, which characteristically produce DILI with minimal or absent
351 accompanying in- flammation, presumably by interfering with bilirubin and bile acids
352 export from the canaliculi. With other medications, which include amoxicillin-clavulanate,
353 macrolide antibiotics and phenothiazine neuroleptics, there are typically variable degrees
354 of portal inflammation and hepatocyte necrosis, in addition to marked cholestasis Drug-
355 induced acute cholestatic and mixed lesions are less severe in the short-term outcome than
356 the hepatocellular type. An exception to this rule is the analysis of the American Registry
357 in which, within 6 months of DILI diagnosis, 14.3% of patients with cholestatic injury died

358 (as compared with only 7.5% of patients with hepatocellular damage). However, most of
359 the fatalities in subjects with cholestatic damage were not related to liver failure in this
360 study [6].

361 The resolution of cholestatic and mixed lesions, nevertheless, are generally slower, with
362 a higher likelihood towards chronicity in one study (9% vs 4% respectively; $p < 0.031$) [60].
363 In contrast, in the hepatocellular pattern with jaundice at presentation, the severity of
364 chronic lesions was greater (higher incidence of cirrhosis and chronic hepatitis) [60, 65].
365 Cardiovascular and central nervous system drugs are the main groups leading to chronic
366 liver damage [60]. Of the whole population included in the Spanish Registry, 5.6% of
367 patients had biochemical evidence of chronicity [60], a figure similar to that reported in
368 the Swedish population (6%), even though the majority of the patients included in this
369 latter study were identified on an outpatient basis and had mild to moderate DILI [66]. A
370 higher prevalence of chronicity (14%) has recently been reported in the US [6]. Further
371 studies on the natural history of DILI are on-going.

372 **PHARMACOGENOMICS IN DILI**

373 As commented above, drug biotransformation is believed to be a relevant process in
374 DILI. The biotransformation of drugs and xenobiotics involves several steps, including
375 Phase I oxidation by cytochrome P450 enzymes followed by Phase II conjugations through
376 enzymes as N-acetyltransferase or glutathione transferase. The activity of drug-
377 metabolizing enzymes shows significant inter-individual and interethnic variation. Genetic
378 and environmental factors are responsible for this variability. Most drug-metabolising
379 enzymes are polymorphic, due to the presence of polymorphisms, that is, mutations which
380 occur with a frequency higher than 1%, in genes coding for such enzymes [67, 68]. Variant
381 genes cause abolished, reduced, altered or increased enzyme activity, because of complete
382 gene deletions, single nucleotide polymorphisms that occur isolated or combined, and gene
383 duplications. Individuals carrying enzyme-inactivating polymorphisms display higher drug
384 plasma concentrations and lower clearance rates when treated at standard doses [67, 69].
385 Therefore, an increased prevalence and severity of adverse drug reactions, including DILI,
386 could be expected among subjects carrying enzyme-inactivating mutations, when receiving
387 drugs that are substrates of the defect enzyme. Polymorphisms which have been associated
388 with an increased risk of drug hepatotoxicity are summarized in Table 3.

389 It should be borne in mind that the biotransformation of a drug most commonly
390 involves more than one metabolic pathway and that several enzymes may be involved in
391 the metabolism of a single drug. As our understanding of drug metabolism increases, more
392 and more evidence suggests that minor metabolic pathways may play a relevant role in

393 adverse drug reactions [70]. Therefore the analysis of the relationship between the
394 occurrence of hepatotoxicity or adverse drug reactions and the metabolism of a drug
395 should consider all the pathways involved in the metabolism of the drug.

396 **Cytochrome P450**

397 The most important enzymes of phase I metabolism are the cytochrome P450 system,
398 a superfamily of isozymes that catalyze the oxidation of drugs and other xenobiotics.
399 Multiple forms of CYP enzymes play important roles in the oxidation of structurally
400 diverse drugs (<http://medicine.iupui.edu/flockhart>; <http://www.imm.ki.se/CYPalleles/>).
401 CYP1, CYP2 and CYP3 families are major forms. Among these, important human drug-
402 metabolizing enzymes are CYP1A2 (caffeine, estradiol, lidocaine, tacrine, theophylline,
403 verapamil, R-warfarin), CYP2C8 (amodiaquine, R-ibuprofen, paclitaxel, pioglitazone,
404 troglitazone) CYP2C9 (diclofenac, ibuprofen, phenytoin, piroxicam,
405 tetrahydrocannabinol, tolbutamide, warfarin), CYP2C19 (diazepam, hexobarbital, S-
406 mephenytoin, omeprazole, pentamidine, propranolol), CYP2D6 (codeine, debrisoquine,
407 dextromethorphan, encainide, haloperidol, metoprolol, mexiletine, paroxetine,
408 phenothiazines, propranolol, risperidone, sertraline, tricyclic antidepressants,
409 venlafaxine), CYP2E1 (chlorzoxazone, halothane, paracetamol, isoflurane, desflurane,
410 enflurane), and finally CYP3A subfamily members CYP3A4 and CYP3A5 mediate the
411 metabolism of many very useful drugs [70-87].

412 With regard to CYP1A2, the enzyme shows interindividual differences due to genetic
413 polymorphisms and to modulation with many compounds such as drugs (fluvoxamine or
414 ciprofloxacin), foods (grapefruit juice or broccoli) and toxins (polycyclic aromatic
415 hydrocarbons present in tobacco smoke) [88]. Several *CYP1A2* allelic variants have been
416 reported, and some of these are related to decreased enzyme activity or decreased
417 expression of the enzyme [89-91]. For an updated list of *CYP1A2* variant alleles see the
418 website <http://www.cypalleles.ki.se/cyp1a2.htm>. CYP1A2 have been associated with 5-
419 n-butyl-7-(3,4,5-trimethoxybenzoylamino) pyrazolo[1,5-a]pyrimidine (OT-7100)
420 hepatotoxicity [92]. The hepatotoxicity of OT-7100, a pyrazolopyrimidine derivative
421 with potential analgesic effects, has been studied *in vitro*, in human liver microsomes, and
422 could be related to the formation of a reactive metabolite produced by CYP1A2 activity.
423 No studies have confirmed the influence of *CYP1A2* polymorphisms in DILI.

424 The *CYP2C* gene cluster is highly conserved. Many of the *CYP2C* genes are within the
425 same haplotype block [93, 94]. Comparison of linkage disequilibrium between pairs of
426 loci carried out in the frame of the HapMap project [95], indicates that the four *CYP2C*
427 genes, namely *CYP2C8*, *CYP2C9*, *CYP2C18* and *CYP2C19*, are allocated in two clusters,

428 the first one including *CYP2C18*, *CYP2C19* and *CYP2C9*, and the second one including
429 *CYP2C8* and part of *CYP2C9* flanking region [94]. Another proposed distribution
430 includes *CYP2C18* and *CYP2C19* in one cluster and *CYP2C9* and *CYP2C8* in the other
431 [93]. Several variant alleles have been described for both *CYP2C8* and *CYP2C9* genes. The
432 variant alleles *CYP2C8*3*, *CYP2C9*2* and *CYP2C9*3*, are the commonest variant alleles
433 among Caucasian individuals [70, 96]. Besides the putative implications of CYP2C
434 enzymes in cancer risk and other diseases [70, 83, 97-100], it has been unambiguously
435 shown that genetic variations in CYP2C enzymes cause variations in drug metabolism. In
436 addition, changes in the expression of CYP2C enzymes may also lead to altered response,
437 as is the case for resistance to paclitaxel [82, 86]. Regarding single nucleotide
438 polymorphisms, the *CYP2C8*3* variant allele causes impaired bio- disposition of ibuprofen
439 *in vivo* [80]. Other variant *CYP2C8* alleles are *CYP2C8*2*, which is related to increased
440 Km for paclitaxel metabolism [101], *CYP2C8*4*, which causes a marginal decrease in
441 metabolic capacity, and *CYP2C8*5*, which causes a frameshift and an early stop codon
442 [102]. The effect of *CYP2C8* variant alleles on drug metabolism, pharmacokinetics and
443 adverse effects remains to be investigated in detail. Because many drugs causing DILI are
444 metabolized in part by CYP2C8, it is of particular interest to elucidate the role of *CYP2C8*
445 variant alleles in the risk to develop DILI in patients receiving CYP2C8 substrates, and we
446 are currently conducting such studies. With regard to CYP2C9, the enzyme encoded by the
447 *CYP2C9*3* allele shows a reduced efficiency of paclitaxel [103] or arachidonic acid
448 metabolism whereas it keeps full activity on amiodarone [102, 104]. Several examples of
449 impaired drug metabolism *in vivo* related to variant alleles have been provided, either for
450 *CYP2C9*2* or for *CYP2C9*3*, with drugs such as acenocoumarol, warfarin, several
451 NSAIDs, hypoglycemics and angiotensin antagonists. Diverse studies have suggested a
452 role for *CYP2C9* polymorphisms in liver toxicity caused by voriconazole, with a trend
453 towards higher toxicity for carriers on non-mutated alleles [105], for valproic acid, with
454 higher risk for carriers of mutated alleles [106], or for leflunomide with higher risk being
455 associated to mutated alleles [107]. Unfortunately no studies involving a large number of
456 patients with DILI caused by these drugs have been conducted. The metabolism of the most
457 NSAIDs is carried out by CYP2C9 and some of them are known to be hepatotoxic. Based
458 on this hypothesis various studies were conducted to test whether *CYP2C9* polymorphisms
459 can be considered as risk factors for DILI [108, 109]. Aithal *et al.* investigated the
460 influence of *CYP2C9*2* and *CYP2C9*3* in hepatotoxicity caused by diclofenac in 24 pa-
461 tients, with negative findings [108]. The study by Pachkoria had a similar sample size (28
462 patients) and did not identify a positive association [109]. Although both studies point to a
463 lack of a major association between *CYP2C9* polymorphisms and DILI, further studies are

464 required to rule out such an association for several rea- sons. First, the statistical power of
465 both studies is too low to reach definitive conclusions, and second, because studies should
466 involve patients with DILI caused by drugs that are metabolized mainly by the CYP2C9
467 enzyme. For instance, the study on diclofenac-induced DILI [108] should have considered
468 other drug-metabolizing enzymes. The primary metabolism of diclofenac is carried out by
469 acyl glucuronidation, by uridine 5'-diphosphoglucuronosyl transferase UGT2B7 and by
470 diverse hydroxylation pathways to produce hydroxymetabolites that are usually further
471 conjugated [110]. CYP2C9 plays a minor role in diclofenac metabolism and therefore a
472 lack of association between *CYP2C9* polymorphisms and diclofenac-induced DILI is not
473 surprising. Daly *et al.* analyzed the polymorphisms of several enzymes involved in the
474 metabolism of diclofenac with relation to the risk of developing DILI and they found a
475 positive association for *UGT2B7* and *CYP2C8* variant alleles [111]. Another factor that
476 should be considered in further studies is that a strong linkage between the *CYP2C8**3 and
477 the *CYP2C9**2 variant alleles has been described to occur in Caucasians [80, 81, 83, 112-
478 114]. This fact, together with increasing evidence that support a role for the CYP2C8
479 enzyme in the metabolism of drugs thought to be substrates of CYP2C9 [70, 73, 80, 81,
480 113], indicates that, in order to be conclusive, further studies should analyze as putative
481 risk factors genetic variations in both genes, *CYP2C8* and *CYP2C9*, simultaneously.

482 With regard to CYP2C19, the variant alleles *CYP2C19**2 and *CYP2C19**3 are the major
483 variant alleles although *CYP2C19**3 is only present in Oriental individuals and is not
484 detected in Caucasians [109, 115]. CYP2C19 deficiency activity has been implicated in
485 alterations in the metabolism, and in toxicity events with the use of several drugs [116],
486 including hepatotoxicity caused by Atrium (a fixed combination preparation containing
487 phenobarbital, febarbamate and difebarbamate) [117], but no clear association of *CYP2C19*
488 variant alleles with DILI has been identified so far. Recently a study on patients with DILI
489 caused by several drugs has shown that the variant *CYP2C19**2 allele does not enhance the
490 risk of DILI [109], although the study requires confirmation because of the low sample
491 size.

492 Regarding the enzyme CYP2D6, *CYP2D6**3, *CYP2D6**4, *CYP2D6**5 (gene deletion),
493 *CYP2D6**6 and *CYP2D6* gene duplication and amplification are the main variant alleles
494 [74]. There are important ethnic differences in *CYP2D6* genetic polymorphisms [118]. In
495 addition, it has been shown that interindividual differences in the processes leading to gene
496 deletion and duplication can also occur [119]. Several studies have found that poor
497 metabolizers were at higher risk of developing adverse effects [120-123]. CYP2D6
498 deficiency has been associated with perhexiline hepatotoxicity [124], probably due to
499 mitochondrial injury which causes steatosis [125]. A recent study attempted to elucidate

500 whether *CYP2D6* polymorphisms has a role in anti tuberculosis drug-induced hepatitis with
501 negative findings [126]. No other studies investigating the putative role of *CYP2D6*
502 polymorphisms in DILI have been published.

503 CYP2E1 is the major enzyme involved in the metabolism of general anaesthetics [85].
504 Other CYP2E1 substrates are paracetamol, chlorzoxazone, and ethanol [67, 127]. CYP2E1
505 has been related to paracetamol hepatotoxicity in humans [128, 129]. Interestingly, Zaher
506 *et al.* demonstrated in a mouse line that deletion of CYP2E1 and CYP1A2 prevented
507 paracetamol toxicity [130]. Knockout mice lacking CYP2E1 were also protected against
508 carbon tetrachloride-induced hepatotoxicity [131]. At present 13 variant *CYP2E1* alleles
509 have been described. Among these the most common is the variant *CYP2E1**5 allele,
510 initially designated as C2 allele, which is located in the 5' flanking region and causes an
511 increase in the expression of the enzyme [132]. Furthermore, there are additional SNPs
512 where the haplotypes have not yet been determined (see
513 <http://www.cypalleles.ki.se/cyp2e1.htm>).

514 Huang *et al.* analyzed the influence of the *CYP2E1**5 variant allele in a case-control
515 study involving 49 patients with DILI, and identified a positive association with patients
516 lacking the *CYP2E1**5 allele and thus at increased risk of developing antituberculosis
517 drug-induced hepatitis [133]. In agreement with these findings, another study reported
518 association of non-mutated *CYP2E1* genotypes with isoniazid-induced hepatotoxicity
519 [134]. Two studies published recently found no association of *CYP2E1* genotypes and
520 antituberculosis drug-induced hepatotoxicity, although the sample size was too small (18
521 and 23 patients) to achieve definitive conclusions [135, 136]. Overall findings and a recent
522 meta-analysis indicate that non-mutated *CYP2E1* alleles confer a moderate risk to develop
523 anti-tuberculosis drug-induced liver injury [137].

524 Regarding CYP2E1 expression, it has been shown that the inhibition of CYP2E1 with
525 alphanhederin or oleanolic acid is able to reduce carbon tetrachloride-induced
526 hepatotoxicity in mice [138, 139]. CYP2E1 is a highly inducible enzyme. Ethanol-induced
527 hepatotoxicity correlated with CYP2E1 expression and the inhibition of the enzyme
528 protects from liver damage [140]. In addition, ethanol is capable of inducing CYP2E1
529 [141], and it is likely that the production of toxic metabolites could be increased in
530 alcoholics and in carriers of the *CYP2E1**5 variant allele. Isoniazid and other CYP2E1
531 inducers increase the extent of metabolism of sevoflurane, increasing the production of
532 the hepatotoxic inorganic fluoride [142]. Therefore, besides gene polymorphisms,
533 induction and inhibition of CYP2E1 are likely to play a major role in DILI. Unfortu-
534 nately, the actual impact *in vivo* of induction of CYP2E1 for DILI remains to be analyzed.

535 CYP3A4 and CYP3A5 enzymes are involved in the metabolism of about half of
536 clinically used drugs [143], both enzymes making up nearly 30% of the total CYP enzymes
537 expressed in the human liver [144]. High interindividual variation in enzyme activity *in*
538 *vivo* and *in vitro* has been reported. The clinical relevance of this inter- individual
539 variability lies in the wide range of drugs, carcinogens and mutagens that are CYP3A4
540 and CYP3A5 substrates [143]. Although over 40 allelic variants and several SNPs where
541 the haplotypes have not yet been determined have been described (see
542 <http://www.cypalleles.ki.se/cyp3a4.htm>), the genetics basis for variability in CYP3A4
543 metabolism remains elusive. In contrast, the major genetic factor for the interindividual
544 variability in CYP3A5 activity has been elucidated. Only two variant alleles, *CYP3A4*1B*
545 and *CYP3A5*3* are common across diverse ethnic populations and have functional
546 relevance. Present evidence indicates that *CYP3A4*1B* and *CYP3A5*3* are functional
547 polymorphisms *in vivo*. Regarding *CYP3A4*1B*, it seems to modify the ability to metabo-
548 lize some CYP3A substrates, such as quinine [145], although it does not influence the
549 metabolism of other substrates such as midazolam or dextromethorphan [87, 146].
550 *CYP3A5*3* is the commonest *CYP3A5* allele and is associated with severely decreased
551 enzyme activity [79]. For CYP3A4 interindividual variability seems to be related to
552 CYP3A4 induction and inhibition. For instance, grape- fruit juice inhibits the enzyme and
553 may also cause pharmacokinetic changes for CYP3A4 substrates [147]. Now it is admitted
554 that induction and inhibition are more relevant for CYP3A4 enzyme activity than
555 variations in the *CYP3A4* gene [148]. Since CYP3A4 can be induced by dietary
556 components, herbal medicinal products and food supplements [149], an increased
557 production of toxic drug metabolites may be expected in individuals consuming CYP3A4
558 inducers.

559 CYP3A4 has been implicated in the toxicity of acetaminophen, and the underlying
560 mechanism is likely to be related to the orphan pregnane X receptor [150]. It has been
561 shown that troglitazone and flucloxacilin are biotransformed into chemically reactive
562 metabo- lites by CYP3A and other enzymes, but it has not been demon- strated that reactive
563 metabolites are responsible for the troglitazone or flucoxacinil induced hepatotoxicity
564 [151-153]. Regarding poly- morphisms, Haas *et al.* analyzed the role of *CYP3A4* and
565 *CYP3A5* polymorphisms in the risk to develop nepiravine hepatotoxicity in 53 patients
566 and found no association of genotypes with this risk [154]. Levin *et al.* analyzed *CYP3A4*
567 and *CYP3A5* polymorphisms with regard to voriconazole hepatotoxicity with negative
568 findings [105]. Although overall findings are negative, it should be stated that there is very
569 little information on *CYP3A4* and *CYP3A5* poly- morphisms in patients with DILI, and that
570 further studies are required to rule out an association of polymorphisms in these genes and

571 DILI risk, especially in patients with DILI related to drugs that are CYP3A4 or CYP3A5
572 substrates.

573 **ARYLAMINE N-ACETYLTRANSFERASE 2 (NAT2)**

574 Human arylamine N-acetyltransferase 2 (NAT2, EC 2.3.1.5) is responsible for the
575 acetylation of numerous xenobiotics and arylamine- or hydrazine-containing drugs [155].
576 NAT2 is involved in the metabolism of antibiotics, such as isoniazid and sulfamethoxazole,
577 and other drugs such as procainamide, hydralazine, dapsone, phenelzine, acebutolol or
578 caffeine. Genetic polymorphisms of *NAT2* are responsible for interindividual variation in
579 the metabolism of these drugs. The determination of NAT2 genotype or phenotype has
580 been proposed to predict adverse reactions in patients with tubercu- losis receiving
581 isoniazid and prior to the concomitant administration of drug combinations such as
582 procainamide-phenytoin or doxycycline-rifampin [156, 157]. The NAT2 enzyme is encoded
583 by a gene located in human chromosome 8 [158]. To date 52 alleles have been registered
584 at [http://www.louisville.edu/medschool/ pharmacology/NAT.html](http://www.louisville.edu/medschool/pharmacology/NAT.html) [159]. Most variant
585 alleles contain combinations of seven common SNPs within the coding region of the gene,
586 designated as G191A (rs1801279), C282T (rs1041983, rs59855457), T341C (rs1801280,
587 rs56935242), C481T (rs1799929, rs60310310), G590A (rs1799930, rs60190029), A803G
588 (rs1208, rs56599719, rs58999469) and G857A (rs1799931, rs58803786). Among these,
589 polymorphisms at positions 191, 341, 590 and 857 are the most studied and characterize
590 the major *NAT2* defect allele clusters (*NAT2**5, *NAT2**6, *NAT2**7 and *NAT2**14
591 respectively) [160]. NAT2 polymorphisms are responsible for variation in the acetyla- tion
592 phenotype. Individuals can be classified as slow or fast acetyla- tors on the basis of their
593 genotype, although some authors consider a third intermediate acetylator status. To infer
594 phenotypes from genotyping data, carriers of zero, one or two functional alleles are defined
595 as slow, intermediate and fast acetylators respectively, after reconstruction of *NAT2*
596 haplotypes from crude genotyping data, which is a complex process, although methods to
597 clear ambiguous diplotypes are available [161]. Interethnic and intraethnic variability of
598 *NAT2* polymorphisms has recently been analyzed [162].

599 Recently, the role of N- acetyltransferases in DILI has been reviewed [163]. Isoniazid
600 is the main drug responsible for hepatotoxicity induced by antituberculosis treatment. It is
601 metabolized to acetylisoniazid by hepatic N-acetyltransferase [164]. In turn, ace-
602 tylisoniazid is hydrolyzed to acetylhydrazine, which is oxidized by CYP2E1 to form
603 hepatotoxic intermediates [165]. However, the disposal of acetylhydrazine also depends on
604 further acetylation by NAT to form a non-toxic metabolite, diacetylhydrazine [166, 167].
605 A study of Huang *et al.* [168] shows that slow acetylators had a higher incidence of

606 antituberculosis drug-induced hepatitis and that once they had hepatotoxicity, they were
607 prone to develop more severe hepatic injury than rapid acetylators. Additionally patients
608 with homozygous genotype CYP2E1 c1/c1 have increased risk of hepatotoxicity (OR
609 2.52). Combined with the acetylation status, the risk of hepatotoxicity increased from 3.94
610 for patients with CYP2E1 c1/c1 and rapid acetylator status to 7.43 for patients with
611 CYP2E1 c1/c1 with slow acetylator status [133]. Cho *et al.* reported that slow acetylation
612 is related to antituberculosis drug-induced hepatotoxicity [135]. Other independent studies
613 supported the hypothesis of slow acetylation as a risk factor for drug-induced hepa-
614 totoxicity [136, 169-172]. However, negative findings have also been reported [134, 173].
615 Overall findings indicate a positive though modest association of the *NAT2* slow
616 acetylation genotype with DILI caused by antituberculosis drugs. A common limitation to
617 most of the studies performed so far is the small sample size and that the genotyping
618 procedures and the assignment of variant alleles after haplotype reconstruction differ across
619 the different studies. This is a major source of heterogeneity. Another source of hetero-
620 geneity is due to the interethnic differences in the frequency for variant *NAT2* alleles, which
621 are rare in Oriental individuals, and therefore studies in Oriental subjects require a high
622 sample size to have adequate statistical power. For further studies it would be advisable to
623 analyze at least the four SNPs that define the major *NAT2* clusters, to include a sample size
624 adequate to the ethnic origin of participants and to use a well-established method to assign
625 phenotypes.

626 **GLUTATHIONE TRANSFERASES (GSTS)**

627 The GSTs are a superfamily of phase II enzymes known to play important roles in
628 protecting against endogenous oxidative stress, as well as in the detoxification of
629 exogenous potential toxins including carcinogens, toxic chemicals and drugs. GSTs
630 catalyze nucleophilic attacks of glutathione on electrophilic substrates, thereby decreas-
631 ing the reactivity of potential toxicants with cellular molecules [174]. Mammalian
632 cytosolic GSTs are divided into eight classes: alpha, kappa, mu, omega, pi, sigma, theta
633 and zeta [175]. Genetic polymorphisms of these enzymes have been demonstrated and cor-
634 related to the susceptibility of many diseases including cancer and alcoholic liver disease
635 [176-180]. However, little is known about the genetic polymorphism of these
636 detoxification enzymes and DILI. To date, studies carried out on DILI patients have been
637 limited to gene deletion of *GSTT1* (pi class) and *GSTM1* (mu class) genes, designed as
638 null alleles [173, 181-185].

639 It has been shown that the risk of anti-TB drugs hepatotoxicity is increased in Indian
640 patients with *GSTM1* homozygous deletion [173]. Huang *et al.* [182] also reported that

641 patients with *GSTM1* homozygous deletion had a high risk of anti-tuberculosis drug-
642 induced hepatotoxicity in a Taiwanese population. However, no significant increase in risk
643 was observed in patients with *GSTT1* homozygous deletion. Recently, our group reported
644 that carriers of double *GSTT1-GSTM1* null genotypes had an increased risk of developing
645 DILI compared with noncarriers Caucasians patients with DILI [181]. Overall findings
646 [173, 181-186] consistently indicate that slow glutathione-S transferase activity is
647 associated with hepatotoxicity, and that this association is a general mechanism
648 independent of the causative drug and of the mechanism leading to either hepatocellular
649 or cholestatic mixed type of injury [181]. However, several issues remain to be analyzed
650 regarding GST enzymes. First, single nucleotide polymorphisms have been reported for
651 the *GSTT1* and *GSTM1* genes [187]. These single nucleotide polymorphisms are relevant
652 for Africans and Hispanic individuals
653 [188] and have not been analyzed with regard to DILI risk. There- fore studies involving
654 these subjects should explore the influence on these polymorphisms besides gene
655 deletion. Even more important is the absence of extensive studies on genetic variations
656 for other GST enzymes besides *GSTM1* and *GSTT1*. Because many GST enzymes are
657 expressed in the liver, it is likely that variations in the genes coding for the rest of GST
658 enzymes will influence DILI risk. These studies are in progress.

659 **UDP-GLUCURONOSYLTRANSFERASES (UGTS)**

660 UGTs are a superfamily of enzymes codified by UGT genes classified into UGT 1A, 2A
661 and 2B subfamilies which catalyse reactions of conjugation with glucuronic acid. These
662 enzymes are responsible for the elimination of drugs and another xenobiotics and
663 endogenous compounds like bile acids, bilirubin hydroxyl-steroids and thyroid hormones
664 [189].

665 The toxicity of some drugs like irinotecan and lamotrigine has been related to impaired
666 UGT1A activity because patients with mutant *UGT1A1* alleles are over-represented
667 amongst those experiencing severe toxicity [190-192]. Inhibition of UGT activity is re-
668 lated to simvastatin-ezetimibe induced liver failure [193]. The mechanism underlying this
669 relationship seems to be due to ezetimibe inhibition of UGT enzymes, which causes
670 increased simvastatin exposure. In a study carried out with human liver microsomes and
671 recombinant UGTs, Yoshigae *et al.* suggested that the liver failure induced by troglitazone
672 is not related with the *UGT1A1* polymorphism, because other UGT1 and UGT2 enzymes
673 may be responsible for troglitazone glucuronidation [194]. Because UGT enzymes overlap
674 in substrate specificity, it is likely that the presence of variant alleles in one of the UGT
675 genes would be insufficient to explain the variability in enzyme activity *in vivo*, that is, the

676 analysis of association of UGT genetic polymorphisms and hepatotoxicity can be extremely
677 complex if the drug is substrate of several UGTs enzymes.

678 UGT2B7 has been implicated in the diclofenac- induced hepatotoxicity [111].
679 Diclofenac metabolism is carried out by diverse hydroxylation pathways and by acyl
680 glucuronidation. About 50% of diclofenac is eliminated as 4-hydroxydiclofenac, a
681 CYP2C9 product. The other metabolite in humans, 5-hydroxydiclofenac, seems to be the
682 product of several enzymes, including CYP2C8, CYP2C18, CYP2C19 and CYP2B6 [110,
683 173]. A role for CYP3A4 in diclofenac metabolism has also been claimed [110, 173],
684 although this issue has not been investigated in sufficient detail. The rest of the drug is
685 metabolized by uridine 5'-diphosphoglucuronosyl transferases UGT2B4 and UGT2B7
686 [195]. Diclofenac glucuronide is further metabolized by CYP2C8 [70, 110, 196]. Daly *et*
687 *al.* [111] found that the variant *UGT2B7*2* allele is more common in patients with
688 hepatotoxicity caused by diclofenac, compared with healthy controls and with patients
689 receiving diclofenac without developing hepatotoxicity. Defective glucuronidation activity
690 as a result of *UGT1A9* mutated alleles, have been claimed to be the cause of two cases of
691 COMT inhibitor –induced hepatic dysfunction [197].

692 **FUTURE PERSPECTIVES**

693 As mentioned above, development of DILI is a complex, multi- step process in which
694 variability in drug metabolism may play a role, but pharmacogenomic studies are still
695 insufficient to confirm or to discount a prominent association of genetic variability in drug
696 metabolism and DILI risk. Variability in drug metabolism or bioactivation is compatible
697 with all proposed mechanisms for DILI. In addition, alterations in Phase II metabolizing
698 enzymes such as GSTs may contribute to the risk by means of general mechanisms
699 independent of the causative drug. Several major issues related to genetic polymorphisms
700 in drug metabolism remain insufficiently studied, for instance, the role of CYP2C8,
701 CYP3A5, and other polymorphisms. Further studies with a sufficient sample size and an
702 exhaustive study of all genetic polymorphisms relevant to the drug responsible for DILI
703 are required. These studies should combine genotyping for drug-metabolising enzymes
704 with the analysis of polymorphisms related to general detoxication mechanisms, such as
705 those of GST enzymes (especially other enzymes besides *GSTM1* and *GSTT1*), superoxide
706 dismutase or glutathione peroxidase enzymes. Regarding the mechanisms that propose
707 immune-mediated hepatotoxicity or the danger-signal hypothesis, it would be worth
708 exploring polymorphisms related to histamine that have clinical implications [198-206],
709 and how these histamine-related polymorphisms may modulate the effect of
710 polymorphisms of drug- metabolising enzymes in the risk to develop immune-mediated

711 DILI.

712 Besides licensed drugs, herbal and natural supplements are recognized as causing
713 hepatotoxicity with increasing frequency as patients turn more and more to alternative
714 medicine [4]. Some com- pounds in herbal and dietary constituents are bioactivated through
715 drug-metabolising enzymes [207], and in addition, these compounds are able to modulate
716 drug-metabolizing enzymes [149, 208] and hence may modify the risk for DILI even if the
717 herbal compounds are not directly hepatotoxic. This highlights the necessity to investigate
718 in detail the hepatotoxic potential of herbal com- pounds, and the mechanisms underlying
719 hepatotoxicity related to herbal and dietary products. This is a promising field that will
720 evolve in the coming years and hopefully will provide useful in- formation to obtain a more
721 comprehensive view of the mechanisms related to DILI and to develop pharmacogenetic
722 analyses to identify at-risk patients in clinical practice.

723

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